Package: RRPP (via r-universe)

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Title Linear Model Evaluation with Randomized Residuals in a Permutation Procedure

Version 2.2.6

Description Linear model calculations are made for many random versions of data. Using residual randomization in a permutation procedure, sums of squares are calculated over many permutations to generate empirical probability distributions for evaluating model effects. Additionally, coefficients, statistics, fitted values, and residuals generated over many permutations can be used for various procedures including pairwise tests, prediction, classification, and model comparison. This package should provide most tools one could need for the analysis of high-dimensional data, especially in ecology and evolutionary biology, but certainly other fields, as well.

Depends R (>= 4.4.0)

License GPL (>= 3)

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`add.trajectories` *Plot Function for RRPP*

Description

Function adds trajectories to a principal component plot

Usage

```
add.trajectories(
  TP,
  traj.pch = 21,
  traj.col = 1,
  traj.lty = 1,
  traj.lwd = 1,
  traj.cex = 1.5,
  traj.bg = 1,
  start.bg = 3,
  end.bg = 2
)
```

Arguments

TP	plot object (from plot.trajectory.analysis)
traj.pch	Plotting "character" for trajectory points. Can be a single value or vector of length equal to the number of trajectories. See par and its description for pch.
traj.col	The color of trajectory lines. Can be a single value or vector of length equal to the number of trajectories. See par and its description for col.
traj.lty	Trajectory line type. Can be a single value or vector of length equal to the number of trajectories. See par and its description for lty.
traj.lwd	Trajectory line width. Can be a single value or vector of length equal to the number of trajectories. See par and its description for lwd.
traj.cex	Trajectory point character expansion. Can be a single value or vector of length equal to the number of trajectories. See par and its description for cex.
traj.bg	Trajectory point background. Can be a single value or vector of length equal to the number of trajectories. See par and its description for bg.
start.bg	Trajectory point background, just the start points. Can be a single value or vector of length equal to the number of trajectories. See par and its description for bg. Green start points are the default.
end.bg	Trajectory point background, just the end points. Can be a single value or vector of length equal to the number of trajectories. See par and its description for bg. Red end points are the default.

Details

The function adds trajectories to a plot made by plot.trajectory.analysis. This function has a restricted set of plot parameters based on the number of trajectories to be added to the plot.

Author(s)

Michael Collyer

References

Adams, D. C., and M. M. Cerney. 2007. Quantifying biomechanical motion using Procrustes motion analysis. J. Biomech. 40:437-444.

Adams, D. C., and M. L. Collyer. 2007. The analysis of character divergence along environmental gradients and other covariates. Evolution 61:510-515.

Adams, D. C., and M. L. Collyer. 2009. A general framework for the analysis of phenotypic trajectories in evolutionary studies. Evolution 63:1143-1154.

Collyer, M. L., and D. C. Adams. 2007. Analysis of two-state multivariate phenotypic change in ecological studies. Ecology 88:683-692.

Collyer, M. L., and D. C. Adams. 2013. Phenotypic trajectory analysis: comparison of shape change patterns in evolution and ecology. Hystrix 24: 75-83.

Collyer, M.L., D.J. Sekora, and D.C. Adams. 2015. A method for analysis of phenotypic change for phenotypes described by high-dimensional data. Heredity. 115:357-365.

See Also

plot.default and par

---

add.tree *Plot tool to add phylogenetic trees to ordination plots*

Description

Function adds a tree based on a description of edges from a class phylo object to an existing plot made from an ordinate object.

Usage

```
add.tree(
  OP,
  tree,
  edge.col = 1,
  edge.lty = 1,
  edge.lwd = 1,
  anc.pts = FALSE,
  return.ancs = FALSE,
  ...
)
```

Arguments

OP	An object with class plot.ordinate.
tree	An object of class phylo.
edge.col	A single value or vector equal to the number of edges for edge colors.
edge.lty	A single value or vector equal to the number of edges for edge line type
edge.lwd	A single value or vector equal to the number of edges for edge line weight.
anc.pts	A logical value for whether to add points for ancestral values.
return.ancs	A logical value for whether ancestral values should be printed.
...	Arguments passed onto points, used only for ancestral points.

Details

With some ordinate plots, it might be desirable to add a tree connecting points in a prescribed way, which would be tedious using points or lines. This function will project a tree from an object of class phylo into a plot with class, plot.ordinate. Using an edges matrix from a phylo object, this function will systematically connect plot points with lines that pass through estimated ancestral character points in the same plot space. Ancestral states are estimated assuming a Brownian motion model of evolutionary divergence.

Author(s)

Michael Collyer

See Also

lines and points

Examples

```
# Examples use residuals from a regression of salamander morphological
# traits against body size (snout to vent length, SVL).
# Observations are species means and a phylogenetic covariance matrix
# describes the relatedness among observations.

data("PlethMorph")
Y <- as.data.frame(PlethMorph[c("TailLength", "HeadLength",
"Snout.eye", "BodyWidth",
"Forelimb", "Hindlimb")])
Y <- as.matrix(Y)
R <- lm.rrpp(Y ~ SVL, data = PlethMorph,
iter = 0, print.progress = FALSE)$LM$residuals

PCA <- ordinate(R, scale. = TRUE)
pc.plot <- plot(PCA, pch = 19, col = "blue")

add.tree(pc.plot, tree = PlethMorph$tree, anc.pts = TRUE,
pch = 19, cex = 0.5, col = "red")
```

---

anova.lm.rrpp *ANOVA for lm.rrpp model fits*

Description

Computes an analysis of variance (ANOVA) table using distributions of random statistics from lm.rrpp. ANOVA can be performed on one model or multiple models. If the latter, the first model is considered a null model for comparison to other models. The ANOVA is functionally similar to a non-parametric likelihood ratio test for all null-full model comparisons Residuals from the null model will be used to generate random pseudo-values via RRPP for evaluation of subsequent models. The permutation schedule from the null model will be used for random permutations. This function does not correct for improper null models. One must assure that the null model is nested within the other models. Illogical results can be generated if this is not the case.

