

# Package: pdi (via r-universe)

July 2, 2026

**Type** Package

**Title** Phenotypic Index Measures for Oak Decline Severity

**Version** 0.4.3

**Description** Oak declines are complex disease syndromes and consist of many visual indicators that include aspects of tree size, crown condition and trunk condition. This can cause difficulty in the manual classification of symptomatic and non-symptomatic trees from what is in reality a broad spectrum of oak tree health condition. Two phenotypic oak decline indexes have been developed to quantitatively describe and differentiate oak decline syndromes in *Quercus robur*. This package provides a toolkit to generate these decline indexes from phenotypic descriptors using the machine learning algorithm random forest. The methodology for generating these indexes is outlined in Finch et al. (2021) <[doi:10.1016/j.foreco.2021.118948](https://doi.org/10.1016/j.foreco.2021.118948)>.

**URL** <https://jasenfinch.github.io/pdi/>

**BugReports** <https://github.com/jasenfinch/pdi/issues>

**License** GPL-3

**Encoding** UTF-8

**Imports** dplyr, magrittr, purrr, randomForest, readxl, stringr, tibble, tidy, tidyselect

**RoxygenNote** 8.0.0

**Suggests** testthat, covr, knitr, rmarkdown, ggplot2

**VignetteBuilder** knitr

**NeedsCompilation** no

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**Repository** <https://cran.r-universe.dev>

**Date/Publication** 2026-07-02 13:20:02 UTC

**RemoteUrl** <https://github.com/cran/pdi>

**RemoteRef** HEAD

**RemoteSha** 7d20ab5983999143bf37ec5b54eea4059bffb08b

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agrilusExitHoleDensity

*Agrilus exit hole density ( $m^{-2}$ )*

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### Description

Calculate *Agrilus biguttatus* exit hole density.

### Usage

```
agrilusExitHoleDensity(n, d, s = 2)
```

### Arguments

n	number of <i>Agrilus</i> exit holes
d	diameter at breast height (m)
s	height to which stem surveyed from the tree base (m)

**Examples**

```
agrilusExitHoleDensity(2,1.02,1.3)
```

---

bleedPrevalence	<i>Estimated bleed prevalence (%)</i>
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**Description**

Calculate estimated bleed prevalence.

**Usage**

```
bleedPrevalence(a, A, b, B, d, s = 3)
```

**Arguments**

a	average active bleed size (mm)
A	number of active bleeds
b	average black stain size (mm)
B	number of black stains
d	diameter at breast height (m)
s	height to which stem surveyed from the tree base (m)

**Examples**

```
bleedPrevalence(30,10,40,5,1,1.3)
```

---

calcDIs	<i>Calculate Decline Indexes</i>
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**Description**

Calculate Phenotypic Decline Index (PDI) and Decline Acuteness Index (DAI).

**Usage**

```
calcDIs(rfModels, PDI = TRUE, DAI = TRUE, invertPDI = TRUE, invertDAI = TRUE)
```

**Arguments**

rfModels	list containing random forest models as returned by rf()
PDI	TRUE/FALSE, calculate PDI?
DAI	TRUE/FALSE, calculate DAI?
invertPDI	invert the PDI scale? TRUE/FALSE. Ignored if argument PDI is FALSE
invertDAI	invert the DAI scale? TRUE/FALSE. Ignored if argument DAI is FALSE

