Package: statuser (via r-universe)

October 31, 2026

Type Package

Title Statistical Tools Designed for End Users

Version 0.4.1

Author Uri Simonsohn [aut, cre]

Maintainer Uri Simonsohn <urisohn@gmail.com>

Description The statistical tools in this package do one of four things: 1) Enhance basic statistical functions with more flexible inputs, smarter defaults, and richer, clearer, and ready-to-use output (e.g., t.test2()) 2) Produce publication-ready commonly needed figures with one line of code (e.g., plot_cdf()) 3) Implement novel analytical tools developed by the authors (e.g., twolines()) 4) Deliver niche functions of high value to the authors that are not easily available elsewhere (e.g., clear(), convert_to_sql(), resize_images()).

License GPL-3

URL https://github.com/urisohn/statuser

BugReports https://github.com/urisohn/statuser/issues

Depends R (>= 3.5.0)

Encoding UTF-8

LazyData true

Imports mgcv, marginaleffects, rsvg, magick, sandwich, lmtest, lmerTest, digest, beeswarm, utils

Suggests testthat (>= 3.0.0), crayon, quantreg, estimatr, broom, modelsummary, knitr, rmarkdown

VignetteBuilder knitr

Config/testthat/edition 3

Config/roxygen2/version 8.1.0

NeedsCompilation no

Repository https://cran.r-universe.dev

Date/Publication 2026-10-31 12:50:13 UTC

RemoteUrl https://github.com/cran/statuser

RemoteRef HEAD

RemoteSha 56173248d48627db9f965ba565e948a9a87e0f39

Contents

clear . . . 3
clear_stimulus_cache . . . 4
convert_to_sql . . . 4
desc_var . . . 5
format_pvalue . . . 7
interprobe . . . 8
interprobe_sim . . . 12
list2 . . . 13
lm2 . . . 14
lm2_notes . . . 17
message2 . . . 18
plot_cdf . . . 19
plot_density . . . 21
plot_freq . . . 23
plot_gam . . . 25
plot_means . . . 27
predict.lm2 . . . 29
print.desc_var . . . 30
print.lm2 . . . 30
print.t.test2 . . . 31
print.table2 . . . 32
resize_images . . . 32
round2 . . . 33
scatter.gam . . . 34
stimulus.beeswarm . . . 36
stimulus.plot . . . 38
summary.lm2 . . . 40
t.test2 . . . 40
table2 . . . 42
tacos_data . . . 44
text2 . . . 44
twolines . . . 46
var_labels . . . 48

Index **49**

clear *Clear Plot, Global Environment, and Console*

Description

Clears plot, global environment, and console. On first use the user is prompted to authorize clearing the environment to comply with CRAN rules.

Usage

```
clear()
```

Details

This function performs three cleanup operations:

- **Plot**: Closes all open graphics devices (except the null device)
- **Global environment**: Removes all objects from the global environment
- **Console**: Clears the console screen (only in interactive sessions)

`clear()` will not modify the global environment unless you have previously typed "yes" when prompted. If you do not type "yes", you are asked again next time; only "yes" is remembered for future sessions.

Warning: This function deletes all objects in the global environment. Save anything that you wish to keep before running.

Value

Invisibly returns NULL. Prints a colored confirmation message.

Examples

```
# Interactive use: clear workspace, console, and plots
# First run may prompt; once you type "yes", your preference is saved.
clear()
```

clear_stimulus_cache *Clear cache used by stimulus.plot() to reuse resampling results from identical previous calls.*

Description

When ‘stimulus.plot()‘ with ‘plot.type = ’effects’‘ is run, resamples are saved in package state. If an identical call is run again (same data, dv, condition, number of simulations, etc.), stored results are loaded instead of re-calculated. Force recalculation by clearing the cache with this function.

Usage

```
clear_stimulus_cache()
```

convert_to_sql *Convert CSV file to SQL INSERT statements*

Description

Reads a CSV file and generates SQL statements to insert all rows. Optionally can also generate a CREATE TABLE statement. The function automatically infers column types (REAL for numeric, DATE for date strings matching YYYY-MM-DD format, TEXT otherwise).

Usage

```
convert_to_sql(input, output, create_table = FALSE)
```

Arguments

input	Character string. Path to the input CSV file.
output	Character string. Path to the output SQL file where the statements will be written.
create_table	Logical. If TRUE, includes a CREATE TABLE statement before the INSERT statements. Default is FALSE.

Details

The function performs the following steps:

1. Reads the CSV file using `read.csv()` with `stringsAsFactors = FALSE`
2. Infers SQL column types:
   - Numeric columns become `REAL`
   - Date columns (matching YYYY-MM-DD format) become `DATE`
   - All other columns become `TEXT`

3. If `create_table = TRUE`, generates a `CREATE TABLE` statement using the base filename (without extension) as the table name
4. Generates `INSERT INTO` statements for each row
5. Writes all SQL statements to the output file

Single quotes in text values are escaped by doubling them (SQL standard). Numeric values are inserted without quotes, while text and date values are wrapped in single quotes.

Value

Invisibly returns NULL. The function writes SQL statements to the specified output file.

Examples

```
# Convert a CSV file to SQL (INSERT statements only)
tmp_csv <- tempfile(fileext = ".csv")
tmp_sql <- tempfile(fileext = ".sql")
write.csv(
  data.frame(id = 1:2, value = c("a", "b"), date = c("2024-01-01", "2024-02-02")),
  tmp_csv,
  row.names = FALSE
)
convert_to_sql(tmp_csv, tmp_sql)

# Convert a CSV file to SQL with CREATE TABLE statement
convert_to_sql(tmp_csv, tmp_sql, create_table = TRUE)
```

---

desc_var *Describe a variable, optionally by groups*

Description

Returns a dataframe with one row per group.

Usage

```
desc_var(y, group = NULL, data = NULL, digits = 3)
```

Arguments

`y`	A numeric vector of values, a column name (character string or unquoted) if `data` is provided, or a formula of the form `y ~ x` or `y ~ x1 + x2` (for multiple grouping variables).
`group`	Optional grouping variable, if not provided computed for the full data. Ignored if `y` is a formula.
`data`	Optional data frame containing the variable(s).
`digits`	Number of decimal places to round to. Default is 3.

Details

The dependent variable (‘y‘) must be numeric. If you pass an existing non-numeric variable (e.g., character, factor), ‘desc_var()‘ will stop with a clear error indicating that the variable must be numeric.

Value

A data frame with one row per group (or one row if no group is specified) containing:

- `group`: Group identifier
- `mean`: Mean
- `sd`: Standard deviation
- `se`: Standard error
- `median`: Median
- `min`: Minimum
- `max`: Maximum
- `mode`: Most frequent value
- `freq_mode`: Frequency of mode
- `mode2`: 2nd most frequent value
- `freq_mode2`: Frequency of 2nd mode
- `n.total`: Number of observations
- `n.missing`: Number of observations with missing (NA) values
- `n.unique`: Number of unique values

Columns may also carry a `"label"` attribute, which provides a short human-readable description of each column.

Examples

```
# With grouping
df <- data.frame(y = rnorm(100), group = rep(c("A", "B"), 50))
desc_var(y, group, data = df)

# Without grouping (full dataset)
desc_var(y, data = df)

# Direct vectors
y <- rnorm(100)
group <- rep(c("A", "B"), 50)
desc_var(y, group)

# With custom decimal places
desc_var(y, group, data = df, digits = 2)

# Using formula syntax: y ~ x
desc_var(y ~ group, data = df)
```

```
# Using formula syntax with multiple grouping variables: y ~ x1 + x2
df2 <- data.frame(y = rnorm(200), x1 = rep(c("A", "B"), 100), x2 = rep(c("X", "Y"), each = 100))
desc_var(y ~ x1 + x2, data = df2)
```

`format_pvalue` *Format P-Values for Display*

Description

Formats p-values for clean display in figures and tables. e.g., p = .0231, p<.0001

Usage

```
format_pvalue(p, digits = 4, include_p = FALSE)
```

Arguments

`p`	A numeric vector of p-values to format.
`digits`	Number of decimal places to round to. Default is 4.
`include_p`	Logical. If TRUE, includes "p" prefix before the formatted value (e.g., "p = .05"). Default is FALSE.

Value

A character vector of formatted p-values.

Examples

```
# Basic usage
format_pvalue(0.05)
format_pvalue(0.0001)

# More rounding
format_pvalue(0.0001,digits=2)

# Vector input
format_pvalue(c(0.05, 0.001, 0.00001, 0.99))

# With p prefix
format_pvalue(0.05, include_p = TRUE)
```

`interprobe` *Test and probe possibly nonlinear interactions*

Description

Tests and probes 2-way regression interactions (i.e., between two predictors, x & z) on their association with the dependent variable, y. When the argument `model` is not specified, interprobe() estimates a Generalized Additive Model (`mgcv::gam`) and thus does not impose linearity on any of the associations (the optional argument `model` can be used to apply interprobe to other models, e.g., linear regression). In terms of testing the interaction, when `model` is not specified, the interaction p-value comes from the null that the association between x and y is independent of z. For legacy purposes, the p-value for the linear model's x*z interaction is also reported. In case of substantive differences between the GAM and linear p-values for the interaction, the plots `interprobe` generates (see below) are likely to help users identify the origin of the disagreement (e.g., a blatantly nonlinear association between x & y could produce a spurious (or eliminate a real) interaction between x & z). In terms of probing the interaction (describing the estimated association between the focal predictor, x, and the dependent variable, y, for a given set of x, z values), `interprobe` reports two traditional analyses: simple slopes and the Johnson-Neyman procedure. Simple slopes map the predicted y for combinations of x & z. Simple slopes are also known as "spotlight analysis". Johnson-Neyman analysis, in turn, maps the estimated marginal effect of x on y, for sets of x & z values. Both simple slopes and Johnson-Neyman keep all other predictors constant, which is necessary to conduct the analysis ceteris paribus (see https://datacolada.org/123).