Usage

```
## S3 method for class 'lm.rrpp'
anova(
  object,
```

```
  ...,
  effect.type = c("F", "cohenf", "SS", "MS", "Rsq", "LR"),
  error = NULL,
  print.progress = TRUE
)
```

Arguments

object	Object from lm.rrpp
...	Additional lm.rrpp model fits or other arguments passed to anova.
effect.type	One of "F", "cohenf", "SS", "MS", "Rsq" to choose from which distribution of statistics to calculate effect sizes (Z). "LR" can be chosen to include a log-likelihood ratio, for multi-model comparisons. If "LR" is used, the effect type is automatically "F". See lm.rrpp.
error	An optional character string to define MS error term for calculation of F values. See lm.rrpp for examples.
print.progress	A logical argument if multiple models are used and one wishes to view progress for sums of squares (SS) calculations.

Author(s)

Michael Collyer

Examples

```
## Not run:
# See examples for lm.rrpp to see how anova.lm.rrpp works in conjunction
# with other functions

data(Pupfish)
names(Pupfish)
Pupfish$logSize <- log(Pupfish$CS) # better to not have functions in formulas

# Single-Model ANOVA

fit <- lm.rrpp(coords ~ logSize + Sex*Pop, SS.type = "I",
data = Pupfish, print.progress = FALSE, iter = 999)
anova(fit)
anova(fit, effect.type = "MS")
anova(fit, effect.type = "Rsq")
anova(fit, effect.type = "cohenf")

# Multi-Model ANOVA (like a Likelihood Ratio Test)
fit.size <- lm.rrpp(coords ~ logSize, SS.type = "I", data = Pupfish,
print.progress = FALSE, iter = 999)
fit.sex <- lm.rrpp(coords ~ logSize + Sex, SS.type = "I", data = Pupfish,
print.progress = FALSE, iter = 999)
fit.pop <- lm.rrpp(coords ~ logSize + Pop, SS.type = "I", data = Pupfish,
print.progress = FALSE, iter = 999)
anova(fit.size, fit.sex, fit.pop,
```

```
print.progress = FALSE) # compares two models to the first

# see lm.rrpp examples for mixed model ANOVA example and how to vary SS type

## End(Not run)
```

`anova.measurement.error` *ANOVA for lm.rrpp model fits used in measurement.error*

Description

Computes an analysis of variance (ANOVA) table using distributions of random statistics from `lm.rrpp`. This function is the same as `anova.lm.rrpp` but includes statistics specific to `measurement.error`, and with restrictions on how P-values and effect.sizes are calculated.

Usage

```
## S3 method for class 'measurement.error'
anova(object, ...)
```

Arguments

`object`	Object from `lm.rrpp`
`...`	Additional lm.rrpp model fits or other arguments passed to anova.

Author(s)

Michael Collyer

Examples

```
# See measurement.error help file examples for use.
```

`betaTest` *Test of beta parameters for lm.rrpp model fits*

Description

For any `lm.rrpp` object, a vector of coefficients can be used for a specific test of a vector of betas (specific population parameters). This test follows the form of (b - beta) in the numerator of a t-statistic, where beta can be a value other than 0 (or 0). However, for this test, a vector (Beta) of length, p, is used for the p variables in the `lm.rrpp` fit. If Beta is a vector of 0s, this test is essentially the same as the test performed for `coef.lm.rrpp`. However, it is possible to test null hypotheses for beta values other than 0, sensu Cicuéndez et al. (2023).

This function can use either the square-root of the inner-product of vectors of coefficients (distance, d) or generalized inner-product based on the inverse of the residual covariance matrix (Mahalanobis distance, md) as statistics. In most cases, either will likely yield similar (or same) P-values. However, Mahalanobis distance might be preferred for generalized least squares fits, which do not have consistent residual covariance matrices for null (intercept only) models over RRPP permutations (the distances are thus standardized by the residual covariances). If high-dimensional data are analyzed, a generalized inverse of the residual covariance matrix will be used because of singular covariance matrices. Results are less trustworthy with Mahalanobis distances, in these cases.

The coefficient number should be provided for specific tests. One can determine this with, e.g., coef(fit). If it is not provided (NULL), tests will be performed on all possible vectors of coefficients (rows of coefficients matrix). These tests will be performed sequentially. If a null model is not specified, then for each vector of coefficients, the corresponding parameter is dropped from the linear model design matrix to make a null model. This process is analogous in some ways to a leave-one-out cross-validation (LOOCV) analysis, testing each coefficient against models containing parameters for all other coefficients. For example, for a linear model fit, y ~ x1 + x2 + 1, where x1 and x2 are single-parameter covariates, the analysis would first drop the intercept, then x1, then x2, performing three sequential analyses. This option could require large amounts of computation time for large models, high-dimensional data, many RRPP permutations, or any combination of these. The test results previously reported via coef.lm.rrpp can be found using X.null. One would have to be cognizant of the null model used for each coefficient, based on which term it represents. The function, `reveal.model.designs` could help determine terms to include in a null model. Regardless, such tests have to be performed iteratively now, but do not require verbose results for initial lm.rrpp fits.

Difference between coef.lm.rrpp test and betaTest:

The test for coef.lm.rrpp uses the square-root of inner-products of vectors (d) as a test statistic and only tests the null hypothesis that the length of the vector is 0. The significance of the test is based on random values produced by RRPP, based on the matrices of coefficients that are produced in all permutations. The null models for generating RRPP distributions are consistent with those used for ANOVA, as specified in the `lm.rrpp` fit by choice of SS type. Therefore, the random coefficients are consistent with those produced by RRPP for generating random estimates used in ANOVA.

