

# Package: eyetools (via r-universe)

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**Type** Package

**Title** Analyse Eye Data

**Version** 0.7.2

**Description** Enables the automation of actions across the pipeline, including initial steps of transforming binocular data and gap repair to event-based processing such as fixations, saccades, and entry/duration in Areas of Interest (AOIs). It also offers visualisation of eye movement and AOI entries. These tools take relatively raw (trial, time, x, and y form) data and can be used to return fixations, saccades, and AOI entries and time spent in AOIs. As the tools rely on this basic data format, the functions can work with data from any eye tracking device. Implements fixation and saccade detection using methods proposed by Salvucci and Goldberg (2000) [doi:10.1145/355017.355028](https://doi.org/10.1145/355017.355028).

**License** GPL-3

**URL** <https://tombeesley.github.io/eyetools/>

**BugReports** <https://github.com/tombeesley/eyetools/issues>

**Depends** R (>= 2.10)

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AOI_seq	<i>Sequence analysis of area of interest entries</i>
---------	--

---

## Description

Analyses the sequence of entries into defined AOI regions across trials. Can only be used with fixation data with a "fix\_n" column denoting fixation events.

## Usage

```
AOI_seq(
  data,
  AOIs,
  AOI_names = NULL,
  sample_rate = NULL,
  long = TRUE,
  participant_ID = "participant_ID"
)
```

**Arguments**

data	A dataframe with fixation data (from fixation_dispersion). Either single or multi participant data
AOIs	A dataframe of areas of interest (AOIs), with one row per AOI (x, y, width_radius, height).
AOI_names	An optional vector of AOI names to replace the default "AOI_1", "AOI_2", etc.
sample_rate	Optional sample rate of the eye-tracker (Hz) for use with raw_data. If not supplied, the sample rate will be estimated from the time column and the number of samples.
long	Whether to return the AOI fixations in long or wide format. Defaults to long
participant_ID	the variable that determines the participant identifier. If no column present, assumes a single participant

**Value**

a dataframe containing the sequence of entries into AOIs on each trial.

If long is TRUE, then each AOI entry is returned on a new row, if FALSE, then a row per trial is returned with all AOI entries in one character string

**Examples**

```
data <- combine_eyes(HCL)
fix_d <- fixation_dispersion(data, participant_ID = "pNum")

AOI_seq(fix_d, AOIs = HCL_AOIs, participant_ID = "pNum")
```

---

AOI\_time

*Time analysis of area of interest entries*


---

**Description**

Analyses total time on defined AOI regions across trials. Works with fixation and raw data as the input (must use one or the other, not both).

**Usage**

```
AOI_time(
  data,
  data_type = NULL,
  AOIs,
  AOI_names = NULL,
  sample_rate = NULL,
  as_prop = FALSE,
  trial_time = NULL,
  participant_ID = "participant_ID"
)
```

**Arguments**

<code>data</code>	A dataframe of either fixation data (from <code>fix_dispersion</code> ) or raw data
<code>data_type</code>	Whether data is a fixation ("fix") or raw data ("raw")
<code>AOIs</code>	A dataframe of areas of interest (AOIs), with one row per AOI (x, y, width_radius, height).
<code>AOI_names</code>	An optional vector of AOI names to replace the default "AOI_1", "AOI_2", etc.
<code>sample_rate</code>	Optional sample rate of the eye-tracker (Hz) for use with data. If not supplied, the sample rate will be estimated from the time column and the number of samples.
<code>as_prop</code>	whether to return time in AOI as a proportion of the total time of trial
<code>trial_time</code>	a vector of the time taken in each trial. Equal to the length of x trials by y participants in the dataset
<code>participant_ID</code>	the variable that determines the participant identifier. If no column present, assumes a single participant

**Details**

AOI\_time can take either single participant data or multiple participants where there is a variable for unique participant identification. The function looks for an identifier named `participant_ID` by default and will treat this as multiple-participant data as default, if not it is handled as single participant data, or the `participant_ID` needs to be specified

**Value**

a dataframe containing the time on the passed AOIs for each trial. One column for each AOI separated by trial.