Usage

```
interprobe(
  x = NULL, z = NULL, y = NULL, model = NULL, data = NULL,
 moderator.on.x.axis = TRUE, k = NULL, spotlights = NULL, spotlight.labels = NULL,
  histogram = TRUE, max.unique = 11, n.bin.continuous = 10, n.max = 50,
  xlab = "", ylab1 = "", ylab2 = "", cols = c("red4", "dodgerblue", "green4"),
  main1 = "GAM Simple Slopes", main2 = "GAM Johnson-Neyman",
  legend.round = c(1, 4), draw = "both", save.as = NULL,
  xlim = NULL, ylim1 = NULL, ylim2 = NULL,
  x.ticks = NULL, y1.ticks = NULL, y2.ticks = NULL,
  legend.simple.slopes = NULL, legend.jn = NULL, quiet = FALSE, probe.bins = 100
)
```

Arguments

`x`	Focal predictor. A name (bare or quoted) when `data` or `model` is provided, or a numeric/factor vector when probing from vectors.
`z`	Moderator. Same options as `x`.
`y`	Dependent variable. Same options as `x`. Not required (and ignored) when `model` is supplied.
`model`	Model to probe. Default `NULL` fits a GAM of `y` on `x` and `z` (see Model estimated). Use `model = "linear"` or `model = "lm"` for an internal linear interaction via

lm2(). Pass a fitted object to probe a model of your choice (the usual path for covariates).

data — Optional data frame containing x, z, and y. Cannot be combined with a fitted model.

moderator.on.x.axis — Logical. If TRUE (default), moderator z is on the x-axis. Ignored (forced to TRUE) when x has 3 or fewer unique values.

k — Integer smoothness parameter passed to mgcv::gam() when estimating with the default GAM engine. Default NULL omits k (mgcv default), except the binary-x testing GAM which uses k = 10.

spotlights — Optional numeric vector of length 3. Values at which curves are computed. Default NULL uses the 15th/50th/85th percentiles (see Spotlights).

spotlight.labels — Optional character vector of length 3. Legend labels. Default NULL builds labels from spotlights and legend.round.

histogram — Logical. If TRUE (default), show a frequency panel under each curve plot (common support).

max.unique — Integer. Threshold for continuous vs discrete treatment of a variable (default 11). Separately, x with 2-3 unique values is always categorical (see How x and z are classified).

n.bin.continuous — Integer. Number of bins in the histogram when binning continuous values (default 10).

n.max — Integer. Sample size at which line darkness/width saturates (default 50).

xlab — Character. X-axis label. Default "" uses the variable name.

ylab1 — Character. Y-axis label for simple slopes. Default "" becomes the name of y.

ylab2 — Character. Y-axis label for the JN panel. Default "" becomes "Marginal effect of <x>".

cols — Character vector of length 2 or 3. Colors for the curves. Length 2 is allowed when the focal predictor has exactly two levels; otherwise length 3 is required (three spotlights or up to three categorical levels). Default c("red4", "dodgerblue", "green4").

main1 — Character. Title for simple slopes. Default "GAM Simple Slopes"; rewritten to drop "GAM" when a non-GAM fitted model is supplied.

main2 — Character. Title for JN. Default "GAM Johnson-Neyman"; rewritten like main1 for non-GAM fitted models.

legend.round — Integer vector length 2. Min/max decimals in legend labels (default c(1, 4)).

draw — Which plots to draw: "both" (default), "simple slopes" (or legacy "simple.slopes"), "jn", or "none" (no plot).

save.as — Optional file path to save the plot (.png or .svg).

xlim — Numeric vector length 2. X-axis limits. Default NULL uses the data range.

ylim1 — Numeric vector length 2. Y-axis limits for simple slopes. Default NULL.

ylim2 — Numeric vector length 2. Y-axis limits for JN. Default NULL.

x.ticks	Optional custom x-axis ticks. Default NULL.
y1.ticks	Optional custom y-axis ticks for panel 1. Default NULL.
y2.ticks	Optional custom y-axis ticks for panel 2. Default NULL.
legend.simple.slopes	Optional legend title for simple slopes. Default NULL.
legend.jn	Optional legend title for JN. Default NULL.
quiet	Logical. If TRUE, suppresses the probing message, interaction tests, and JN region printout (default FALSE).
probe.bins	Integer. Grid resolution for continuous probing (default 100; larger = smoother/slower).

Value

Invisibly returns a list with:

- simple.slopes: data.frame of predicted values (y.hat) and confidence limits, plus the evaluation grid for x and z
- johnson.neyman: data.frame of marginal effects (marginal.effect) and confidence limits on the same grid
- frequencies: data.frame with bin frequencies used for shading/histogram (bin, f1–f3; after a plot is drawn, also bin_from / bin_to)
- gam_results: the model fitted inside interprobe() when none was supplied. Despite the name, this is an lm2 fit when model = "linear" / "lm"
- gam_results_testing: when a GAM is estimated internally and x has exactly 2 unique values, a separate GAM used for interaction testing (with a ti() term and numeric coding of x)
- lm2_results: when a GAM is estimated internally, the corresponding linear fit lm2(y ~ x * z) (or NULL if estimatr is not installed). Not set on the model = "linear" path

Input options

Call interprobe() in one of three ways:

1. data plus names x, z, and y
2. vectors x, z, and y (no data)
3. a fitted model plus names x and z (y is taken from the model)

You cannot pass both data and model. If both model and y are supplied, y is ignored.

Model estimated

By default interprobe() fits a GAM with mgcv::gam(..., method = "REML"). GAM probing avoids the linear-interaction-effect assumption of the classical y ~ x * z model: the conditional effect of x can bend freely with z (and vice versa). The frequency panel under each plot shows where the data support the curves (common support across the moderator range).

Default formulas:

- If x has 3 or fewer unique values: y ~ s(z, by = x) + x (with x as a factor)

- If x has 4 or more unique values: `y ~ s(z) + s(x) + ti(x, z)`

Set `model = "linear"` (or `"lm"`) to fit `lm2(y ~ x * z)` instead. Pass a fitted model (e.g. `lm()`, `lm2()`, `glm()`, `mgcv::gam()`) to include covariates; other predictors are held at their means on the prediction grid. Any model supported by `marginaleffects` can be probed.

How x and z are classified

Classification uses the number of unique observed values:

- x with 2-3 unique values: **categorical** (one curve per level; spotlights unused)
- x with 4 to `max.unique` unique values: **discrete** (still uses three spotlights)
- x with more than `max.unique` unique values: **continuous**
- z at or below `max.unique`: evaluated at observed values
- z above `max.unique`: evaluated on a grid of `probe.bins` points from min to max

If x has 3 or fewer unique values, `moderator.on.x.axis = FALSE` is ignored and the moderator is forced onto the x-axis.

Spotlights

When `spotlights` is `NULL` and x is not categorical, curves are drawn at the 15th, 50th, and 85th percentiles (`stats::quantile(..., type = 3)`) of x if the moderator is on the x-axis, otherwise of z. Default legend labels are built from those percentiles and `legend.round`.

Console output

Unless `quiet = TRUE`, prints the pair being probed. When a GAM is estimated internally, also prints an HC3 linear interaction test and a GAM interaction F test. Johnson-Neyman regions of significance are printed only for a categorical focal predictor when the JN panel is drawn.

References

Simonsohn, U. (2024). Interacting with curves: How to validly test and probe interactions in the real (nonlinear) world. *Advances in Methods and Practices in Psychological Science*, 7(1), Article 25152459231207787. doi:10.1177/25152459231207787

Montealegre, D., & Simonsohn, U. (2026). *Johnson Neyman 2.0* (working paper).

Run `citation("statuser")` for ready-to-use citation entries.

See Also

`vignette("interprobe", package = "statuser")`, `lm2`, `gam`, `tacos_data`

Examples

```
# This is a stylized dataset inspired by Montealegre & Simonsohn (2026).
# It depicts how salsa spiciness relates to taco enjoyment,
# moderated by respondents' tolerance for spicy food.
data(tacos_data)

# Case A: experiment with Mild vs Hot salsa (binary focal predictor)
tacos_bin <- tacos_data[!is.na(tacos_data$spicy_bin), ]
interprobe(x = spicy_bin, z = tolerance, y = enjoyment, data = tacos_bin)

# Case B: three salsa levels (Mild / Medium / Hot)
interprobe(x = salsa, z = tolerance, y = enjoyment, data = tacos_data)

# Case C: continuous spiciness rating (1-7) by continuous tolerance
interprobe(x = spicy, z = tolerance, y = enjoyment, data = tacos_data)

# Same as Case A, but force a classical linear interaction
interprobe(
  x = spicy_bin, z = tolerance, y = enjoyment,
  data = tacos_bin, model = "linear"
)
# Probe a user-fitted model (here with age as a covariate)
lm1 <- lm(enjoyment ~ spicy_bin * tolerance + age, data = tacos_bin)
interprobe(model = lm1, x = spicy_bin, z = tolerance)
```

`interprobe_sim` *Simulated nonlinear interaction for interprobe demos*

Description

A small simulated dataset with a binary treatment, a continuous moderator, and an outcome whose treatment effect rises nonlinearly with the moderator and then flattens: $2(1 - e^{-mod/3})$. Useful for examples and for checking that GAM probing recovers a curved interaction that a linear `y ~ treat * mod` model would misshape.