The betaTest analysis allows different null hypotheses to be used (vector length is not necessarily 0) and unless otherwise specified, uses a null model that lacks one vector of parameters and a full model that contains all vectors of parameters, for the parameter for which coefficients are

estimated. This is closest to a type III SS method of estimation, but each parameter is dropped from the model, rather than terms potentially comprising several parameters. Additionally, betaTest calculates Mahalanobis distance, in addition to Euclidean distance, for vectors of coefficients. This statistic is probably better for more types of models (like generalized least squares fits).

High-dimensional data:

If data are high-dimensional (more variables than observations), or even highly multivariate, using Mahalanobis distance can require significant computation time and will require using a generalized inverse. One might wish to consider first whether using principal component scores or other ordinate scores could achieve the same goal. (See `ordinate`.) For example, one could use the first few principal components as a surrogate for a high-dimensional trait, and test whether the surrogate trait is different than Beta. This would require that the PC scores make sense compared to the original variables, but it would be more computationally tractable.

Generalized Least Squares:

To the extent that is possible, tests for GLS estimated coefficients should use Mahalanobis distance. The reason is that the covariance matrix for the data (not to be confused with the residual covariance matrix of a linear model) might not be consistent across RRPP permutations. To assure that random distances are comparable in terms of scale, a generalized (Mahalanobis) distance is safer. However, this can impose a computational burden for high-dimensional data (see above).

Usage

```
betaTest(
  fit,
  X.null = NULL,
  include.md = FALSE,
  coef.no = NULL,
  Beta = NULL,
  print.progress = FALSE
)
```

Arguments

`fit`	Object from `lm.rrpp`
`X.null`	Optional object that is either a linear model design matrix or a model fit from `lm.rrpp`, from which a linear model design matrix can be extracted. Note that if any transformation of a design matrix is required (GLS estimation), it is assumed that the matrix was transformed prior to analysis. If X.null is a `lm.rrpp` object, transformation is inherent.
`include.md`	A logical vector for whether to include Mahalanobis distances in the results. For highly multivariate data, this will slow down computations, significantly.
`coef.no`	The row or rows of a matrix of coefficients for which to perform the test. This can be learned by performing coef(fit), prior to the test. If left NULL, the analysis will cycle through every possible vector of coefficients (rows of a coefficients matrix).

Beta	A single value (for univariate data) or a numeric vector with length equal to the number of variables used in the fit object. If left NULL, 0 is used for each parameter. This should not be a matrix. If one wishes to use different Beta vectors for different coefficients, then multiple tests should be performed. (Because tests are performed sequentially, multiple tests using the same Beta vector produces results that are the same as for multiple rows of coefficients, using the same Beta vector.)
print.progress	A logical value for whether to print test progress to the screen. This might be useful if a large number of coefficient vectors are tested, so that one can track completion.

Value

Function returns a list with the following components:

obs.d	Length of observed b - Beta vector
obs.md	The observed b - Beta vector length, after accounting for residual covariance matrix; the Mahalanobis distance
Beta	Hypothesized beta values in the Beta vector.
obs.B.mat	The observed matrix of coefficients (before subtracting Beta).
coef.no	The rows of the observed matrix of coefficients, for which to subtract Beta.
random.stats	Random distances produced with RRPP.

Author(s)

Michael Collyer

References

Tejero-Cicuéndez, H., I. Menéndez, A. Talavera, G. Riaño, B. Burriel-Carranza, M. Simó-Riudalbas, S. Carranza, and D.C. Adams. 2023. Evolution along allometric lines of least resistance: Morphological differentiation in Pristurus geckos. Evolution. 77:2547–2560.

See Also

coef.lm.rrpp

Examples

```
## Not run:
data(PlethMorph)
fit <- lm.rrpp(TailLength ~ SVL,
data = PlethMorph,
verbose = TRUE)

## Allometry test (Beta = 0)

T1 <- betaTest(fit, coef.no = 2, Beta = 0)
summary(T1)
```

```
# Including Mahalanobis distance

T1 <- betaTest(fit, coef.no = 2,
Beta = 0, include.md = TRUE)
summary(T1)


# compare to
coef(fit, test = TRUE)

# Note that if Beta is not provided

T1 <- betaTest(fit, coef.no = 2)
summary(T1)

# Note that if coef.no is not provided

T1 <- betaTest(fit)
summary(T1)

# Note that if X.null is provided

T1 <- betaTest(fit, X.null = model.matrix(fit)[, 1],
coef.no = 2)
summary(T1)


## Isometry test (Beta = 1)
# Failure to reject H0 suggests isometric-like association.

T2 <- betaTest(fit, coef.no = 2, Beta = 1)
summary(T2)


## More complex tests

# Multiple covariates

fit2 <- lm.rrpp(HeadLength ~ SVL + TailLength,
data = PlethMorph,
SS.type = "II",
verbose = TRUE)

fit.null1 <- lm.rrpp(HeadLength ~ SVL,
data = PlethMorph,
verbose = TRUE)

fit.null2 <- lm.rrpp(HeadLength ~ TailLength,
data = PlethMorph,
verbose = TRUE)

## allometries
```

```
T3 <- betaTest(fit2, fit.null2, coef.no = 2, Beta = 0)
T4 <- betaTest(fit2, fit.null1, coef.no = 3, Beta = 0)
summary(T3)
summary(T4)

# compare to
coef(fit2, test = TRUE)

## isometries
T5 <- betaTest(fit2, fit.null2, coef.no = 2, Beta = 1)
T6 <- betaTest(fit2, fit.null1, coef.no = 3, Beta = 1)

summary(T5)
summary(T6)

# Intercept test
T7 <- betaTest(fit2, fit.null1, coef.no = 1)
summary(T7)

# multivariate data

PlethMorph$Y <- cbind(PlethMorph$HeadLength, PlethMorph$TailLength)
fit3 <- lm.rrpp(Y ~ SVL,
data = PlethMorph,
verbose = TRUE)

T8 <- betaTest(fit3, coef.no = 2, Beta = c(0, 0))
T9 <- betaTest(fit3, coef.no = 2, Beta = c(1, 1))

summary(T8)
summary(T9)

## GLS example

fit4 <- lm.rrpp(TailLength ~ SVL,
data = PlethMorph,
Cov = PlethMorph$PhyCov,
verbose = TRUE)

T10 <- betaTest(fit4, include.md = TRUE)

summary(T10)

# compare to
coef(fit4, test = TRUE)

anova(fit4)


## End(Not run)
```

classify *Deprecated functions in RRPP*

Description

The following function has been deprecated in RRPP

Usage

```
classify()
```

Details

This function has been deprecated. Use `prep.lda` instead.

