

# Package: SpaTopic (via r-universe)

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**Type** Package

**Title** Topic Inference to Identify Tissue Architecture in Multiplexed Images

**Version** 1.1.0

**Date** 2024-04-22

**Description** A novel spatial topic model to integrate both cell type and spatial information to identify the complex spatial tissue architecture on multiplexed tissue images without human intervention. The Package implements a collapsed Gibbs sampling algorithm for inference. 'SpaTopic' is scalable to large-scale image datasets without extracting neighborhood information for every single cell. For more details on the methodology, see <https://xiyupeng.github.io/SpaTopic/>.

**License** GPL (>= 3)

**Depends** R (>= 3.5.0),

**Imports** Rcpp (>= 0.12.0), RANN (>= 2.6.0), sf (>= 1.0-12), methods (>= 3.4), foreach (>= 1.5.0), iterators (>= 1.0),

**LinkingTo** Rcpp, RcppArmadillo, RcppProgress,

**Suggests** knitr, rmarkdown, SeuratObject (>= 4.9.9.9086), doParallel (>= 1.0),

**VignetteBuilder** knitr

**RoxygenNote** 7.2.3

**Encoding** UTF-8

**LazyData** true

**URL** <https://github.com/xiyupeng/SpaTopic>

**BugReports** <https://github.com/xiyupeng/SpaTopic/issues>

**NeedsCompilation** yes

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**Repository** CRAN

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## Contents

gibbs.res-class . . . . .	2
lung5 . . . . .	3
Seurat5obj_to_SpaTopic . . . . .	3
SpaTopic-Package . . . . .	4
SpaTopic_inference . . . . .	5
stratified_sampling_sf . . . . .	7
<b>Index</b>	<b>9</b>

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gibbs.res-class	<i>A class of the output from 'SpaTopic'</i>
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## Description

Outputs from function [SpaTopic\\_inference](#). A `list` contains the following members:

- `$Perplexity`. The perplexity is for the training data. Let  $N$  be the total number of cells across all images.  $Perplexity = \exp(-\loglikelihood/N)$
- `$Deviance`.  $Deviance = -2\loglikelihood$ .
- `$loglikelihood`. The model log-likelihood.
- `$loglike.trace`. The log-likelihood for every collected posterior sample. NULL if `trace = FALSE`.
- `$Beta`. Topic content matrix with rows as celltypes and columns as topics
- `$Theta`. Topic prevalent matrix with rows as regions and columns as topics
- `$Ndk`. Number of cells per topic (col) per region (row).
- `$Nwk`. Number of cells per topic (col) per celltype (row).
- `$Z.trace`. Number of times cell being assigned to each topic across all posterior samples. We can further compute the posterior distributions of  $Z$  (topic assignment) for individual cells.
- `$doc.trace`. `Ndk` for every collected posterior sample. NULL if `trace = FALSE`.
- `$word.trace`. `Nwk` for every collected posterior sample. NULL if `trace = FALSE`.

## See Also

[SpaTopic\\_inference](#)

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lung5	<i>Example input data for 'SpaTopic'</i>
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**Description**

multiplexed image data on tumor tissue sample from non small cell lung cancer patient

**Usage**

```
lung5
```

**Format**

```
## 'lung5' A data frame with 100149 rows and 4 columns:
```

**image** Image ID

**X** X coordinate of the cell

**Y** Y coordinate of the cell

**type** cell type

**Source**

<<https://nanosttring.com/products/cosmx-spatial-molecular-imager/ffpe-dataset/nsclc-ffpe-dataset/>>

**See Also**

[SpaTopic\\_inference](#), [Seurat5obj\\_to\\_SpaTopic](#)

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Seurat5obj\_to\_SpaTopic

*Convert a Seurat v5 object as the input of 'SpaTopic'*

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**Description**

Prepare 'SpaTopic' input from one Seurat v5 object

**Usage**

```
Seurat5obj_to_SpaTopic(object, group.by, image = "image1")
```

**Arguments**

**object** Seurat v5 object

**group.by** character. The name of the column that contains celltype information in the Seurat object.

**image** character. The name of the image. Default is "image1".

**Value**

Return a data frame as the input of 'SpaTopic'

**See Also**

[lung5](#)

**Examples**

```
## nano.obj is a Seurat v5 object
#dataset<-Seurat5obj_to_SpaTopic(object = nano.obj,
#                                group.by = "predicted.annotation.l1",image = "image1")
## Expect output
data("lung5")
```

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SpaTopic-Package      *'SpaTopic' R package*

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**Description**

The 'SpaTopic' R package is centered around the 'SpaTopic' algorithm to infer the spatial tissue architectures from multiplexed images.

**Details**

The package implements a Collapsed Gibbs sampling algorithm to infer topics, corresponding to distinct tissue microenvironments across multiple tissue images. Without obtaining the cell neighborhood info of every single cell, 'SpaTopic' runs much faster than other KNN-based methods on large-scale images.

The main functions in the 'SpaTopic' package

- Prepare input [Seurat5obj\\_to\\_SpaTopic](#)
- Model Inference [SpaTopic\\_inference](#)

**Author(s)**

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SpaTopic_inference	<i>'SpaTopic': fast topic inference to identify tissue architecture in multiplexed images</i>
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### Description

This is the main function of 'SpaTopic', implementing a Collapsed Gibbs Sampling algorithm to learn topics, which referred to different tissue microenvironments, across multiple multiplexed tissue images. The function takes cell labels and coordinates on tissue images as input, and returns the inferred topic labels for every cell, as well as topic contents, a distribution over celltypes. The function recovers spatial tissue architectures across images, as well as indicating cell-cell interactions in each domain.