**Examples**

```
data <- combine_eyes(HCL)
fix_d <- fixation_dispersion(data, participant_ID = "pNum")

# fixation data
AOI_time(data = fix_d, data_type = "fix", AOIs = HCL_AOIs, participant_ID = "pNum")

#raw data
AOI_time(data = data, data_type = "raw", AOIs = HCL_AOIs,
         sample_rate = 120, participant_ID = "pNum")
```

---

`combine_eyes`*Combine binocular data into single X/Y coordinate pairs*

---

**Description**

Combines the data from binocular samples into X/Y coordinate pairs. Two methods can be used: "average" or "best\_eye". For "average", the result is based on the average of the two eyes for each sample, or for samples where there is data from only a single eye, that eye is used. For "best\_eye", a summary of the proportion of missing samples is computed, and the eye with the fewest missing samples is used.

**Usage**

```
combine_eyes(data, method = "average")
```

**Arguments**

<code>data</code>	raw data with columns <code>time</code> , <code>left_x</code> , <code>left_y</code> , <code>right_x</code> , <code>right_y</code> , and <code>trial</code>
<code>method</code>	either "average" or "best_eye" - see description.

**Value**

a dataframe of x-2 variables (with `left_x` and `right_x` condensed to `x`, and `left_y` and `right_y` condensed to `y`) and the same number of observations as the input data

**Examples**

```
combine_eyes(HCL, method = "average")
```

---

`compare_algorithms`*A battery of metrics and plots to compare the two algorithms (dispersion and VTI)*

---

**Description**

A tool for comparing the two different algorithms present in this package. This function is useful for assessing the data as well as exploring which algorithm is likely to fit data more appropriately. The raw data is run through both algorithms (using the same specified dispersion tolerances, etc.) before making comparisons of the underlying data. Can only be used for single participant data.

**Usage**

```
compare_algorithms(
  data,
  plot_fixations = TRUE,
  print_summary = TRUE,
  sample_rate = NULL,
  threshold = 100,
  min_dur = 150,
  min_dur_sac = 20,
  disp_tol = 100,
  NA_tol = 0.25,
  run_interp = TRUE,
  smooth = FALSE
)
```

**Arguments**

<code>data</code>	A dataframe with raw data (time, x, y, trial) for one participant
<code>plot_fixations</code>	Whether to plot the detected fixations. default as TRUE
<code>print_summary</code>	Whether to print the summary table. default as TRUE
<code>sample_rate</code>	sample rate of the eye-tracker. If default of NULL, then it will be computed from the timestamp data and the number of samples. Supplied to the VTI algorithm
<code>threshold</code>	velocity threshold (degrees of VA / sec) to be used for identifying saccades. Supplied to the VTI algorithm
<code>min_dur</code>	Minimum duration (in milliseconds) of period over which fixations are assessed. Supplied to both algorithms.
<code>min_dur_sac</code>	Minimum duration (in milliseconds) for saccades to be determined. Supplied to the VTI algorithm
<code>disp_tol</code>	Maximum tolerance (in pixels) for the dispersion of values allowed over fixation period. Supplied to both algorithms
<code>NA_tol</code>	the proportion of NAs tolerated within any window of samples that is evaluated as a fixation. Supplied to the dispersion algorithm
<code>run_interp</code>	include a call to <code>eyetools::interpolate</code> on each trial. Supplied to the VTI algorithm
<code>smooth</code>	include a call to <code>eyetools::smoother</code> on each trial. Supplied to the VTI algorithm

**Value**

a list of the fixation data, correlation output, and data used for plotting

**Examples**

```
data <- combine_eyes(HCL)
compare_algorithms(data[data$pNum == 118,])
```

---

conditional\_transform *conditional\_transform*

---

### Description

A function to perform conditional transformations of the x/y raw data. The function takes the dataframe and performs a single axis flip based on the values specified in the `cond_column`. The primary use of this function is to correct or normalise the data when counterbalancing stimulus placement within experiments (e.g., having a target stimulus appear on the left and right equally often)

### Usage

```
conditional_transform(  
  data,  
  flip = c("x", "y"),  
  cond_column,  
  cond_values,  
  resolution_x = 1920,  
  resolution_y = 1080,  
  message = TRUE  
)
```