coef.lm.rrpp *coef for lm.rrpp model fits*

Description

Computes ordinary or generalized least squares coefficients over the permutations of an `lm.rrpp` model fit with predefined random permutations. For each coefficient vector, the Euclidean distance is calculated as an estimate of the amount of change in Y, the n x p matrix of dependent variables; larger distances mean more change in location in the data space associated with a one unit change in the model design, for the parameter described. Random coefficients are based on either RRPP or FRPP, as defined by the `lm.rrpp` model fit.

The test argument will be deprecated:

The following details will also be removed:

This function can be used to test the specific coefficients of an lm.rrpp fit. The test statistics are the distances (d), which are also standardized (Z-scores). The Z-scores might be easier to compare, as the expected values for random distances can vary among coefficient vectors.

If RRPP is used, all distributions of coefficient vector distances are based on appropriate null models, as defined by SS type. Please be aware that this can result in two seemingly strange but reasonable phenomena. First, if type II or type III SS is used, the intercept will not appear in test results (because the function seeks model parameter differences to know for which coefficients to calculate Euclidean distances). Even if it appears for type I SS, this is merely an artifact of sequential model building and there really is no meaningful test of intercept = 0. Second, Euclidean distances might not always be logical, especially when viewing univariate coefficients, in which case the expected d is |b|. Coefficients without a test are based on the full model; tests are based on the estimates of coefficients (b), given a null model. For example, for a model, y ~ b1 + b2 + b3, with type I SS, b2 will be estimated and tested, using a null model, y ~ b1 and a full model, y ~ b1 + b2. The estimate for b2 might not be the same in the test as when estimated from the model, y ~ b1 + b2 + b3. Therefore, the d statistic might not reflect what one would expect from the full model (like when using type III SS).

Difference between coef.lm.rrpp test and betaTest:

The test for coef.lm.rrpp uses the square-root of inner-products of vectors (d) as a test statistic and only tests the null hypothesis that the length of the vector is 0. The significance of the test is based on random values produced by RRPP, based on the matrices of coefficients that are produced in all permutations. The null models for generating RRPP distributions are consistent with those used for ANOVA, as specified in the `lm.rrpp` fit by choice of SS type. Therefore, the random coefficients are consistent with those produced by RRPP for generating random estimates used in ANOVA.

The betaTest analysis allows different null hypotheses to be used (vector length is not necessarily 0) and unless otherwise specified, uses a null model that lacks one vector of parameters and a full model that contains all vectors of parameters, for the parameter for which coefficients are estimated. This is closest to a type III SS method of estimation, but each parameter is dropped from the model, rather than terms potentially comprising several parameters. Additionally, betaTest calculates Mahalanobis distance, in addition to Euclidean distance, for vectors of coefficients. This statistic is probably better for more types of models (like generalized least squares fits).

coef.lmm.rrpp: The coef.lmm.rrpp function computes combined fixed and random effects by subject for `lmm.rrpp` model fits. This function does not operate the same as `coef.lm.rrpp`, for which coefficients and fixed effects are the same, and tests can be performed on fixed effects. It is merely a wrapper to combine the results of `ranef.lmm.rrpp` and `fixef.lmm.rrpp`.

Usage

```
## S3 method for class 'lm.rrpp'
coef(object, SE = FALSE, test = FALSE, confidence = 0.95, ...)
```

Arguments

`object`	Object from `lm.rrpp`
`SE`	Whether to include standard errors of coefficients. Standard errors are muted if test = TRUE.
`test`	Logical argument that if TRUE, performs hypothesis tests (Null hypothesis is vector distance = 0) for the observed coefficients. If FALSE, only the observed coefficients are returned.
`confidence`	The desired confidence limit to print with a table of summary statistics, if test = TRUE. Because distances are directionless, confidence limits are one-tailed.
`...`	Other arguments (currently none)

Author(s)

Michael Collyer

See Also

`betaTest`

Examples

```
## Not run:
# See examples for lm.rrpp to see how anova.lm.rrpp works in conjunction
# with other functions

data(Pupfish)
names(Pupfish)
Pupfish$logSize <- log(Pupfish$CS)

fit <- lm.rrpp(coords ~ logSize + Sex*Pop,
SS.type = "I", data = Pupfish, verbose = TRUE)

coef(fit) # Just coefficients
coef(fit, SE = TRUE) # Coefficients with SE
coef(fit, test = TRUE,
confidence = 0.99) # Test of coefficients

## End(Not run)
```

`coef.lmm.rrpp` *coef for lmm.rrpp model fits*

Description

The coef.lmm.rrpp function computes combined fixed and random effects by subject for `lmm.rrpp` model fits. This function does not operate the same as `coef.lm.rrpp`, for which coefficients and fixed effects are the same, and tests can be performed on fixed effects. It is merely a wrapper to combine the results of `ranef.lmm.rrpp` and `fixef.lmm.rrpp`.

Usage

```
## S3 method for class 'lmm.rrpp'
coef(object, type = c("matrix", "list"), ...)
```

Arguments

`object`	Object from `lmm.rrpp`
`type`	Whether to combine coefficients in a matrix or a list by variable.
`...`	Other arguments (currently none)

Author(s)

Michael Collyer

convert2ggplot *Convert RRPP plots to ggplot objects*

Description

Function attempts to coerce plot information from an RRPP plot object to an amenable ggplot object.