### Usage

```
SpaTopic_inference(
  tissue,
  ntopics,
  sigma = 50,
  region_radius = 400,
  kneigh = 5,
  npoints_selected = 1,
  ini_LDA = TRUE,
  ninit = 10,
  niter_init = 100,
  beta = 0.05,
  alpha = 0.01,
  trace = FALSE,
  seed = 123,
  thin = 20,
  burnin = 1000,
  niter = 200,
  display_progress = TRUE,
  do.parallel = FALSE,
  n.cores = 1,
  axis = "2D"
)
```

### Arguments

tissue	(Required). A data frame or a list of data frames. One for each image. Each row represent a cell with its image ID, X, Y coordinates on the image, celltype, with column names (image, X, Y, type), respectively. You may add another column Y2 for 3D tissue image.
ntopics	(Required). Number of topics. Topics will be obtained as distributions of cell types.

<code>sigma</code>	Default is 50. The lengthscale of the Nearest-neighbor Exponential Kernel. Sigma controls the strength of decay of correlation with distance in the kernel function. Please check the paper for more information. Need to be adjusted based on the image resolution
<code>region_radius</code>	Default is 400. The radius for each grid square when sampling region centers for each image. Need to be adjusted based on the image resolution and pattern complexity.
<code>kneigh</code>	Default is 5. Only consider the top 5 closest region centers for each cell.
<code>npoints_selected</code>	Default is 1. Number of points sampled for each grid square when sampling region centers for each image. Used with <code>region_radius</code> .
<code>ini_LDA</code>	Default is TRUE. Use warm start strategy for initialization and choose the best one to continue. If 0, it simply uses the first initialization.
<code>ninit</code>	Default is 10. Number of initialization. Only retain the initialization with the highest log likelihood (perplexity).
<code>niter_init</code>	Default is 100. Warm start with 100 iterations in the Gibbs sampling during initialization.
<code>beta</code>	Default is 0.05. A hyperparameter to control the sparsity of topic content (topic-celltype) matrix Beta. A smaller value introduces more sparse in Beta.
<code>alpha</code>	Default is 0.01. A hyperparameter to control the sparsity of document (region) content (region-topic) matrix Theta. For our application, we keep it very small for the sparsity in Theta.
<code>trace</code>	Default is FALSE. Compute and save log likelihood, Ndk, Nwk for every posterior samples. Useful when you want to use DIC to select number of topics, but it is time consuming to compute the likelihood for every posterior samples.
<code>seed</code>	Default is 123. Random seed.
<code>thin</code>	Default is 20. Key parameter in Gibbs sampling. Collect a posterior sample for every thin=20 iterations.
<code>burnin</code>	Default is 1000. Key parameter in Gibbs sampling. Start to collect posterior samples after 1000 iterations. You may increase the number of iterations for burn-in for highly complex tissue images.
<code>niter</code>	Default is 200. Key parameter in Gibbs sampling. Number of posterior samples collected for model inference.
<code>display_progress</code>	Default is TRUE. Display the progress bar.
<code>do.parallel</code>	Default is FALSE. Use parallel computing through R package foreach.
<code>n.cores</code>	Default is 1. Number of cores used in parallel computing.
<code>axis</code>	Default is "2D". You may switch to "3D" for 3D tissue images. However, the model inference for 3D tissue is still under test.

### Value

Return a `gibbs.res-class` object. A list of outputs from Gibbs sampling.

**See Also**[gibbs.res-class](#)**Examples**

```
## tissue is a data frame containing cellular information from one image or
## multiple data frames from multiple images.

data("lung5")
## NOT RUN, it takes about 90s
library(sf)
#gibbs.res<-SpaTopic_inference(lung5, ntopics = 7,
#                               sigma = 50, region_radius = 400)

## generate a fake image 2 and make an example for multiple images
## NOT RUN
#lung6<-lung5
#lung6$image<-"image2" ## The image ID of two images should be different
#gibbs.res<-SpaTopic_inference(list(A = lung5, B = lung6),
#                                  ntopics = 7, sigma = 50, region_radius = 400)
```

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stratified\_sampling\_sf

*Spatially stratified random sample points from an image.*


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**Description**

Spatially stratified random sample points from an image by R package sf

**Usage**

```
stratified_sampling_sf(
  points,
  cellsize = c(600, 600),
  num_samples_per_stratum = 1
)
```

**Arguments**

`points` a data frame contains all points in a image with X, Y coordinates.  
`cellsize` a vector of length 2 contains the size of each grid square. Default c(600,600).  
`num_samples_per_stratum` number of point selected from each grid square. Default 1.

**Value**

Return a vector contains index of sampled points.

**Examples**

```
data("lung5")  
pt_idx<-stratified_sampling_sf(lung5, cellsize = c(600,600))
```



# Index

\* **datasets**

lung5, [3](#)

\* **package**

SpaTopic-Package, [4](#)

`gibbs.res-class`, [2](#)

`list`, [2](#)

lung5, [3](#), [4](#)

`Seurat5obj_to_SpaTopic`, [3](#), [3](#), [4](#)

SpaTopic-Package, [4](#)

`SpaTopic_inference`, [2-4](#), [5](#)

`stratified_sampling_sf`, [7](#)