### Arguments

<code>data</code>	a dataframe that includes columns x and y and the column specified in <code>cond_column</code> . Can be raw, fixation, or saccade data.
<code>flip</code>	either "x", to flip across vertical midline, or "y" to flip across horizontal midline
<code>cond_column</code>	a column name, on which the flips are conditional
<code>cond_values</code>	a single value or vector stating which values in <code>cond_column</code> result in a flip
<code>resolution_x</code>	screen size in pixels for the x axis
<code>resolution_y</code>	screen size in pixels for the y axis
<code>message</code>	whether to output messages during function. Useful to turn off when using in a vectorised fashion where it is running multiple times

### Value

a dataframe of the equivalent format as the input data

### Examples

```
data <- combine_eyes(HCL)  
data <- merge(data, HCL_behavioural)  
conditional_transform(data, flip = "x",  
  cond_column = "cue_order",  
  cond_values = 2)
```

---

dist\_to\_visual\_angle    *Compute visual angle from distance metrics*

---

### Description

Takes a single value or vector of distances and returns the visual angle equivalent.

### Usage

```
dist_to_visual_angle(  
  vector,  
  dist_type = "cm",  
  view_dist_cm = 60,  
  screen_width_cm = 51,  
  screen_width_pixels = 1920  
)
```

### Arguments

vector	vector of distances (or single distance)
dist_type	default is "cm". Specify "pixel" for conversion from pixel values.
view_dist_cm	viewing distance in cm. Default of 60cm.
screen_width_cm	used in conversion of pixel values. Default is 51 cm (24" monitor).
screen_width_pixels	used in conversion of pixel values. Default is 1920 pixels.

### Value

an equivalent-sized object to the input

### Examples

```
# calculate visual angle for stimulus of 5cm  
dist_to_visual_angle(5)  
  
# calculate visual angle of stimuli 2 and 10cm width at 50 cm viewing angle  
dist_to_visual_angle(c(2,10), view_dist_cm = 50)  
  
# calculate visual angle of 150 pixel wide  
dist_to_visual_angle(150, dist_type = "pixels")
```

---

fixation\_dispersion     *Fixation detection using a dispersion method*

---

### Description

Detects fixations by assessing dispersion of the eye position, using a method that is similar to that proposed by Salvucci & Goldberg (1996). Evaluates the maximum dispersion (distance) between x/y coordinates across a window of data. Looks for sufficient periods in which this maximum dispersion is below the specified dispersion tolerance. NAs are considered breaks in the data and are not permitted within a valid fixation period. Runs the interpolation algorithm by default to fix small breaks in the data.

### Usage

```
fixation_dispersion(  
  data,  
  min_dur = 150,  
  disp_tol = 100,  
  run_interp = TRUE,  
  NA_tol = 0.25,  
  progress = TRUE,  
  participant_ID = "participant_ID"  
)
```

### Arguments

data	A dataframe with raw data (time, x, y, trial) for one participant (the standardised raw data form for eyetools)
min_dur	Minimum duration (in milliseconds) of period over which fixations are assessed
disp_tol	Maximum tolerance (in pixels) for the dispersion of values allowed over fixation period
run_interp	include a call to eyetools::interpolate on each trial
NA_tol	the proportion of NAs tolerated within any window of samples that is evaluated as a fixation
progress	Display a progress bar
participant_ID	the variable that determines the participant identifier. If no column present, assumes a single participant

### Details

It can take either single participant data or multiple participants where there is a variable for unique participant identification. The function looks for an identifier named `participant_ID` by default and will treat this as multiple-participant data as default, if not it is handled as single participant data, or the `participant_ID` needs to be specified

**Value**

a dataframe containing each detected fixation by trial, with mean x/y position in pixel, start and end times, and duration.

**References**

Salvucci, D. D., & Goldberg, J. H. (2000). Identifying fixations and saccades in eye-tracking protocols. *Proceedings of the Symposium on Eye Tracking Research & Applications - ETRA '00*, 71–78.