Usage

```
convert2ggplot(object)
```

Arguments

object	A plot object produced from plot.lm.rrpp and type equals either "PC" or "regression, plot.predict.lm.rrpp, or plot.ordinate. Essentially, any RRPP plot except a series of diagnostic plots should work.

Details

This function will attempt to use the plot arguments from an RRPP plot object to make a ggplot that can be additionally updated, as desired. Not all plot characteristics can be converted. For example, text arguments are not currently passed to ggplot, as the text function and geom_text arguments do not easily align. However, one can use text arguments produced by a RRPP plot object and geom_text to augment a ggplot object the way they like.

This function assumes no responsibility for arguments made by ggplot. It merely produces a ggplot object that should resemble an RRPP plot default. Any augmentation of ggplot objects can be done either by direct intervention of the ggplot produced or reformatting the initial RRPP plot produced. One should not expect direct correspondence between R base plot parameters and ggplot parameters. For example, error bars will generally appear as different widths, without an easy way to control them, changing from one format to the other.

Author(s)

Michael Collyer

Examples

```
## Not run:
### Linear Model Example

data(Pupfish)
fit <- lm.rrpp(coords ~ log(CS) + Sex*Pop, SS.type = "I",
data = Pupfish, print.progress = FALSE)

# Predictions (holding alternative effects constant)

shapeDF <- expand.grid(Sex = levels(Pupfish$Sex),
```

```
Pop = levels(Pupfish$Pop))
rownames(shapeDF) <- paste(shapeDF$Sex, shapeDF$Pop, sep = ".")

shapePreds <- predict(fit, shapeDF)
summary(shapePreds, PC = TRUE)

# Plot prediction

P <- plot(shapePreds, PC = TRUE, ellipse = TRUE)
convert2ggplot(P)

### Ordination Example

data("PlethMorph")

Y <- as.data.frame(PlethMorph[c("TailLength", "HeadLength",
                                "Snout.eye", "BodyWidth",
                                "Forelimb", "Hindlimb")])
Y <- as.matrix(Y)
R <- lm.rrpp(Y ~ SVL, data = PlethMorph,
             print.progress = FALSE)$LM$residuals

# PCA (on correlation matrix)


PCA.ols <- ordinate(R, scale. = TRUE)
PCA.ols$rot
prcomp(R, scale. = TRUE)$rotation # should be the same


PCA.gls <- ordinate(R, scale. = TRUE,
                    transform. = FALSE,
                    Cov = PlethMorph$PhyCov)

P <- plot(PCA.gls)
convert2ggplot(P)

## End(Not run)
```

`effect.size` *Obtain Effect-size from a vector of values*

Description

A function to find the effect size (Z-score) of a target, from a vector of values presumably obtained in random permutations.

Usage

```
effect.size(x, center = TRUE, ...)
```

Arguments

x	The vector of data to use.
center	Logical value for whether to center x.
...	Arguments passed to powerTrans.

Author(s)

Michael Collyer

---

fishy *Simulated Fish Data for Measurement Error Analysis*

Description

Simulated Fish Data for Measurement Error Analysis

Details

Data as simulated in Collyer and Adams (2024), resembling a fish shape, comprising Procrustes coordinates for 11 anatomical landmarks. Data represent 120 configurations for 60 subjects, each with two replicates of measurement. The 60 subjects represent 20 subjects each from three groups.

Author(s)

Michael Collyer

References

Collyer, M.L, and D.C. Adams. 2024. Interrogating random and systematic measurement error in morphometric data. Evolutionary Biology. In press.

---

fitted.lm.rrpp *Extract fitted values*

Description

Extract fitted values

Usage

```
## S3 method for class 'lm.rrpp'
fitted(object, ...)
```

Arguments

object	plot object (from lm.rrpp)
...	Arguments passed to other functions

Author(s)

Michael Collyer

Examples

```
# See examples for lm.rrpp
```

---

fixef.lmm.rrpp *fixef for lmm.rrpp model fits*

Description

Returns fixed effects from a lmm.rrpp object.

Usage

```
## S3 method for class 'lmm.rrpp'
fixef(object, ...)
```

Arguments

object	Object from lmm.rrpp
...	Other arguments passed.

Author(s)

Michael Collyer

flower *Flower Production and Temperature*

Description

A data set containing variables for flower production and temperature, averaged yearly for 30 years, from a dry tropical forest.

Details

The data in this data set are samples from a larger study that compared flower production between forest types over 30 years, based on climate variables. The data in this set are from a dry tropical forest, include variables for annual flower production, temperature, and year. These data are suitable for considering the temporal non-independence of observations in generalized least squares (GLS) estimation, for the example in the book, Permutational Biometry (Collyer & Adams, 2025).

References

Pau, S., Wolkovich, E. M., Cook, B. I., Nytch, C. J., Regetz, J., Zimmerman, J. K., and Joseph Wright, S. (2013). Clouds and temperature drive dynamic changes in tropical flower production. Nature Climate Change, 3:838–842.

Collyer, M. L., & Adams D. C. (2025). Permutational Biometry: Volume 1, Univariate Data. Iowa State University Digital Press. NOT YET PUBLISHED.

focusMEonSubjects *Plot Function for RRPP*

Description

Reduces a plot.measurement.error to a single research subject. This can be for cases when many overlapping subjects in a plot obscure interpretation for specific subjects.

Usage

```
focusMEonSubjects(x, subjects = NULL, shadow = TRUE, ...)
```

Arguments

x	Plot from plot.measurement.error
subjects	The specific subject to plot
shadow	A logical value for whether to show other subject values as shadows of their locations.
...	Other arguments passed onto plot

Author(s)

Michael Collyer

getANOVAStats *Utility Function for RRPP*

Description

A function mostly for internal processing but can be used to extract ANOVA statistics for other uses, such as plotting histograms

Usage

```
getANOVAStats(fit, stat = c("SS", "MS", "Rsq", "F", "cohenf", "all"))
```

Arguments

fit	Object from lm.rrpp.
stat	The ANOVA statistic to extract. Returns every RRPP permutation of the statistic. If "all", a list of each statistic is returned.