**Examples**

```

library(dplyr)

## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',
  package = 'pdi'),full.names = TRUE)

## Prepare data
d <- map(files,readPhenotypeSheet) %>%
  map(preparePhenotypeData) %>%
  bind_rows() %>%
  siteAdjustment() %>%
  mutate(`Live crown ratio (%)` = liveCrownRatio(`Total height (m)`,
    `Lower crown height (m)`),
    `Crown condition (%)` = crownCondition(`Missing crown (%)`,
      `Crown transparency (%)`),
    `Crown volume (m^3)` = crownVolume(`Crown radius (m)`,
      `Total height (m)`,
      `Lower crown height (m)`,
      `Crown condition (%)`),
    `Bleed prevalence (%)` = bleedPrevalence(`Active bleed length (mm)`,
      `Active bleeds`,
      `Black staining length (mm)`,
      `Black staining`,
      `Diameter at breast height (m)`),
    `Agrilus exit hole density (m^-2)` = agrilusExitHoleDensity(`Agrilus exit holes`,
      `Diameter at breast height (m)`))
)

t <- makeAnalysisTable(d)

## Generate random forest models
m <- rf(t,cls = NULL,nreps = 10)

## Calculate decline indexes
DIs <- calcDIs(m,DAI = FALSE,invertPDI = FALSE) %>%
  bind_cols(d %>%
    select(Location,ID,Status))

```

---

crownCondition

*Crown condition*


---

**Description**

Calculate crown condition (%).

**Usage**

```
crownCondition(m, t)
```

**Arguments**

m                    missing crown (%)  
t                    crown transparency (%)

**Examples**

`crownCondition(50,60)`

---

`crownProductionEfficiency`  
*Crown production efficiency*

---

**Description**

Calculate the crown production efficiency

**Usage**

`crownProductionEfficiency(crown_surface_area, crown_volume)`

**Arguments**

`crown_surface_area`  
                         crown surface area (m<sup>2</sup>)  
`crown_volume`      crown volume (m<sup>3</sup>)

**Examples**

`crownProductionEfficiency(34,35)`

---

`crownSurfaceArea`      *Crown surface area (m<sup>2</sup>)*

---

**Description**

Calculate the crown surface area

**Usage**

`crownSurfaceArea(r, h, l, c)`

**Arguments**

r	crown radius (m)
h	total height (m)
l	lower crown height (m)
c	crown condition (%)

**Examples**

```
crownSurfaceArea(3,15,10,50)
```

---

crownVolume	<i>Estimated crown volume (m<sup>3</sup>)</i>
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---

**Description**

Calculate estimated crown volume.

**Usage**

```
crownVolume(r, h, l, c)
```

**Arguments**

r	crown radius (m)
h	total height (m)
l	lower crown height (m)
c	crown condition (%)

**Examples**

```
crownVolume(3,15,10,50)
```

---

```
descriptorContributions
      Descriptor contributions
```

---

**Description**

Calculate average descriptor contributions to random forest models.

**Usage**

```
descriptorContributions(rfModels)
```

**Arguments**

rfModels            list containing random forest models as returned by rf()

**Details**

See see ?randomForest::importance for details on random forest importance metrics.

**Examples**

```
library(dplyr)

## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',
  package = 'pdi'),full.names = TRUE)

## Prepare data
d <- map(files,readPhenotypeSheet) %>%
  map(preparePhenotypeData) %>%
  bind_rows() %>%
  siteAdjustment() %>%
  mutate(`Live crown ratio (%)` = liveCrownRatio(`Total height (m)`,
    `Lower crown height (m)`),
    `Crown condition (%)` = crownCondition(`Missing crown (%)`,
    `Crown transparency (%)`),
    `Crown volume (m^3)` = crownVolume(`Crown radius (m)`,
    `Total height (m)`,
    `Lower crown height (m)`,
    `Crown condition (%)`),
    `Bleed prevalence (%)` = bleedPrevalence(`Active bleed length (mm)`,
    `Active bleeds`,
    `Black staining length (mm)`,
    `Black staining`,
    `Diameter at breast height (m)`),
    `Agrilus exit hole density (m^-2)` = agrilusExitHoleDensity(`Agrilus exit holes`,
    `Diameter at breast height (m)`))
)
```

```
t <- makeAnalysisTable(d)

## Generate random forest models
m <- rf(t,cls = NULL,nreps = 10)

descriptor_contributions <- m %>%
  descriptorContributions()
```

---

liveCrownRatio	<i>Live crown ratio</i>
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---

### Description

Calculate the live crown ratio

### Usage

```
liveCrownRatio(h, l)
```

### Arguments

h	total height (m)
l	lower crown height (m)