Format

A data frame with 400 rows and 3 variables:

treat Factor with levels `Control`, `Treated`

mod Numeric moderator on $[0, 10]$

y Numeric outcome

Source

Simulated in '`data-raw/interprobe_sim.R`' with seed 20241030.

Examples

```
data(interprobe_sim)
head(interprobe_sim)

interprobe(x = treat, z = mod, y = y, data = interprobe_sim, quiet = TRUE)
```

`list2` *Enhanced alternative to list()*

Description

List with objects that are automatically named.

Usage

```
list2(...)
```

Arguments

`...`	Objects to include in the list. Objects are automatically named based on their variable names unless explicit names are provided.

Details

list2(x , y) is equivalent to list(x = x , y = y)

list2(x , y2 = y) is equivalent to list(x = x , y2 = y)

Based on: https://stackoverflow.com/questions/16951080/can-lists-be-created-that-name-themselves-bas

Value

A named list. Each element is named after the variable passed to the function (or the explicit name if provided). The structure is identical to a standard R list created with `list`.

Examples

```
x <- 1:5
y <- letters[1:3]
z <- matrix(1:4, nrow = 2)

# Create named list from objects
my_list <- list2(x, y, z)
names(my_list)  # "x" "y" "z"

# Works with explicit names too
my_list2 <- list2(a = x, b = y)
names(my_list2)  # "a" "b"
```

lm2 *Enhanced alternative to lm()*

Description

Runs a linear regression with better defaults (robust SE), and richer & better formatted output than lm. For robust and clustered errors it relies on lm_robust. The output reports classical and robust errors, number of missing observations per variable, an effect size column (standardized regression coefficient), and a red.flag column per variable flagging the need to conduct specific diagnostics. It relies by default on HC3 for standard errors; lm_robust relies on HC2 (and Stata's 'reg y x, robust' on HC1), which can have inflated false-positive rates in smaller samples (Long & Ervin, 2000).

Usage

```
lm2(
  formula,
  data = NULL,
  se_type = "HC3",
  notes = TRUE,
  clusters = NULL,
  fixed_effects = NULL,
  round = NULL,
  ...
)
```

Arguments

formula	A formula as in lm (e.g., y ~ x1 + x2).
data	An optional data frame containing the variables in the model.
se_type	The type of standard error to use. Default is "HC3". Without clusters: "HC0", "HC1", "HC2", or "HC3". When clusters is specified, se_type is automatically set to "CR2".
notes	deprecated. Ignored. Explanatory notes are always built when the model is printed; use lm2_notes to view them.
clusters	An optional variable indicating clusters for cluster-robust standard errors. When specified, se_type is automatically set to "CR2" (bias-reduced cluster-robust estimator). Passed to lm_robust.
fixed_effects	An optional right-sided formula containing the fixed effects to be projected out (absorbed) before estimation. Useful for models with many fixed effect groups (e.g., ~ firm_id or ~ firm_id + year). Passed to lm_robust.
round	Optional integer controlling printed numeric formatting. NULL (default): magnitude-based decimals (1–3 places) with zero-chop. -1: R default format() (respects options("digits")), with leading-dot style for |x| < 1. Non-negative k: fixed k decimal places.
...	Additional arguments passed to lm_robust (and, where applicable, the same style of arguments as lm).

Details

Robust standard errors and clustered standard errors are computed using `lm_robust`; see the documentation of that function for details. With clusters, `lm_robust` uses CR2 by default. The output shows both standard errors; when clustering is used it reports all three. The red.flag column is based on the difference between robust and classical standard errors.

The `red.flag` column provides diagnostic warnings:

- `!`, `!!`, `!!!`: Robust and classical standard errors differ by more than 25%, 50%, or 100%, respectively. Large differences may suggest model misspecification or outliers (but they may also be benign). When encountering a red flag, authors should plot the distributions to look for outliers or skewed data, and use `scatter.gam` to look for possible nonlinearities in the relevant variables. King & Roberts (2015) propose a higher cutoff, at 100%, and a bootstrapped significance test; `statuser` does not follow either recommendation. The former seems too liberal, the latter too time consuming to include in every regression, plus the focus here is on individual variables rather than joint tests.
- `X`: For interaction terms, the component variables are correlated ($|r| > 0.3$ or $p < .05$), which means the interaction term is likely to be biased. See Simonsohn (2024) "Interacting with curves" doi:10.1177/25152459231207787.

Value

An object of class `c("lm2", "lm_robust", "lm")`. This inherits from `lm_robust` and can be used with packages like `marginaleffects`. The object contains all components of an `lm_robust` object plus additional attributes:

statuser_table A data frame with columns: `term`, `estimate`, `SE.robust`, `SE.classical`, `t`, `df`, `p.value`, `B` (standardized coefficient), and optionally `SE.cluster` when clustered standard errors are used.

classical_fit The underlying `lm` object with classical standard errors.

na_counts Integer vector of missing value counts per variable.

n_missing Total number of observations excluded due to missing values.

has_clusters Logical indicating whether clustered standard errors were used.

When printed, displays a formatted regression table with robust and classical standard errors, effect sizes, and diagnostic red flags.

References

King, G., & Roberts, M. E. (2015). How robust standard errors expose methodological problems they do not fix, and what to do about it. *Political Analysis*, 23(2), 159-179.

Long, J. S., & Ervin, L. H. (2000). Using heteroscedasticity consistent standard errors in the linear regression model. *The American Statistician*, 54(3), 217-224.

Simonsohn, U. (2024). Interacting with curves: How to validly test and probe interactions in the real (nonlinear) world. *Advances in Methods and Practices in Psychological Science*, 7(1), 1-22.

See Also

`lm_robust`, `scatter.gam`, `lm2_notes`

Examples

```
# Basic usage with data argument
lm2(mpg ~ wt + hp, data = mtcars)

# Without data argument (variables from environment)
y <- mtcars$mpg
x1 <- mtcars$wt
x2 <- mtcars$hp
lm2(y ~ x1 + x2)

# RED FLAG EXAMPLES

# Example 1: red flag catches a nonlinearity
# True model is quadratic: y = x^2
set.seed(123)
x <- runif(200, -3, 3)
y <- x^2 + rnorm(200, sd = 2)

# lm2() shows red flag due to misspecification
lm2(y ~ x)

# Follow up with scatter.gam() to diagnose it
scatter.gam(x, y)

# Example 2: red flag catches an outlier in y
# True model is y = x, but one observation has a very large y value
set.seed(123)
x <- sort(rnorm(200))
y <- round(x + rnorm(200, sd = 2), 1)
y[200] <- 100  # Outlier

# lm2() flags x
lm2(y ~ x)

# Look at distribution of y to spot the outlier
plot_freq(y)

# Example 3: red flag catches an outlier in one predictor
# True model is y = x1 + x2, but x2 has an extreme value
set.seed(123)
x1 <- round(rnorm(200),.1)
x2 <- round(rnorm(200),.1)
y <- x1 + x2 + rnorm(200, sd = 0.5)
x2[200] <- 50  # Outlier in x2

# lm2() flags x2 (but not x1)
lm2(y ~ x1 + x2)

# Look at distribution of x2 to spot the outlier
plot_freq(x2)

# CLUSTERED STANDARD ERRORS
```

```
# When observations are grouped (e.g., students within schools),
# use clusters to account for within-group correlation
set.seed(123)
n_clusters <- 20
n_per_cluster <- 15
cluster_id <- rep(1:n_clusters, each = n_per_cluster)
cluster_effect <- rnorm(n_clusters, sd = 2)[cluster_id]
x <- rnorm(n_clusters * n_per_cluster)
y <- 1 + 0.5 * x + cluster_effect + rnorm(n_clusters * n_per_cluster)
mydata <- data.frame(y = y, x = x, cluster_id = cluster_id)

# Clustered SE (CR2) - note the SE.cluster column in output
lm2(y ~ x, data = mydata, clusters = cluster_id)

# FIXED EFFECTS
# Use fixed_effects to absorb group-level variation (e.g., firm or year effects)
# This is useful for panel data or when you have many fixed effect levels
set.seed(456)
n_firms <- 30
n_years <- 5
firm_id <- rep(1:n_firms, each = n_years)
year <- rep(2018:2022, times = n_firms)
firm_effect <- rnorm(n_firms, sd = 3)[firm_id]
x <- rnorm(n_firms * n_years)
y <- 2 + 0.8 * x + firm_effect + rnorm(n_firms * n_years)
panel <- data.frame(y = y, x = x, firm_id = factor(firm_id), year = factor(year))

# Absorb firm fixed effects (coefficient on x is estimated, firm dummies are not shown)
lm2(y ~ x, data = panel, fixed_effects = ~ firm_id)

# Two-way fixed effects (firm and year)
lm2(y ~ x, data = panel, fixed_effects = ~ firm_id + year)
```

`lm2_notes` *Print explanatory notes from the most recent* `lm2` *print*

Description

After printing an `lm2` object, call `lm2_notes()` to display the full `Notes:` block (significance legend, column definitions, red-flag guidance, etc.).