Author(s)

Michael Collyer

Examples

```
## Not run:
data(Pupfish)
fit <- lm.rrpp(coords ~ log(CS) + Sex*Pop, SS.type = "I",
data = Pupfish, print.progress = FALSE, iter = 999)
anova(fit)
Fstats <- getANOVAStats(fit, stat = "F")
par(mfrow = c(2, 2))
hist(Fstats$Fs[1,], breaks = 50, main = "log(CS)", xlab = "F")
abline(v = Fstats$Fs[1, 1])
hist(Fstats$Fs[2,], breaks = 50, main = "Sex", xlab = "F")
abline(v = Fstats$Fs[2, 1])
hist(Fstats$Fs[3,], breaks = 50, main = "Pop", xlab = "F")
abline(v = Fstats$Fs[3, 1])
hist(Fstats$Fs[4,], breaks = 50, main = "Sex:Pop", xlab = "F")
abline(v = Fstats$Fs[4, 1])

## End(Not run)
```

getModelCov *Utility Function for RRPP*

Description

A function mostly for internal processing but can be used to extract either the model covariance matrix (Cov) or the projection matrix for transformations made from the covariance matrix (Pcov), which is basically the square-root of the covariance matrix. This matrix is the model covariance used for estimation, not the residual covariance matrix (see `getResCov`). There are also options for S3 or S4 format versions, or a forcing of symmetry for Pcov.

Usage

```
getModelCov(
  fit,
  type = c("Cov", "Pcov"),
  format = c("S3", "S4"),
  forceSym = TRUE
)
```

Arguments

`fit`	Object from `lm.rrpp`
`type`	Whether the Cov or Pcov matrix is returned
`format`	Whether an S3 or S4 format is returned
`forceSym`	Logical value for whether a symmetric matrix should be returned for Pcov, even if Pcov was triangular as a solution.

Author(s)

Michael Collyer

getModels *Utility Function for RRPP*

Description

A function mostly for internal processing but can be used to obtain terms, design matrices, or QR decompositions used for each reduced or full model that is fitted in an `lm.rrpp` fit.

Usage

```
getModels(fit, attribute = c("terms", "X", "qr", "all"))
```

Arguments

fit	Object from lm.rrpp
attribute	The various attributes that are used to extract RRPP Model information, including "terms", "X" (design matrices), "qr" (QR decompositions), or "all".

Author(s)

Michael Collyer

---

getPermInfo *Utility Function for RRPP*

Description

A function mostly for internal processing but can be used to extract RRPP permutation information for other reasons.

Usage

```
getPermInfo(
  fit,
 attribute = c("perms", "perm.method", "block", "perm.seed", "perm.schedule", "all")
)
```

Arguments

fit	Object from lm.rrpp
attribute	The various attributes that are used to generate RRPP permutation schedules. If there are n observations, each iteration has some randomization of 1:n, restricted by the arguments that match attributes provided by this function.

Author(s)

Michael Collyer

getResCov *Utility Function for RRPP*

Description

A function mostly for internal processing but can be used to extract the residual covariance matrix. This matrix is the residual covariance matrix, not the model covariance matrix used for estimation (see getModelCov). Options for averaging over degrees of freedom or number of observations, plus standardization, are also available.

Usage

```
getResCov(fit, useDf = TRUE, standardize = FALSE)
```

Arguments

fit	Object from lm.rrpp
useDf	Logical value for whether the degrees of freedom from the linear model fit should be used (if TRUE), as opposed to the number of observations (if FALSE).
standardize	Logical value for whether residuals should be standardized. If TRUE, a correlation matrix is produced.

Author(s)

Michael Collyer

getTerms *Utility Function for RRPP*

Description

A function mostly for internal processing but can be used to extract The terms for each reduced and full model used in an lm.rrpp fit.

Usage

```
getTerms(fit)
```

Arguments

fit	Object from lm.rrpp

Author(s)

Michael Collyer

ICCstats *Intraclass correlation statistics from an lm.rrpp model fits*

Description

Function performs analyses concerned with the repeatability (reliability) of multivariate data (measurements) collected from the same research subjects. Although there is no requirement for repeated measurements on all research subjects, the analysis assumes that multiple observations are made.

Usage

```
ICCstats(
  fit,
  subjects = NULL,
  with_in = NULL,
  groups = NULL,
  multivariate = FALSE,
  print.AOV = TRUE
)
```

Arguments

fit	The lm.rrpp, previously evaluated.
subjects	A single character value indicating which term in an ANOVA table corresponds to research subjects.
with_in	One or more character values indicating which terms in an ANOVA table are measured within subjects (replications, plus maybe interactions). If NULL, the only replication within-subject will be considered as residuals.
groups	An optional character value to indicate if a factor in the model frame of the lm.rrpp fit that could account for subject variation. Using this argument might minimize the importance of subject variation, if subjects have disparate values that could inflate ICC. Note that this name could be different than what is shown in the ANOVA table, if measurement.error was used. Use names(fitLMdata), substituting fit with the name assigned to the measurement.error object, to know the groups factor, if used.
multivariate	Logical value for whether to include to calculate ICC matrix generalizations and perform eigenanalysis.
print.AOV	Logical value for whether to include ANOVA table as screen output, when calculating ISS statistics. Note that this function can return ICC statistics, even if they do not make sense. It is possible to generate ICC stats with any ANOVA table, with at least one term.

Details

Function uses ANOVA statistics or SSCP matrices to find the ratio of among-subject to within-subject variance. The former is a dispersion-based approach and the latter is a multivariate generalization of the ICC statistic (as a matrix product). The multivariate generalizations of the statistics described by Liljequist et al. (2019) are used to find matrix products, from which eigenanalysis is performed, providing ICC statistics by eigenvectors.