**Examples**

```
data <- combine_eyes(HCL)
fixation_dispersion(data, participant_ID = "pNum")
```

---

fixation\_VTI

*Fixation detection using a velocity threshold identification method*

---

**Description**

Determine fixations by assessing the velocity of eye-movements, using a method that is similar to that proposed by Salvucci & Goldberg (1996). Applies the algorithm used in VTI\_saccade and removes the identified saccades before assessing whether separated fixations are outside of the dispersion tolerance. If they are outside of this tolerance, the fixation is treated as a new fixation regardless of the length of saccade separating them. Compared to fixation\_dispersion(), fixation\_VTI() is more conservative in determining a fixation as smaller saccades are discounted and the resulting data is treated as a continued fixation (assuming it is within the pixel tolerance set by disp\_tol). Returns a summary of the fixations found per trial, including start and end coordinates, timing, duration, mean velocity, and peak velocity.

**Usage**

```
fixation_VTI(  
  data,  
  sample_rate = NULL,  
  threshold = 100,  
  min_dur = 150,  
  min_dur_sac = 20,  
  disp_tol = 100,  
  run_interp = TRUE,  
  smooth = FALSE,  
  progress = TRUE,  
  participant_ID = "participant_ID"  
)
```

**Arguments**

<code>data</code>	A dataframe with raw data (time, x, y, trial) for one participant
<code>sample_rate</code>	sample rate of the eye-tracker. If default of NULL, then it will be computed from the timestamp data and the number of samples
<code>threshold</code>	velocity threshold (degrees of VA / sec) to be used for identifying saccades.
<code>min_dur</code>	Minimum duration (in milliseconds) of period over which fixations are assessed
<code>min_dur_sac</code>	Minimum duration (in milliseconds) for saccades to be determined
<code>disp_tol</code>	Maximum tolerance (in pixels) for the dispersion of values allowed over fixation period
<code>run_interp</code>	include a call to <code>eyetools::interpolate</code> on each trial.
<code>smooth</code>	include a call to <code>eyetools::smoother</code> on each trial
<code>progress</code>	Display a progress bar
<code>participant_ID</code>	the variable that determines the participant identifier. If no column present, assumes a single participant

**Details**

It can take either single participant data or multiple participants where there is a variable for unique participant identification. The function looks for an identifier named `participant_ID` by default and will treat this as multiple-participant data as default, if not it is handled as single participant data, or the `participant_ID` needs to be specified

**Value**

a dataframe containing each detected fixation by trial, with mean x/y position in pixel, start and end times, and duration.

**References**

Salvucci, D. D., & Goldberg, J. H. (2000). Identifying fixations and saccades in eye-tracking protocols. *Proceedings of the Symposium on Eye Tracking Research & Applications - ETRA '00*, 71–78.

**Examples**

```
data <- combine_eyes(HCL)
fixation_VTI(data, participant_ID = "pNum")
```

---

HCL	<i>Example dataset from that contains binocular eye data from two participants from a simple contingency learning task (the data are from Beesley, Nguyen, Pearson, &amp; Le Pelley, 2015). In this task there are two stimuli that appear simultaneously on each trial (to the left and right of the screen). Participants look at these cues and then make a decision by selecting an "outcome response" button.</i>
-----	--

---

### Description

The dataset contains data from two participants and the first six trials of the study.

### Usage

HCL

### Format

A dataframe of 31,041 observations and seven variables

**pNum** participant number

**time** timestamp of the sample (milliseconds)

**left\_x** x coordinate of the left eye

**left\_y** y coordinate of the left eye

**right\_x** x coordinate of the right eye

**right\_y** y coordinate of the right eye

**trial** trial number ...

---

HCL_AOIs	<i>Example AOIs for use with HCL</i>
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---

### Description

This dataframe contains three rectangular areas of interest (AOIs), set out for use with the HCL dataset. Values are in pixels.