Usage

```
lm2_notes()
```

Value

Invisibly returns the notes character string, or `NULL` if none stored.

See Also

lm2, print.lm2

`message2` *Enhanced alternative to message()*

Description

Add options to set color and to end execution of code (to be used as error message)

Usage

```
message2(..., col = "cyan", font = 1, stop = FALSE)
```

Arguments

`...`	Message content to be printed. Multiple arguments are pasted together.
`col`	text color. Default is "cyan".
`font`	Integer. 1 for plain text (default), 2 for bold text.
`stop`	Logical. If TRUE, stops execution (like `stop()`) but without printing "Error:".

Details

This function prints colored messages to the console. If ANSI color codes are supported by the terminal, the message will be colored. Otherwise, it will be printed as plain text. If `stop = TRUE`, execution will be halted after printing the message.

Value

No return value, called for side effects. Prints a colored message to the console. If `stop = TRUE`, execution is halted after printing the message.

Examples

```
message2("This is a plain cyan message", col = "cyan", font = 1)
message2("This is a bold cyan message", col = "cyan", font = 2)
message2("This is a bold red message", col = "red", font = 2)

cat("this will be shown")
try(message2("This stops execution", stop = TRUE), silent = TRUE)
cat("this will be shown after the try")
```

plot_cdf *Plot Empirical Cumulative Distribution Functions by Group*

Description

Plots empirical cumulative distribution functions (ECDFs) separately for each unique value of a grouping variable, with support for vectorized plotting parameters. If no grouping variable is provided, plots a single ECDF.

Usage

```
plot_cdf(
  formula,
  y2 = NULL,
  data = NULL,
  order = NULL,
  show.ks = TRUE,
  show.quantiles = TRUE,
  ...
)
```

Arguments

formula	Two possible uses (similar to `t.test()`): • Single Variable (possibly by subgroup): `plot_cdf(y)` or `plot_cdf(y~x)` • Contrast Two Variables: `plot_cdf(y1, y2)`
y2	optional second variable when contrasting two variables `plot_cdf(y1,y2)`
data	An optional data frame containing the variables in the formula. If `data` is not provided, variables are evaluated from the calling environment.
order	Controls the order in which groups appear in the plot and legend. Use `-1` to reverse the default order. Alternatively, provide a vector specifying the exact order (e.g., `c("B", "A", "C")`). If `NULL` (default), groups are ordered by their factor levels (if the grouping variable is a factor) or sorted alphabetically/numerically. Only applies when using grouped plots.
show.ks	Logical. If TRUE (default), shows Kolmogorov-Smirnov test results when there are exactly 2 groups. If FALSE, KS test results are not displayed.
show.quantiles	Logical. If TRUE (default), shows horizontal lines and results at 25th, 50th, and 75th percentiles when there are exactly 2 groups. If FALSE, quantile lines and results are not displayed.
...	Additional arguments passed to plotting functions. Can be single values (applied to all groups) or vectors (applied element-wise to each group). Common parameters include `col`, `lwd`, `lty`, `pch`, `type`, `xlim`, and `ylim`.

Value

Invisibly returns a list containing:

- `ecdfs`: A list of ECDF function objects, one per group. Each can be called as a function to compute cumulative probabilities (e.g., `result$ecdfs[[1]](5)` returns P(X <= 5) for group 1).
- `ks_test`: (Only when exactly 2 groups) The Kolmogorov-Smirnov test result comparing the two distributions. Access p-value with `result$ks_test$p.value`.
- `quantile_regression_25`: (Only when exactly 2 groups) Quantile regression model for the 25th percentile.
- `quantile_regression_50`: (Only when exactly 2 groups) Quantile regression model for the 50th percentile (median).
- `quantile_regression_75`: (Only when exactly 2 groups) Quantile regression model for the 75th percentile.
- `warnings`: Any warnings captured during execution (if any).

Examples

```
# Basic usage with single variable (no grouping)
y <- rnorm(100)
plot_cdf(y)

# Basic usage with formula syntax and grouping
group <- rep(c("A", "B", "C"), c(30, 40, 30))
plot_cdf(y ~ group)

# With custom colors (scalar - same for all)
plot_cdf(y ~ group, col = "blue")

# With custom colors (vector - different for each group)
plot_cdf(y ~ group, col = c("red", "green", "blue"))

# Multiple parameters
plot_cdf(y ~ group, col = c("red", "green", "blue"), lwd = c(1, 2, 3))

# With line type and point character
plot_cdf(y ~ group, col = c("red", "green", "blue"), lty = c(1, 2, 3), lwd = 2)

# Using data frame
df <- data.frame(value = rnorm(100), group = rep(c("A", "B"), 50))
plot_cdf(value ~ group, data = df)
plot_cdf(value ~ group, data = df, col = c("red", "blue"))

# Custom x-axis limits
plot_cdf(y ~ group, xlim = c(0, 10))

# Compare two vectors
y1 <- rnorm(50)
y2 <- rnorm(50, mean = 1)
plot_cdf(y1, y2)
```

```
# Formula syntax without data (variables evaluated from environment)
widgetness <- rnorm(100)
gender <- rep(c("M", "F"), 50)
plot_cdf(widgetness ~ gender)

# Using the returned object
df <- data.frame(value = c(rnorm(50, 0), rnorm(50, 1)), group = rep(c("A", "B"), each = 50))
result <- plot_cdf(value ~ group, data = df)

# Use ECDF to find P(X <= 0.5) for group A
result$ecdfs[[1]](0.5)

# Access KS test p-value
result$ks_test$p.value

# Summarize median quantile regression
summary(result$quantile_regression_50)
```

`plot_density` *Plot density of a variable, optionally by another variable*

Description

Plots the distribution of a variable by group, simply: `plot_density(y ~ x)`

Usage

```
plot_density(
  formula,
  y2 = NULL,
  data = NULL,
  order = NULL,
  show_means = TRUE,
  ...
)
```

Arguments

`formula`	Two possible uses (similar to `t.test()`): • Single Variable (possibly by subgroup): `plot_density(y)` or `plot_density(y~x)` • Contrast Two Variables: `plot_density(y1, y2)`
`y2`	optional second variable when contrasting two variables `plot_density(y1,y2)`
`data`	An optional data frame containing the variables in the formula.
`order`	Controls the order in which groups appear in the plot and legend. Use `-1` to reverse the default order. Alternatively, provide a vector specifying the exact order (e.g., `c("B", "A", "C")`). If `NULL` (default), groups are ordered by their factor levels (if the grouping variable is a factor) or sorted alphabetically/numerically. Only applies when using grouped plots.

show_means	Logical. If TRUE (default), shows points at means.
...	Additional arguments passed to plotting functions.

Details

Plot parameters like col, lwd, lty, and pch can be specified as:

- A single value: applied to all groups
- A vector: applied to groups in order of unique group values

Value

Invisibly returns a list with the following element:

densities A named list of density objects (class "density"), one for each group. Each density object contains x (evaluation points), y (density estimates), bw (bandwidth), and other components as returned by density. If no grouping variable is provided, the list contains a single element named "all".

The function is primarily called for its side effect of creating a plot.

Examples

```
# Basic usage with formula syntax (no grouping)
y <- rnorm(100)
plot_density(y)

# With grouping variable
group <- rep(c("A", "B", "C"), c(30, 40, 30))
plot_density(y ~ group)

# With custom colors (scalar - same for all)
plot_density(y ~ group, col = "blue")

# With custom colors (vector - different for each group)
plot_density(y ~ group, col = c("red", "green", "blue"))

# Multiple parameters
plot_density(y ~ group, col = c("red", "green", "blue"), lwd = c(1, 2, 3))

# With line type
plot_density(y ~ group, col = c("red", "green", "blue"), lty = c(1, 2, 3), lwd = 2)

# Using data frame
df <- data.frame(value = rnorm(100), group = rep(c("A", "B"), 50))
plot_density(value ~ group, data = df)
plot_density(value ~ group, data = df, col = c("red", "blue"))

# Compare two vectors
y1 <- rnorm(50)
y2 <- rnorm(50, mean = 1)
plot_density(y1, y2)
```

plot_freq *Plot frequencies for a variable (histogram without binning)*

Description

Creates a frequency plot showing the frequency of every observed value, optionally by group. Most frequent values are labeled by default. The plotted variable may be numeric, character, or factor. Category order on the x-axis is controlled by `x.order`.

Usage

```
plot_freq(
  formula,
  y2 = NULL,
  data = NULL,
  freq = TRUE,
  order = NULL,
  x.order = NULL,
  col = "dodgerblue",
  lwd = 9,
  width = NULL,
  value.labels = "auto",
  ticks.max = 30,
  show.x.value = "auto",
  show.legend = TRUE,
  legend.title = NULL,
  col.text = NULL,
  ...
)
```

Arguments

`formula`	Two possible uses (similar to `t.test()`): • Single Variable (possibly by subgroup): `plot_freq(y)` or `plot_freq(y~x)` • Contrast Two Variables: `plot_freq(y1, y2)` The outcome may be numeric, character, or factor. When comparing two variables, both must be the same kind (both numeric, or both character/factor).
`y2`	optional second variable when contrasting two variables `plot_freq(y1,y2)`
`data`	An optional data frame containing the variables in the formula.
`freq`	Logical. If TRUE (default), displays frequencies. If FALSE, displays percentages.
`order`	Controls the order in which groups appear in the plot and legend. Use `-1` to reverse the default order. Alternatively, provide a vector specifying the exact order (e.g., `c("B", "A", "C")`). If `NULL` (default), groups are ordered by their factor levels (if the grouping variable is a factor) or sorted alphabetically/numerically. Only applies when using grouped plots or comparing two variables.