Three statistics describe the ICC for the population, agreement of measurements among subjects, and consistency between measurements. The last statistic does not necessarily measure the sameness between measurements but the consistency of change between measurements, which might be indicative of a systematic measurement error. If groups are used, these three statistics are repeated, using the SSCP for groups-adjusted data. This approach accounts for group differences, which would avoid large subject variation compared to measurement error inflating ICC values. If there are inherently disparate groups from which subjects are sampled, this approach can elucidate better agreement and consistency in light of group differences.

This function is most useful for analyses performed with `measurement.error`, but any `lm.rrpp` fit can be used, so long as research subjects can be defined.

It is essential that all arguments are terms that can be found in the model frame of the model fit, as provoke by ANOVA. Using anova(fit) will elucidate the row names of the ANOVA that could be used.

Value

Objects of class "ICCstats" return the following:

`ICC_disp`	The intraclass correlation coefficient (ICC) based on the dispersion of values.
`ICC_mult`	The eigenvalues of ICC matrices

Author(s)

Michael Collyer

References

Liljequist, D., Elfving, B., & Skavberg Roaldsen, K. (2019). Intraclass correlation–A discussion and demonstration of basic features. PloS one, 14(7), e0219854.

Examples

```
## Not run:
# Measurement error analysis on simulated data of fish shapes

data(fishy)

# Analysis unconcerned with groups

ME1 <- measurement.error(
  Y = "coords",
  subjects = "subj",
  replicates = "reps",
```

```
  data = fishy)

anova(ME1)
ICCstats(ME1, subjects = "Subjects", with_in = "Systematic ME")

# Analysis concerned with groups

ME2 <- measurement.error(
  Y = "coords",
  subjects = "subj",
  replicates = "reps",
  groups = "groups",
  data = fishy)

anova(ME2)
ICCstats(ME2, subjects = "Subjects",
  with_in = "Systematic ME", groups = "groups")
ICCstats(ME2, subjects = "Subjects",
  with_in = c("Systematic ME", "Systematic ME:Groups"),
  groups = "groups")

## End(Not run)
```

`interSubVar` *Reveal the inter-subject variability from a measurement error analysis*

Description

Function produces both a list of inter-subject Euclidean distance matrices, based on replicate measurements of the same subjects, and one matrix that summarizes the variability among the inter-subject distances, across subjects. This function can be considered a tool for the evaluation of subject estimate precision. The function, `plot.interSubVar` can produce a heat map of the inter-subject variability.

Usage

```
interSubVar(ME, type = c("range", "sd", "var", "cv"))
```

Arguments

`ME`	A measurement error object
`type`	A value to indicate the type of variability (statistic) to measure, which can be one of range (the maximum value minus the minimum value, not the two values), standard deviation (sd), variance (var), or coefficient of variation (cv). No attempt is made to assure the distribution of values is appropriate for the statistics. For example, if only two replicates are available, using sd, var, or cv might not be wise. Or if the replicated values are exact, cv will be NA (and other stats will be 0). Choice of statistic should consider the distribution of values.

Value

An object of class `interSubVar` is a list containing the following

`var.map`	A distance matrix object with values that map the variability statistic used for inter-subject Euclidean distances.
`distance.mats`	The inter-subject distance matrices for every replicate.
`subject.order`	A vector of subject levels in the order that was used to guarantee consistent sorting across distance matrices.
`var.map`	The variability type (statistic) that was used.

Author(s)

Michael Collyer

Examples

```
## Not run:
# Measurement error analysis on simulated data of fish shapes

data(fishy)

# Analysis unconcerned with groups

ME1 <- measurement.error(
  Y = "coords",
  subjects = "subj",
  replicates = "reps",
  data = fishy)

anova(ME1)
ICCstats(ME1, subjects = "Subjects", with_in = "Systematic ME")
plot(ME1)

# Analysis concerned with groups

ME2 <- measurement.error(
  Y = "coords",
  subjects = "subj",
  replicates = "reps",
  groups = "groups",
  data = fishy)

anova(ME2)
ICCstats(ME2, subjects = "Subjects",
  with_in = "Systematic ME", groups = "groups")
P <- plot(ME2)
focusMEonSubjects(P, subjects = 18:20, shadow = TRUE)

## End(Not run)
```

kcomp *K-component analysis*

Description

Function performs a K-component analysis, which is an eigen decomposition of a ratio of ordinary least-squares (OLS) to generalized least squares (GLS) covariance matrices.

Usage

```
kcomp(Y, Cov, transform. = TRUE, scale. = FALSE, tol = NULL, rank. = NULL)
```

Arguments

Y	An n x p data matrix.
Cov	An required n x n covariance matrix to describe the non-independence among observations in Y, and provide a GLS-centering of data.
transform.	An optional argument to transform GLS-centered residuals, if TRUE. If FALSE, only GLS-centering is performed.
scale.	A logical value indicating whether the variables should be scaled to have unit variance before the analysis takes place. The default is FALSE.
tol	A value indicating the magnitude below which components should be omitted. (Components are omitted if their standard deviations are less than or equal to tol times the standard deviation of the first component.)
rank.	Optionally, a number specifying the maximal rank, i.e., maximal number of K components to be used. This argument can be set as alternative or in addition to tol, useful notably when the desired rank is considerably smaller than the dimensions of the matrix of the K matrix.

Details

The function performs a K-component analysis (KCA), as described in Mitteroecker et al. (2025). The analysis is similar to many that can be performed with the `ordinate` function, but whereas that function finds ordinate scores for a rotation or shear of a data space, KCA performs a relative eigen decomposition of a matrix product that represents a ratio between two covariance matrices (Mitteroecker and Bookstein, 2014). The eigenvalues are helpful for understanding the importance of the GLS estimation of a covariance matrix. For example, if the GLS estimation is made with respect to a matrix of phylogenetic covariances, the eigenvalues express the distribution of phylogenetic signal across components of the data space.