### Examples

```
liveCrownRatio(15,10)
```

---

makeAnalysisTable	<i>Make analysis table</i>
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---

### Description

prepare data table ready for random forest analysis

### Usage

```
makeAnalysisTable(phenoData)
```

### Arguments

phenoData	tibble containing phenotype data
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**Examples**

```

library(dplyr)

## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',
  package = 'pdi'),full.names = TRUE)

## Prepare data
d <- map(files,readPhenotypeSheet) %>%
  map(preparePhenotypeData) %>%
  bind_rows() %>%
  siteAdjustment() %>%
  mutate(`Live crown ratio (%)` = liveCrownRatio(`Total height (m)`,
    `Lower crown height (m)`),
    `Crown condition (%)` = crownCondition(`Missing crown (%)`,
      `Crown transparency (%)`),
    `Crown volume (m^3)` = crownVolume(`Crown radius (m)`,
      `Total height (m)`,
      `Lower crown height (m)`,
      `Crown condition (%)`),
    `Bleed prevalence (%)` = bleedPrevalence(`Active bleed length (mm)`,
      `Active bleeds`,
      `Black staining length (mm)`,
      `Black staining`,
      `Diameter at breast height (m)`),
    `Agrilus exit hole density (m^-2)` = agrilusExitHoleDensity(`Agrilus exit holes`,
      `Diameter at breast height (m)`))
)

t <- makeAnalysisTable(d)

```

mds

*Multidimensional scaling***Description**

perform multidimensional scaling of random forest proximities

**Usage**

```
mds(rfModels, dimensions = 2)
```

**Arguments**

`rfModels` list containing random forest models as returned by `rf()`  
`dimensions` number of dimensions to scale to

**Examples**

```

library(dplyr)

## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',
  package = 'pdi'),full.names = TRUE)

## Prepare data
d <- map(files,readPhenotypeSheet) %>%
  map(preparePhenotypeData) %>%
  bind_rows() %>%
  siteAdjustment() %>%
  mutate(`Live crown ratio (%)` = liveCrownRatio(`Total height (m)`,
    `Lower crown height (m)`),
    `Crown condition (%)` = crownCondition(`Missing crown (%)`,
      `Crown transparency (%)`),
    `Crown volume (m^3)` = crownVolume(`Crown radius (m)`,
      `Total height (m)`,
      `Lower crown height (m)`,
      `Crown condition (%)`),
    `Bleed prevalence (%)` = bleedPrevalence(`Active bleed length (mm)`,
      `Active bleeds`,
      `Black staining length (mm)`,
      `Black staining`,
      `Diameter at breast height (m)`),
    `Agrilus exit hole density (m^-2)` = agrilusExitHoleDensity(`Agrilus exit holes`,
      `Diameter at breast height (m)`
  )

t <- makeAnalysisTable(d)

## Generate random forest models
m <- rf(t,cls = NULL,nreps = 10)

mds_data <- mds(m,2)

```

minMaxScale

*Min-max scaling***Description**

Variable min-max scaling.

**Usage**

```
minMaxScale(vec)
```

**Arguments**

vec                    vector of numbers to scale

**Examples**

```
set.seed(1234)

d <- runif(20,1,10)

minMaxScale(d)
```

---

phenotypingTemplate    *Phenotyping template*

---

**Description**

Export a copy of the oak phenotyping data collection spreadsheet.

**Usage**

```
phenotypingTemplate(path = ".")
```

**Arguments**

path                    directory path for export output

**Examples**

```
## Not run:
phenotypingTemplate()

## End(Not run)
```

---

preparePhenotypeData    *Prepare phenotype data*

---

**Description**

Process parsed phenotype data sheets into a tibble suitable for random forest analysis.