`x.order` Controls the order of *categories* on the x-axis when the plotted variable is character or factor (ignored for numeric `x`, with a warning if set).

- `NULL` (default): factor level order if `x` is a factor; otherwise alphabetical.
- `-1`: reverse of that default.
- `"alpha"`: alphabetical (even for factors).
- `"freq"`: most frequent to least frequent (ties broken alphabetically).
- `"freq.asc"`: least frequent to most frequent (ties broken alphabetically).
- Character vector: exact category order (must include all observed categories).

For grouped plots, frequency-based orders use overall (pooled) counts.

`col` Color for the bars.

`lwd` Line width for the frequency bars. Default is 9.

`width` Numeric. Width of the frequency bars. If NULL (default), width is automatically calculated based on the spacing between values.

`value.labels` Controls value labeling. If numeric, shows labels for the `value.labels` highest-frequency values (including ties). Use `-1` or `"all"` to show all labels and `0` to show none. Use `"auto"` to label all values when there are 30 or fewer unique values; otherwise label the single most frequent value. For backward compatibility, `TRUE` is treated as `-1` and `FALSE` as `0`.

`ticks.max` Integer. Maximum number of unique x values to label on the x-axis. If there are more than `ticks.max` unique values, `pretty()` ticks are used instead of labeling every value.

`show.x.value` Either `"auto"` (default), `TRUE`, or `FALSE`. If enabled, draws a small `x=<value>` label just above each frequency value label. When `"auto"`, x labels are shown only when plot_freq is not already labeling all x values on the x-axis.

`show.legend` Logical. If TRUE (default), displays a legend when `group` is specified. If FALSE, no legend is shown.

`legend.title` Character string. Title for the legend when `group` is specified. If NULL (default), no title is shown.

`col.text` Color for the value labels. If not specified, uses `col` for non-grouped plots or group colors for grouped plots.

`...` Pass on any argument accepted by `plot()` e.g., `xlab='x-axis'` , `main='Distribution of X'`

Details

This function creates a frequency plot where each observed value is shown with its frequency. Unlike a histogram, there is no binning: each distinct observed value (or category) gets its own bar. For character and factor inputs, use `x.order` to control category order on the x-axis (default: factor levels if available, otherwise alphabetical).

Value

Invisibly returns a data frame with values and their frequencies.

Examples

```
# Simple example
x <- c(1, 1, 2, 2, 2, 5, 5)
plot_freq(x)

# Character categories (alphabetical by default)
cats <- c("a", "a", "b", "c")
plot_freq(cats)
plot_freq(cats, x.order = "freq")  # most frequent first

# Pass on some common \code{plot()} arguments
plot_freq(x, col = "steelblue", xlab = "Value", ylab = "Frequency",ylim=c(0,7))

# Add to an existing plot
plot_freq(x, col = "dodgerblue")


# Compare two vectors
y1 <- c(1, 1, 2, 2, 2, 5, 5)
y2 <- c(1, 2, 2, 3, 3, 3)
plot_freq(y1, y2)

# Using a data frame with grouping
df <- data.frame(value = c(1, 1, 2, 2, 2, 5, 5), group = c("A", "A", "A", "B", "B", "A", "B"))
plot_freq(value ~ 1, data = df)  # single variable
plot_freq(value ~ group, data = df)  # with grouping

# Control group order in legend and plot
plot_freq(value ~ group, data = df, order = c("B", "A"))  # B first, then A
plot_freq(value ~ group, data = df, order = -1)  # Reverse default order
```

`plot_gam` *Plot GAM Model*

Description

Plots fitted GAM values for focal predictor, keeping any other predictors in the model at a specified quantile (default: median)

Usage

```
plot_gam(
  model,
  predictor,
  quantile.others = 50,
  col = "blue4",
  bg = adjustcolor("dodgerblue", 0.2),
  plot2 = "auto",
```

```
  col2 = NULL,
  bg2 = "gray90",
  ...
)
```

Arguments

model	A GAM model object fitted using mgcv::gam().
predictor	Character string specifying the name of the predictor variable to plot on the x-axis.
quantile.others	Number between 1 and 99 for quantile at which all other predictors are held constant. Default is 50 (median).
col	Color for the prediction line. Default is "blue4".
bg	Background color for the confidence band. Default is adjustcolor('dodgerblue', .2).
plot2	How to plot the distribution in the lower plot. Options: 'auto' (default, auto-select based on number of unique values), 'freq' (always plot frequencies), 'density' (always plot the density) or 'none' (neither). When 'auto', plots frequencies when the predictor has fewer than 30 unique values, density otherwise.
col2	Color for the lines/bars in the bottom distribution plot. Default is "dodgerblue"
bg2	Background color for the bottom distribution plot. Default is "gray90".
...	Additional arguments passed to plot() and lines().

Value

Invisibly returns a list containing:

- predictor_values: The sequence of predictor values used
- predicted: The predicted values
- se: The standard errors
- lower: Lower confidence bound (predicted - 2*se)
- upper: Upper confidence bound (predicted + 2*se)

Examples

```
library(mgcv)
# Fit a GAM model
data(mtcars)
mtcars$cyl <- factor(mtcars$cyl)  # Convert to factor before fitting GAM
model <- gam(mpg ~ s(hp) + s(wt) + cyl, data = mtcars)

# Plot effect of hp (with other variables at median)
plot_gam(model, "hp")

# Plot effect of hp (with other variables at 25th percentile)
```

```
plot_gam(model, "hp", quantile.others = 25)

# Customize plot
plot_gam(model, "hp", main = "Effect of Horsepower", col = "blue", lwd = 2)
```

`plot_means` *Barplot of means*

Description

Plots means, with confidence intervals, and (optionally) p-values for differences of means and interactions

Usage

```
plot_means(
  formula,
  data = NULL,
  cluster = NULL,
  tests = "auto",
  quiet = FALSE,
  order = NULL,
  legend.title = NULL,
  col = NULL,
  col.text = NULL,
  values.cex = 1,
  values.align = "top",
  values.round = 1,
  pvalue.cex = 0.9,
  pvalue.col = "gray50",
  ci.level = 95,
  buffer.top = "auto",
  ...
)
```

Arguments

`formula`	A formula, e.g., `y ~ x1+x2`, where x1 & x2 are grouping variables (e.g., condition indicators in a 2x2 experiment). The formula can include up to three grouping variables. The plot will show contiguous bars for x1, and sets of bars for x2 and x3
`data`	Optional data frame containing variables in the formula.
`cluster`	Optional clustering variable when there are repeated observations per cluster e.g., `cluster="participant_ID"`. When provided, inference for reported tests is based on regressions with clustered standard errors (via `lm2(..., clusters=...)`).

tests	specifies which comparisons of means to report. Syntax involves putting column numbers in a character string, with a - to symbolize a comparison and a + symbolizing combination. For example tests="1-2" reports t-test comparing columns 1 and 2. tests="1-2,3-4" t-tests comparing columns 1 & 2 and another 3 & 4. To run an interaction use parentheses: tests="(4-3)-(2-1)" and can also be combined with simple tests. Main effects can be specified using '+', for example (1+2)-(3+4) compares all observations in the first two columns with all observations in the next two columns.
quiet	Logical. When TRUE, suppresses console messages from plot_means().
order	Controls the order of x1 groups (bar order and colors). Use -1 to reverse the default order (e.g., if plot shows 'male' first and 'female' second, order=-1 will flip that).
legend.title	Character string. Title for the legend. If NULL, no title is shown.
col	Color(s) for x1 bars. If NULL, colors are chosen automatically.
col.text	Color for confidence intervals and other non-bar annotations. If NULL, defaults to a dark gray.
values.cex	Numeric scalar controlling text size for mean value labels (and related annotations).
values.align	Where within the bars to put the mean value labels: "top", "middle", "bottom", or "none".
values.round	Non-negative integer. Number of decimal places for mean value labels.
pvalue.cex	Numeric scalar controlling p-value label size.
pvalue.col	Color for p-value brackets/labels.
ci.level	Confidence interval level for ciL/ciH in $means. Default is 95. You can also pass a proportion (e.g., 0.95).
buffer.top	Either "auto" (default) or a numeric value. Extra vertical headroom (as a fraction of the data y-range) added above the maximum y value to make room for annotations. When "auto", uses 0.35 when an interaction p-value is shown (scenario 2) and 0.25 otherwise.
...	Additional arguments passed to plot() (e.g., main, ylim, ylab). If ylim is supplied, the lower limit may be extended slightly downward (below your requested minimum) to leave room for n= sample-size labels; it is not pulled down to 0 unless your lower ylim already includes 0. The upper ylim is unchanged (aside from headroom for p-value brackets).

Details

When tests="auto", the function reports a small default set of differences-of-means tests (when applicable) and, in 2x2 designs, an interaction test:

Differences in means If cluster is NULL, these are Welch two-sample t-tests computed with t.test(..., var.equal=FALSE). If cluster is provided, these comparisons are computed from a regression using lm2() with clustered standard errors.

Interaction The interaction is tested using a linear regression fit with lm2(), even when cluster is NULL; when cluster is provided, the interaction test uses clustered standard errors.