### Usage

HCL\_AOIs

**Format**

A data frame with 3 rows and 4 variables:

**x** centred x coordinate of the AOI

**y** centred y coordinate of the AOI

**width\_radius** either the width of the AOI, or the radius for circular AOIs

**height** the height of the AOI; should be NA for circular AOIs ...

---

HCL\_behavioural

*Example dataset of behavioural data to complement dataset HCL.*

---

**Description**

This contains information on stimuli (such as the side the predictive cue was presented on) as well as response data, including accuracy and response times

**Usage**

HCL\_behavioural

**Format**

A dataframe of 12 observations and eight variables

**pNum** participant number

**trial** trial number

**P\_cue** Are these necessary columns?

**NP\_cue** Are these necessary columns?

**cue\_order** whether the predictive cue os presented on the left (1) or the right (2)

**correct\_out** NAre these necessary columns?

**accuracy** response accuracy

**RT** response time in milliseconds ...

---

hdf5_to_csv	<i>hdf5_to_csv</i>
-------------	--------------------

---

**Description**

A function to convert TOBII-generated hdf5 files to csv

**Usage**

```
hdf5_to_csv(filename)
```

**Arguments**

filename            the hdf5 file generated from TOBII

**Value**

A list of csv files collected from the eyetracker content, if only one eyetracking event is present, return this as a csv file

**Examples**

```
## Not run:  
raw_data <- hdf5_to_csv("example_TOBII.hdf5")  
  
## End(Not run)
```

---

interpolate	<i>Interpolation of missing data (NAs)</i>
-------------	--

---

**Description**

Extends the zoo::na.approx and zoo::na.spline functions to include a report which provides the proportion of missing data before and after the interpolation process. This is handy for evaluating the effectiveness of the repair.

**Usage**

```
interpolate(  
  data,  
  maxgap = 25,  
  method = "approx",  
  report = FALSE,  
  participant_ID = "participant_ID"  
)
```

**Arguments**

data	dataframe with columns time, x, y, trial (the standardised raw data form for eyeproc)
maxgap	maximum number of consecutive NAs to fill. Any longer gaps will be left unchanged (see zoo package)
method	"approx" for linear interpolation or "spline" for cubic spline interpolation
report	default is FALSE. If TRUE, then the return value is a list containing the returned data frame and the report.
participant_ID	the variable that determines the participant identifier. If no column present, assumes a single participant

**Details**

It can take either single participant data or multiple participants where there is a variable for unique participant identification. The function looks for an identifier named participant\_ID by default and will treat this as multiple-participant data as default, if not it is handled as single participant data, or the participant\_ID needs to be specified

**Value**

a dataframe of the same shape of the input data

**Examples**

```
data <- combine_eyes(HCL)
interpolate(data, maxgap = 20, participant_ID = "pNum")
```

---

plot\_seq

*Plot of raw data over time*

---

**Description**

A tool for visualising the timecourse of raw data over a single trial. If data from multiple trials are present, then a single trial will be sampled at random. Alternatively, the trial\_number can be specified. Data can be plotted across the whole trial, or can be split into bins to present distinct plots for each time window.

**Usage**

```
plot_seq(
  data = NULL,
  trial_number = NULL,
  AOIs = NULL,
  bg_image = NULL,
  res = c(0, 1920, 0, 1080),
```

```

    flip_y = FALSE,
    plot_header = FALSE,
    bin_time = NULL,
    bin_range = NULL
  )

```

### Arguments

data	A dataframe with raw data. If multiple trials are used, then one trial is sampled at random.
trial_number	can be used to select a particular trial within the data
AOIs	A dataframe of areas of interest (AOIs), with one row per AOI (x, y, width_radius, height).
bg_image	The filepath of an image to be added to the plot, for example to show a screenshot of the task.
res	resolution of the display to be shown, as a vector (xmin, xmax, ymin, ymax)
flip_y	reverse the y axis coordinates (useful if origin is top of the screen)
plot_header	display the header title text which explains graphical features of the plot.
bin_time	if wanting to split data into bins, the time (in ms) for each bin of data to be displayed
bin_range	if wanting to split data into bins, the first and last bin to be display, e.g., c(1,5)

### Value

a plot of the raw data representing changes over time

### Examples

```

data <- combine_eyes(HCL)

# plot the raw data
plot_seq(data = data[data$PNum == 118,])

# with AOIs
plot_seq(data = data[data$PNum == 118,], AOIs = HCL_AOIs)

# plot raw data with bins
plot_seq(data = data[data$PNum == 118,], bin_time = 500)

```

---

plot_spatial	<i>Plot raw data and fixations</i>
--------------	------------------------------------

---

### Description

A tool for visualising raw eye-data, processed fixations, and saccades. Can use all three data types together and independently. Fixations can be labeled in the order they were made. Can overlay areas of interest (AOIs) and customise the resolution.