**Usage**

```
preparePhenotypeData(phenotypeData)
```

**Arguments**

phenotypeData    parsed phenotype data collection sheet returned from readPhenotypeSheet

**Examples**

```
library(dplyr)

## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',
  package = 'pdi'),full.names = TRUE)

## Prepare data
d <- map(files,readPhenotypeSheet) %>%
  map(preparePhenotypeData)
```

---

readPhenotypeSheet      *Read phenotyping sheet*

---

**Description**

Parse .xlsx phenotype data collection sheets.

**Usage**

```
readPhenotypeSheet(file)
```

**Arguments**

file                      file path to excel file to parse

**Examples**

```
library(dplyr)

## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',
  package = 'pdi'),full.names = TRUE)

## Prepare data
d <- readPhenotypeSheet(files[1])
```

---

rf                              *Random forest analysis*

---

**Description**

Perform random forest repetitions.

**Usage**

```
rf(analysisTable, cls, params = list(), nreps = 100, seed = 1234)
```

**Arguments**

analysisTable	tibble of phenotype data suitable for random forest analysis as returned by preparePhenotypeData
cls	analysisTable column to use as response vector. NULL for unsupervised analyses.
params	additional arguments to pass to randomForest::randomForest
nreps	number of repetitions
seed	random number seed

**Examples**

```
library(dplyr)

## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',
  package = 'pdi'),full.names = TRUE)

## Prepare data
d <- map(files,readPhenotypeSheet) %>%
  map(preparePhenotypeData) %>%
  bind_rows() %>%
  siteAdjustment() %>%
  mutate(`Live crown ratio (%)` = liveCrownRatio(`Total height (m)`,
    `Lower crown height (m)`),
    `Crown condition (%)` = crownCondition(`Missing crown (%)`,
      `Crown transparency (%)`),
    `Crown volume (m^3)` = crownVolume(`Crown radius (m)`,
      `Total height (m)`,
      `Lower crown height (m)`,
      `Crown condition (%)`),
    `Bleed prevalence (%)` = bleedPrevalence(`Active bleed length (mm)`,
      `Active bleeds`,
      `Black staining length (mm)`,
      `Black staining`,
      `Diameter at breast height (m)`),
    `Agrilus exit hole density (m^-2)` = agrilusExitHoleDensity(`Agrilus exit holes`,
      `Diameter at breast height (m)`)
  )

t <- makeAnalysisTable(d)

## Generate random forest models
m <- rf(t,cls = NULL,nreps = 10)
```

---

siteAdjustment

*Site adjustment*


---

**Description**

Perform a site adjustment of selected descriptors.

**Usage**

```
siteAdjustment(
  phenoData,
  descriptors = c("Diameter at breast height (m)", "Lower crown height (m)",
    "Timber height (m)", "Total height (m)", "Crown radius (m)")
)
```

**Arguments**

phenoData      phenoData tibble containing phenotype data  
 descriptors    columns of phenoData on which to perform site correction

**Examples**

```
library(dplyr)

## Retrieve file paths for example data
files <- list.files(system.file("phenotypeDataCollectionSheets",
  package = "pdi"
), full.names = TRUE)

## Prepare data
d <- map(files, readPhenotypeSheet) %>%
  map(preparePhenotypeData) %>%
  bind_rows() %>%
  siteAdjustment()
```

---

siteAdjustmentFactors *Site adjustment factors*

---

**Description**

Return site adjustment factors of selected phenotypic descriptors.

**Usage**

```
siteAdjustmentFactors(
  phenoData,
  descriptors = c("Diameter at breast height (m)", "Lower crown height (m)",
    "Timber height (m)", "Total height (m)", "Crown radius (m)")
)
```

**Arguments**

phenoData      phenoData tibble containing phenotype data  
 descriptors    columns of phenoData on which calculate site correction factors

**Examples**

```
library(dplyr)

## Retrieve file paths for example data
files <- list.files(system.file("phenotypeDataCollectionSheets",
  package = "pdi"
), full.names = TRUE)

## Prepare data
d <- map(files, readPhenotypeSheet) %>%
  map(preparePhenotypeData) %>%
  bind_rows() %>%
  siteAdjustment()

sa_factors <- siteAdjustmentFactors(d)
```

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