The regression-based tests use heteroskedasticity-robust inference (HC3) when `cluster` is `NULL`. HC3 is a common small-sample adjustment to White-type robust standard errors and is used to reduce sensitivity to heteroskedasticity. When `cluster` is provided, `plot_means()` instead uses clustered standard errors (robust to within-cluster correlation).

In the returned `$means` table, `ciL` and `ciH` are the lower and upper bounds of a `ci.level`% confidence interval for the mean (when available). The same confidence level is used for the confidence-interval whiskers drawn in the figure.

Value

A minimal list returned invisibly with two elements:

`means` A data frame of means (and, when available, confidence intervals) aligned to the plotting grid.

`tests` A data frame of comparisons used for p-value annotation (or `NULL` if not applicable).

Examples

```
df <- data.frame(y = rnorm(100), group = rep(c("A", "B"), 50))
plot_means(y ~ group, data = df)

df2 <- data.frame(
  y = rnorm(200),
  x1 = rep(c("A", "B"), 100),
  x2 = rep(c("X", "Y"), each = 100)
)
plot_means(y ~ x1 + x2, data = df2)

df3 <- data.frame(
  y = rnorm(600),
  x1 = rep(c("control", "treatment"), times = 300),
  x2 = rep(rep(c("low", "high"), each = 150), times = 2),
  x3 = rep(c("online", "lab"), each = 300)
)
plot_means(y ~ x1 + x2 + x3, data = df3)
```

---

predict.lm2 *Predict method for lm2 objects*

Description

Predict method for lm2 objects

Usage

```
## S3 method for class 'lm2'
predict(object, newdata, ...)
```

Arguments

object	An object of class lm2
newdata	An optional data frame in which to look for variables with which to predict. If omitted, the original model data is used.
...	Additional arguments passed to predict.lm_robust, including se.fit and interval.

Value

A vector of predicted values (or a list with fit and se.fit if se.fit = TRUE, or a matrix with fit, lwr, upr if interval is specified)

---

print.desc_var *Print method for desc_var objects*

Description

Print method for desc_var objects

Usage

```
## S3 method for class 'desc_var'
print(x, ...)
```

Arguments

x	An object of class desc_var
...	Additional arguments passed to print.data.frame

Value

Invisibly returns the original object

---

print.lm2 *Print method for lm2 objects*

Description

Print method for lm2 objects

Usage

```
## S3 method for class 'lm2'
print(x, notes, round = NULL, ...)
```

Arguments

x	An object of class lm2
notes	deprecated. Ignored. Use lm2_notes after printing.
round	Optional integer: NULL uses the value from lm2(); -1 for R default format(); non-negative k for fixed decimals.
...	Additional arguments (ignored)

Value

Invisibly returns the original object

---

print.t.test2 *Print method for t.test2 output*

Description

Print method for t.test2 output

Usage

```
## S3 method for class 't.test2'
print(x, ...)
```

Arguments

x	An object of class t.test2
...	Additional arguments passed to print

Value

Invisibly returns the input object x. Called for its side effect of printing a formatted t-test summary to the console, including means, confidence intervals, test statistics, p-values, sample sizes, and APA-formatted results.

print.table2 *Print method for table2 output with centered column variable name*

Description

Print method for table2 output with centered column variable name

Usage

```
## S3 method for class 'table2'
print(x, ...)
```

Arguments

x	An object of class table2
...	Additional arguments passed to print

Value

Invisibly returns the input object x. Called for its side effect of printing a formatted cross-tabulation table to the console. The output includes frequencies, optional relative frequencies (row, column, or overall proportions), and chi-squared test results when applicable.

resize_images *Resize Images*

Description

Saves images to PNG with a specified width. As input it accepts (SVG, PDF, EPS, JPG, JPEG, TIF, TIFF, BMP, PNG) Saves to subdirectory '/resized' within input folder (or same directory as file if input is a single file)

Usage

```
resize_images(path, width)
```

Arguments

path	Character string. Path to a folder containing image files, or path to a single image file.
width	Numeric vector. Target width(s) in pixels for the output PNG files. Can be a single value (recycled for all files) or a vector matching the number of files found.

Details

This function:

- Searches for image files with extensions: svg, pdf, eps, jpg, jpeg, tif, tiff, bmp, png
- Creates a "resized" subfolder in the target directory if it doesn't exist
- Converts each file to PNG format at the specified width(s)
- Saves output files as: `originalname_width.png` in the resized subfolder

Supported input formats:

- Vector graphics: SVG, PDF, EPS (rasterized using rsvg/magick)
- Raster images: JPG, JPEG, TIF, TIFF, BMP, PNG

Value

Invisibly returns TRUE on success.

Note

Dependencies required: `rsvg`, `magick`, and `tools` (base R). SVG files are rasterized using `rsvg::rsvg()`, while PDF/EPS and other formats are handled by `magick::image_read()`.

Examples

```
# Create a temporary PNG file and resize it
tmp_png <- tempfile(fileext = ".png")
grDevices::png(tmp_png, width = 400, height = 300)
old_par <- graphics::par(no.readonly = TRUE)
graphics::par(mar = c(2, 2, 1, 1))
graphics::plot(1:2, 1:2, type = "n")
grDevices::dev.off()
graphics::par(old_par)
resize_images(tmp_png, width = 80)
```

---

round2 *Round with Fixed Decimals*

Description

Like `round()`, but always returns a character vector with exactly `digits` decimal places. By default, drops leading zeros before the decimal point when every finite value has absolute value less than 1 (e.g. `0.34` becomes `.34`).

Usage

```
round2(x, digits = 0, leading.zero = FALSE)
```

Arguments

x	A numeric vector.
digits	Number of decimal places. Default is 0.
leading.zero	Logical. If FALSE (default), drop leading zeros before the decimal when every finite value satisfies |x| < 1. If any finite value has |x| >= 1, leading zeros are kept for the whole vector. If TRUE, leading zeros are always kept.

Value

A character vector of the same length as x.

Examples

```
round2(0.34, 2)
round2(0.34, 2, leading.zero = TRUE)
round2(c(0.34, 0.5), 2)
round2(c(0.34, 1.25), 2)
round2(1.2, 2)
```

---

scatter.gam *Scatter Plot with GAM Smooth Line*

Description

Creates a scatter plot with a GAM (Generalized Additive Model) smooth line. Supports both scatter.gam(x, y) and scatter.gam(y ~ x).

Usage

```
scatter.gam(
  x,
  y,
  data.dots = TRUE,
  three.dots = FALSE,
  data = NULL,
  k = NULL,
  plot.dist = NULL,
  dot.pch = 16,
  dot.col = adjustcolor("gray", 0.7),
  jitter = FALSE,
  ...
)
```

Arguments

x	A numeric vector of x values, or a formula of the form y ~ x.
y	A numeric vector of y values. Not used if x is a formula.
data.dots	Logical. If TRUE, displays data on scatterplot
three.dots	Logical. If TRUE, divides x into tertiles and puts markers on the average x & y for each
data	An optional data frame containing the variables x and y.
k	Optional integer specifying the basis dimension for the smooth term in the GAM model (passed to s(x, k=k)). If NULL (default), uses the default basis dimension.
plot.dist	Character string specifying how to plot the distribution of x underneath the scatter plot. Options: NULL (default, auto-select based on number of unique values), "none" (no distribution plot), "plot_freq" (always use plot_freq()), or "hist" (always use hist()). When NULL, uses plot_freq() if there are 25 or fewer unique values, otherwise uses hist().
dot.pch	Plotting character for data points when data.dots = TRUE. Default is 16 (filled circle).
dot.col	Color for data points when data.dots = TRUE. Default is adjustcolor('gray', 0.7) (semi-transparent gray).
jitter	Logical. If TRUE, applies a small amount of jitter to data points to reduce overplotting. Default is FALSE.
...	Additional arguments passed to plot() and gam(). Common plot arguments include: • main: Custom title for the plot (e.g., main = "My Title") • col: Color of the GAM smooth line (e.g., col = "red") • lwd: Line width of the GAM smooth line (e.g., lwd = 2) • xlim, ylim: Axis limits (e.g., xlim = c(0, 10)) • xlab, ylab: Axis labels (e.g., xlab = "Age")

Details

This function fits a GAM model with a smooth term for x and plots the fitted smooth line. The function uses the mgcv package's gam() function.

When three.dots = TRUE, the x variable is divided into three equal-sized groups (tertiles), and the mean x and y values for each group are plotted as points. This provides a simple summary of the relationship across the range of x.

Value

Invisibly returns the fitted GAM model object.

See Also

scatter.smooth for a simpler loess-based scatter plot smoother.

Examples

```
# Generate sample data for examples
x <- rnorm(100)
y <- 2*x + rnorm(100)

# Plot GAM smooth line only
scatter.gam(x, y)

# Equivalent call using formula syntax (y ~ x)
scatter.gam(y ~ x)

# Include scatter plot with underlying data points behind the GAM line
scatter.gam(x, y, data.dots = TRUE)

# Include summary points showing mean x and y for each tertile bin
scatter.gam(x, y, three.dots = TRUE)

# Customize the plot with a custom title, line color, and line width
scatter.gam(x, y, data.dots = TRUE, col = "red", lwd = 2, main = "GAM Fit")

# Control smoothness of the GAM line by specifying the basis dimension
scatter.gam(x, y, k = 10)
```

---

stimulus.beeswarm *Make beeswarm plots for compared-stimulus designs*

Description

Beeswarm plot for compared-stimulus designs from "Stimulus Sampling Reimagined" (Simonsohn, Montealegre, & Evangelidis, 2025).