### Usage

```
plot_spatial(
  raw_data = NULL,
  fix_data = NULL,
  sac_data = NULL,
  AOIs = NULL,
  trial_number = NULL,
  bg_image = NULL,
  res = c(0, 1920, 0, 1080),
  flip_y = FALSE,
  show_fix_order = TRUE,
  plot_header = FALSE
)
```

### Arguments

raw_data	data in standard raw data form (time, x, y, trial)
fix_data	data output from fixation function
sac_data	data output from saccade function
AOIs	A dataframe of areas of interest (AOIs), with one row per AOI (x, y, width_radius, height). If using circular AOIs, then the 3rd column is used for the radius and the height should be set to NA.
trial_number	can be used to select particular trials within the data
bg_image	The filepath of an image to be added to the plot, for example to show a screenshot of the task.
res	resolution of the display to be shown, as a vector (xmin, xmax, ymin, ymax)
flip_y	reverse the y axis coordinates (useful if origin is top of the screen)
show_fix_order	label the fixations in the order they were made
plot_header	display the header title text which explains graphical features of the plot.

### Value

a plot of the raw data

**Examples**

```

data <- combine_eyes(HCL)
data <- data[data$Num == 118,]
# plot the raw data
plot_spatial(raw_data = data[data$Num == 118,])

# plot both raw and fixation data together
plot_spatial(raw_data = data, fix_data = fixation_dispersion(data))

#plot one trial
plot_spatial(raw_data = data, fix_data = fixation_dispersion(data), trial_number = 1)

```

---

saccade\_VTI

*Velocity threshold identification of saccades*


---

**Description**

Use the velocity threshold algorithm from Salvucci & Goldberg (1996) to determine saccadic eye movements. Returns a summary of the saccades found per trial, including start and end coordinates, timing, duration, mean velocity, and peak velocity.

**Usage**

```

saccade_VTI(
  data,
  sample_rate = NULL,
  threshold = 150,
  min_dur = 20,
  participant_ID = "participant_ID"
)

```

**Arguments**

data	A dataframe with raw data (time, x, y, trial) for one participant
sample_rate	sample rate of the eye-tracker. If default of NULL, then it will be computed from the timestamp data and the number of samples
threshold	velocity threshold (degrees of VA / sec) to be used for identifying saccades
min_dur	minimum duration (ms) expected for saccades. This helps to avoid identification of very short saccades occurring at the boundary of velocity threshold
participant_ID	the variable that determines the participant identifier. If no column present, assumes a single participant

**Details**

It can take either single participant data or multiple participants where there is a variable for unique participant identification. The function looks for an identifier named `participant_ID` by default and will treat this as multiple-participant data as default, if not it is handled as single participant data, or the `participant_ID` needs to be specified

**Value**

a data frame giving the saccades found by trial

**Examples**

```
data <- combine_eyes(HCL)
saccade_VTI(data, participant_ID = "pNum")
```

---

 smoother

*Smoothing of raw data*


---

**Description**

A wrapper for the `stats::loess` function, with default parameters suitable for smoothing raw eye data

**Usage**

```
smoother(data, span = 0.1, plot = FALSE, participant_ID = "participant_ID")
```

**Arguments**

<code>data</code>	A dataframe with raw data (time, x, y, trial) for one participant
<code>span</code>	From <code>stats::loess</code> . The parameter alpha which controls the degree of smoothing.
<code>plot</code>	whether to plot the raw and smoothed plot for inspection
<code>participant_ID</code>	the variable that determines the participant identifier. If no column present, assumes a single participant

**Details**

It can take either single participant data or multiple participants where there is a variable for unique participant identification. The function looks for an identifier named `participant_ID` by default and will treat this as multiple-participant data as default, if not it is handled as single participant data, or the `participant_ID` needs to be specified

**Value**

a dataframe of the same shape as the input data

**Examples**

```
data <- combine_eyes(HCL)

smoother(data, participant_ID = "pNum")

#with an inspection plot
smoother(data, span = .02, participant_ID = "pNum", plot = TRUE)
```

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