Usage

```
stimulus.beeswarm(
  data,
  dv,
  stimulus,
  condition,
  flip.conditions = FALSE,
  dv.is.percentage = FALSE,
  simtot = 500,
  confidence = 95,
  ylim = c(),
  ylab1 = "",
  ylab2 = "",
  xlab1 = "",
  xlab2 = NULL,
```

```
  dot.spacing = "auto",
  col1 = "blue4",
  col2 = "red4",
  main = "",
  watermark = TRUE,
  save.as = "",
  svg.width = "",
  svg.height = "",
  ...
)
```

Arguments

`data`	Data frame containing variables to be analyzed.
`dv`	Name of the dependent variable.
`stimulus`	Name of the variable containing the stimulus ID.
`condition`	Name of the variable containing the condition indicator.
`flip.conditions`	If 'TRUE', reverse the order of condition labels.
`dv.is.percentage`	If 'TRUE', format values as percentages.
`simtot`	Number of bootstraps for the confidence band under homogeneity.
`confidence`	Confidence level for the band (default 95).
`ylim`	Optional y-axis limits.
`ylab1`, `ylab2`	Labels on the y-axis (optional).
`xlab1`, `xlab2`	Labels on the x-axis (optional).
`dot.spacing`	Horizontal distance between stimulus labels ('"auto"' by default).
`col1`, `col2`	Colors for the two conditions.
`main`	Plot title.
`watermark`	If 'TRUE', display package version watermark.
`save.as`	File path for saving the figure ('.svg' or '.png', optional).
`svg.width`	Optional width when saving.
`svg.height`	Optional height when saving.
`...`	Additional arguments passed to [graphics::plot.default()].

Value

Invisibly, a two-column matrix of beeswarm coordinates.

`stimulus.plot` *Make stimulus plots as in "Stimulus Sampling Reimagined"*

Description

Build stimulus plots comparing results for individual stimuli in an experiment (Simonsohn, Montealegre, & Evangelidis, 2025).

Usage

```
stimulus.plot(
  plot.type = "means",
  data,
  dv,
  condition,
  stimulus,
  participant = "",
  save.as = "",
  svg.width = "",
  svg.height = "",
  sort.by = "",
  flip.conditions = FALSE,
  model = c(),
  overall.estimate = c(),
  overall.ci = c(),
  overall.p = c(),
  overall.label = c(),
  ylab1 = "",
  ylab2 = "",
  xlab1 = "Stimuli",
  xlab2 = "",
  decimals = "auto",
  null.method = "shuffle",
  dv.is.percentage = FALSE,
  legend.title = "",
  simtot = 1000,
  watermark = TRUE,
  seed = 2024,
  ylim = c(),
  main = "",
  ...
)
```

Arguments

`plot.type`	Either ‘"means"‘ or ‘"effects"‘; determines what is plotted on the y-axis.
`data`	Data frame containing variables to be analyzed.

dv	Name of the dependent variable (e.g., ‘dv = ’y’‘); quotes are not required.
condition	Name of the variable containing the condition indicator.
stimulus	Name of the variable containing the stimulus ID.
participant	Name of the variable containing participant IDs; necessary for valid inference when ‘plot.type = ’effects’‘ and each participant provided more than one observation.
save.as	File path for saving the figure (‘.svg‘ or ‘.png‘, optional).
svg.width	Optional width when saving to SVG/PNG.
svg.height	Optional height when saving to SVG/PNG.
sort.by	Variable to sort stimuli by. Defaults to sorting by observed effect size.
flip.conditions	If ‘TRUE‘, subtract the first condition from the second instead of the default.
model	Method used to compute overall average: ‘’regression’‘, ‘’intercepts’‘, ‘’slopes’‘, and/or ‘’all’‘.
overall.estimate	Scalar or vector of overall average effect computed outside statuser.
overall.ci	Confidence interval bounds for ‘overall.estimate‘.
overall.p	P-value for overall average effect computed outside statuser.
overall.label	Label for overall averages on the x-axis.
ylab1, ylab2	Labels on the y-axis (optional).
xlab1, xlab2	Labels on the x-axis (optional).
decimals	Number of decimals for value labels (‘"auto"‘ by default).
null.method	Null method for effects plots: ‘"shuffle"‘ or ‘"demean"‘.
dv.is.percentage	If ‘TRUE‘, format the dependent variable as percentages.
legend.title	Text with title above legend (optional).
simtot	Number of resamples for heterogeneity under null (effects plots).
watermark	If ‘TRUE‘, display package version watermark.
seed	Seed used when resampling for effects plots.
ylim	Optional y-axis limits.
main	Plot title.
...	Additional arguments passed to [graphics::plot.default()].

Value

Invisibly, a data frame (means plot) or list (effects plot).

summary.lm2 *Summary method for lm2 objects*

Description

Summary method for lm2 objects

Usage

```
## S3 method for class 'lm2'
summary(object, ...)
```

Arguments

object	An object of class lm2
...	Additional arguments passed to print.lm2

Value

Invisibly returns the original object

t.test2 *Enhanced alternative to t.test()*

Description

The basic t-test function in R, t.test, does not report the observed difference of means, does not stipulate which mean is subtracted from which (i.e., whether it computed A-B or B-A), and presents the test results on the console in a verbose unorganized paragraph of text. t.test2 improves on all those counts, and in addition, it reports the number of observations per group and if any observations are missing it issues a warning. It returns a dataframe instead of a list. The function is also registered as t.test2 for the t generic to satisfy S3 registration checks while keeping direct calls to t.test2(...) unchanged.

Usage

```
## S3 method for class 'test2'
t(x, ...)
```

Arguments

x	The first data argument passed to t.test. This can be a numeric vector or a formula.
...	Further arguments passed to t.test. The first unnamed argument (or a named y) is the optional second sample. A numeric vector is a second sample; a character, factor, or logical vector with exactly two levels and the same length as x is a grouping variable, so t.test2(dfy, dfgroup) matches t.test2(y ~ group, data = df). Numeric vectors are never treated as groups; use a formula for numeric group codes such as 0/1. The formals are function(x, ...) so this S3 method for t stays consistent with the generic.

Value

A data frame with class c("t.test2", "data.frame") containing a single row with the following columns:

mean columns One or two columns containing group means, named after the input variables (e.g., men, women) or Group 1, Group 2 for long names.

diff column For two-sample tests, the difference between means (e.g., men-women).

d For two-sample independent tests only, Cohen's d computed as the mean difference divided by the pooled standard deviation.

ci The confidence level as a string (e.g., "95 percent").

ci.L, ci.H Lower and upper bounds of the confidence interval.

t The t-statistic.

df Degrees of freedom.

p.value The p-value.

N columns Sample sizes, named N(group1), N(group2) or N1, N2. For paired tests, a single N column.

correlation For paired tests only, the correlation between pairs.

Attributes store additional information including missing value counts and test type (one-sample, two-sample, paired, Welch vs. Student).

Examples

```
# Two-sample t-test
men <- rnorm(100, mean = 5, sd = 1)
women <- rnorm(100, mean = 4.8, sd = 1)
t.test2(men, women)

# Paired t-test
x <- rnorm(50, mean = 5, sd = 1)
y <- rnorm(50, mean = 5.2, sd = 1)
t.test2(x, y, paired = TRUE)

# One-sample t-test
data <- rnorm(100, mean = 5, sd = 1)
```

```
t.test2(data, mu = 0)

# Formula syntax
data <- data.frame(y = rnorm(100), group = rep(c("A", "B"), 50))
t.test2(y ~ group, data = data)

# Outcome vector and grouping variable
t.test2(data$y, data$group)
```

`table2` *Enhanced alternative to table()*

Description

The function `table` does not show variable names when tabulating from a dataframe, requires running another function, `prop.table`, to tabulate proportions and yet another function, `chisq.test` to test difference of proportions. `table2` does what those three functions do, producing easier to read output, and always shows variable names.

Usage

```
table2(
  ...,
  data = NULL,
  exclude = if (useNA == "no") c(NA, NaN),
  useNA = c("no", "ifany", "always"),
  dnn = NULL,
  deparse.level = 1,
  prop = NULL,
  digits = 3,
  chi = FALSE,
  chi2 = NULL,
  correct = FALSE
)
```

Arguments

`...`	Variables to tabulate, as in `table` (e.g., vectors or `df$col` expressions). Extra named arguments below may also be passed here.
`data`	Unused; reserved for compatibility. Prefer `df$col` in `...`.
`exclude`	Values to exclude, passed to `table`.
`useNA`	Whether to include `NA` levels, passed to `table`.
`dnn`	Optional dimension names, passed to `table`.
`deparse.level`	Passed to `table` for naming dimensions from deparsed expressions when `dnn` is not set.

prop	report a table with: • prop="all": Proportions for full table (each cell / total) • prop="row": Proportions by row ('rows' also accepted) • prop="col": Proportions by columns ('cols', 'column', 'columns' also accepted)
digits	Number of decimal values to show for proportions
chi	Logical. If TRUE, performs a chi-square test on frequency table, reports results in APA format. chi2 is accepted as an alias.
chi2	Alias for chi. If both are provided, they must agree.
correct	Logical. If TRUE, applies Yates' continuity correction for 2x2 tables in the chi-square test. Default is FALSE (no correction).

Value

A list (object of class "table2") with the following components:

- freq: frequency table
- prop: proportions table
- chisq: chi-square test

Examples

```
# Create example data
df <- data.frame(
  group = c("A", "A", "B", "B", "A"),
  status = c("X", "Y", "X", "Y", "X")
)

# Enhanced table with variable names (2 variables)
table2(df$group, df$status)

# Enhanced table with variable names (3 variables)
df3 <- data.frame(
  x = c("A", "A", "B", "B"),
  y = c("X", "Y", "X", "Y"),
  z = c("high", "low", "high", "low")
)
table2(df3$x, df3$y, df3$z)

# Table with proportions
table2(df$group, df$status, prop = 'all')  # Overall proportions
table2(df$group, df$status, prop = 'row')  # Row proportions
table2(df$group, df$status, prop = 'col')  # Column proportions

# Table with chi-square test
table2(df$group, df$status, chi = TRUE,prop='all')
```

tacos_data *Fictitious taco spiciness study for interprobe examples*

Description

Simulated data inspired by the taco vignette in Montealegre and Simonsohn (2026). Respondents vary in spice `tolerance`; salsa `spicy`ness ranges from 1 (mild) to 7 (hot); `enjoyment` is highest when spiciness roughly matches tolerance. Hot vs mild salsa is disliked at low tolerance and preferred at high tolerance (nonlinear interaction).

Format

A data frame with 1000 rows and 6 variables:

spicy Numeric salsa spiciness on $[1, 7]$

salsa Factor with levels `Mild`, `Medium`, `Hot` (cut from `spicy`: $[1, 3]$, $(3, 5]$, $(5, 7]$)

spicy_bin Factor `Mild`/`Hot` when `spicy` is near 1 (≤ 1.5) or near 7 (≥ 6.5); `NA` otherwise (use `!is.na(spicy_bin)` for the binary design)

tolerance Numeric spice tolerance on about 1–7

enjoyment Numeric taco enjoyment on about 15–100 (kept well above 0)

age Integer age (nuisance covariate for fitted-model examples)

Source

Simulated in '`data-raw/tacos_data.R`' with seed 20260727.

Examples

```
data(tacos_data)
head(tacos_data)

tacos_bin <- tacos_data[!is.na(tacos_data$spicy_bin), ]
interprobe(x = spicy_bin, z = tolerance, y = enjoyment, data = tacos_bin)
```

text2 *Enhanced alternative to text()*

Description

Adds to text() optional background color and horizontal alignment (align='center')

Usage

```
text2(
  x,
  y,
  labels,
  align = "center",
  bg = "papayawhip",
  cex = 1,
  pad = 0.03,
  pad_v = 0.25,
  ...
)
```

Arguments

x, y	coordinates for text placement
labels	text to display
align	alignment in relation to x coordinate ('left','center','right')
bg	background color
cex	character expansion factor
pad	left/right padding in percentage (e.g., .03)
pad_v	top/bottom padding in percentage (e.g., .25)
...	Additional arguments passed to text.

Value

No return value, called for side effects. Adds text with an optional background rectangle to an existing plot.

Examples

```
# Create a simple plot
plot(1:10, 1:10, type = "n", main = "text2() - Alignment & Color")

# Alignment with respect to x=5
text2(5, 8, "align='left' from 5", align = "left", bg = "yellow1")
text2(5, 7, "align='right' from 5", align = "right", bg = "blue", col = "white")
text2(5, 6, "align='center' from 5", align = "center", bg = "black", col = "white")
abline(v = 5, lty = 2)

# Multiple labels with different alignments
text2(c(2, 5, 8), c(5, 5, 5),
      labels = c("Left", "Center", "Right"),
      align = c("left", "center", "right"),
      bg = c("pink", "lightblue", "lightgreen"))

# Text with custom font color (passed through ...)
text2(5, 3, "Red Text", col = "red", bg = "white")
```

```
# Padding examples
plot(1:10, 1:10, type = "n", main = "Padding Examples")

# Default padding (pad=0.03, pad_v=0.25)
text2(5, 8, "Default padding", bg = "lightblue")

# More horizontal padding
text2(5, 6, "Wide padding", pad = 0.2, bg = "lightgreen")

# More vertical padding
text2(5, 4, "Tall padding", pad_v = 0.8, bg = "lightyellow")

# Both padding increased
text2(5, 2, "Extra padding", pad = 0.15, pad_v = 0.6, bg = "pink")
```

`twolines` *Two-Lines Test of U-Shapes*

Description

Implements the two-lines test for U-shaped (or inverted U-shaped) relationships introduced by Simonsohn (2018).

Usage

```
twolines(
  f,
  graph = 1,
  link = "gaussian",
  data = NULL,
  pngfile = "",
  quiet = FALSE
)
```

Arguments

`f`	A formula object specifying the model (e.g., y ~ x1 + x2 + x3). The first predictor is the one tested for a u-shaped relationship.
`graph`	Integer. If 1 (default), produces a plot. If 0, no plot is generated.
`link`	Character string specifying the link function for the GAM model. Default is "gaussian".
`data`	An optional data frame containing the variables in the formula. If not provided, variables are evaluated from the calling environment.
`pngfile`	Optional character string. If provided, saves the plot to a PNG file with the specified filename.
`quiet`	Logical. If TRUE, suppresses the Robin Hood details messages. Default is FALSE.

Details

Reference: Simonsohn, Uri (2018) "Two lines: A valid alternative to the invalid testing of U-shaped relationships with quadratic regressions." AMPPS, 538-555. doi:10.1177/2515245918805755

The test beings fitting a GAM model, predicting y with a smooth of x, and optionally with covariates. It identifies the interior most extreme value of fitted y, and adjusts from the matching x-value to set the breakpoint relying on the Robin Hood procedure introduced also by Simonsohn (2018). It then estimates the (once) interrupted regression using that breakpoint, and reports the slope and significance of the average slopes at either side of it. A U-shape is significant if the slopes are of opposite sign and are both individually significant.

Value

A list containing:

- All elements from `reg2()`: `b1`, `b2`, `p1`, `p2`, `z1`, `z2`, `u.sig`, `xc`, `glm1`, `glm2`, `rob1`, `rob2`, `msg`, `yhat.smooth`
- `yobs`: Observed y values (adjusted for covariates if present)
- `y.hat`: Fitted values from GAM
- `y.ub`, `y.lb`: Upper and lower bounds for fitted values
- `y.most`: Most extreme fitted value
- `x.most`: x-value associated with most extreme fitted value
- `f`: Formula as character string
- `bx1`, `bx2`: Linear and quadratic coefficients from preliminary quadratic regression
- `minx`: Minimum x value
- `midflat`: Median of flat region
- `midz1`, `midz2`: Z-statistics at midpoint

Examples

```
# Simple example with simulated data
set.seed(123)
x <- rnorm(100)
y <- -x^2 + rnorm(100)
data <- data.frame(x = x, y = y)
result <- twolines(y ~ x, data = data)

# With covariates
z <- rnorm(100)
y <- -x^2 + 0.5*z + rnorm(100)
data <- data.frame(x = x, y = y, z = z)
result <- twolines(y ~ x + z, data = data)

# Without data argument (variables evaluated from environment)
x <- rnorm(100)
y <- -x^2 + rnorm(100)
result <- twolines(y ~ x)
```

`var_labels` *Get or set variable labels*

Description

Assign labels to describe variables in a data frame. Can be called on a single variable or full data, to assign or to read existing labels.

Usage

```
var_labels(x)

var_labels(x) <- value
```

Arguments

`x`	A vector or a data frame.
`value`	Character label(s) to assign.

Details

For a vector, this reads/writes a base-R `"label"` attribute. For a data frame, this reads/writes the `"label"` attribute of each column.

Value

- If `x` is a data frame: a named character vector with one entry per column (missing labels are returned as `NA_character_`). - Otherwise: a single character string (or `NA_character_`).

Examples

```
df <- data.frame(x = 1:3, y = 4:6)

# Set labels for all columns
var_labels(df) <- c("this is x", "this is y")
var_labels(df)

# Set a label for a single column
var_labels(df$x) <- "this is x"
var_labels(df$x)
```

Index

∗ **datasets**
    interprobe_sim, 12
    tacos_data, 44

chisq.test, *42*
clear, 3
clear_stimulus_cache, 4
convert_to_sql, 4

density, *22*
desc_var, 5

format_pvalue, 7

gam, *11*

interprobe, 8
interprobe_sim, 12

list, *13*
list2, 13
lm, *14*
lm2, *11*, 14, *18*
lm2_notes, *14, 15*, 17, *31*
lm_robust, *14, 15*

message2, 18

plot_cdf, 19
plot_density, 21
plot_freq, 23
plot_gam, 25
plot_means, 27
predict.lm2, 29
predict.lm_robust, *30*
print.desc_var, 30
print.lm2, *18*, 30, *40*
print.t.test2, 31
print.table2, 32
prop.table, *42*

resize_images, 32
round2, 33

scatter.gam, *15*, 34
scatter.smooth, *35*
stimulus.beeswarm, 36
stimulus.plot, 38
summary.lm2, 40

t, *40, 41*
t.test, *40, 41*
t.test2, 40
table, *42*
table2, 42
tacos_data, *11*, 44
text, *45*
text2, 44
twolines, 46

var_labels, 48
var_labels<- *(*var_labels*)*, 48