The matrix product (K matrix) is the inverse of the OLS covariance matrix times the GLS covariance matrix. Redundancies between these matrices will mean the K matrix is likely not full rank. Therefore, an algorithm is used to compute scores in appropriate dimensions. First, data are aligned to the square-root matrix of the GLS covariance matrix (Collyer and Adams, 2021), using the `ordinate` function. Second, relative eigen decomposition is performed, and the number of real eigenvectors that can be computed are retained. Lastly, the same number of aligned components are used for projection of data onto relative eigenvectors (the K components).

Value

An object of class kcomp is a list containing the following

values	The eigenvalues of the K matrix,
vectors	The eigenvectors of the K matrix.
scores	The projected scores of data onto the eigenvectors.

Author(s)

Michael Collyer

References

Collyer, M.L. and D.C. Adams. 2021. Phylogenetically-aligned Component Analysis. Methods in Ecology and evolution. In press.

Bookstein, F. L., & Mitteroecker, P. (2014). Comparing covariance matrices by relative eigenanalysis, with applications to organismal biology. Evolutionary biology, 41, 336-350.

Mitteroecker, P., Collyer, M. L., & Adams, D. C. (2024). Exploring Phylogenetic Signal in Multivariate Phenotypes by Maximizing Blomberg's K. Systematic Biology, syae035.

See Also

plot.kcomp, ordinate, plot.default, rrpp.data.frame

Examples

```
data(PlethMorph)
PCA <- ordinate(as.data.frame(PlethMorph[c("TailLength",
"HeadLength", "Snout.eye", "BodyWidth","Forelimb", "Hindlimb")],
scale. = TRUE))
Y <- PCA$x
KCA <- kcomp(Y, Cov = PlethMorph$PhyCov, tol = 0.001)
```

---

lm.rrpp *Linear Model Evaluation with a Randomized Residual Permutation Procedure*

Description

Function performs a linear model fit over many random permutations of data, using a randomized residual permutation procedure.

Usage

```
lm.rrpp(
  f1,
  iter = 999,
  turbo = FALSE,
  seed = NULL,
  int.first = FALSE,
  RRPP = TRUE,
  full.resid = FALSE,
  block = NULL,
  SS.type = c("I", "II", "III"),
  data = NULL,
  Cov = NULL,
  print.progress = FALSE,
  Parallel = FALSE,
  verbose = FALSE,
  ...
)
```

Arguments

f1	A formula for the linear model (e.g., y~x1+x2). Can also be a linear model fit from lm.
iter	Number of iterations for significance testing
turbo	A logical value that if TRUE, suppresses coefficient estimation in every random permutation. This will affect subsequent analyses that require random coefficients (see coef.lm.rrpp) but might be useful for large data sets for which only ANOVA is needed.
seed	An optional argument for setting the seed for random permutations of the resampling procedure. If left NULL (the default), the exact same P-values will be found for repeated runs of the analysis (with the same number of iterations). If seed = "random", a random seed will be used, and P-values will vary. One can also specify an integer for specific seed values, which might be of interest for advanced users.
int.first	A logical value to indicate if interactions of first main effects should precede subsequent main effects
RRPP	A logical value indicating whether residual randomization should be used for significance testing
full.resid	A logical value for whether to use the full model residuals, only (sensu ter Braak, 1992). This only works if RRPP = TRUE and SS.type = III. Rather than permuting reduced model residuals, this option permutes only the full model residuals in every random permutation of RRPP.
block	An optional factor for blocks within which to restrict resampling permutations.
SS.type	A choice between type I (sequential), type II (hierarchical), or type III (marginal) sums of squares and cross-products computations.
data	A data frame for the function environment, see rrpp.data.frame

Cov	An optional argument for including a covariance matrix to address the non-independence of error in the estimation of coefficients (via GLS). If included, any weights are ignored.
print.progress	A logical value to indicate whether a progress bar should be printed to the screen. This is helpful for long-running analyses.
Parallel	Either a logical value to indicate whether parallel processing should be used, a numeric value to indicate the number of cores to use, or a predefined socket cluster. This argument defines parallel processing via the parallel library. If TRUE, this argument invokes forking or socket cluster assignment of all processor cores, except one. If FALSE, only one core is used. A numeric value directs the number of cores to use, but one core will always be spared. If a predefined socket cluster (Windows) is provided, the cluster information will be passed to parallel.
verbose	A logical value to indicate if all possible output from an analysis should be retained. Generally this should be FALSE, unless one wishes to extract, e.g., all possible terms, model matrices, QR decomposition, or random permutation schemes.
...	Arguments typically used in lm, such as weights or offset, passed on to LM.fit (an internal RRPP function) for estimation of coefficients. If both weights and a covariance matrix are included, weights are ignored (since inverses of weights are the diagonal elements of weight matrix, used in lieu of a covariance matrix.)

Details

The function fits a linear model using ordinary least squares (OLS) or generalized least squares (GLS) estimation of coefficients over any number of random permutations of the data. A permutation procedure that randomizes vectors of residuals is employed. This procedure can randomize two types of residuals: residuals from null models or residuals from an intercept model. The latter is the same as randomizing full values, and is referred to as as a full randomization permutation procedure (FRPP); the former uses the residuals from null models, which are defined by the type of sums of squares and cross-products (SSCP) sought in an analysis of variance (ANOVA), and is referred to as a randomized residual permutation procedure (RRPP). Types I, II, and III SSCPs are supported.

Users define the SSCP type, the permutation procedure type, whether a covariance matrix is included (GLS estimation), and a few arguments related to computations. Results comprise observed linear model results (coefficients, fitted values, residuals, etc.), random sums of squares (SS) across permutation iterations, and other parameters for performing ANOVA and other hypothesis tests, using empirically-derived probability distributions.

lm.rrpp emphasizes estimation of standard deviates of observed statistics as effect sizes from distributions of random outcomes. When performing ANOVA, using the anova function, the effect type (statistic choice) can be varied. See anova.lm.rrpp for more details. Please recognize that the type of SS must be chosen prior to running lm.rrpp and not when applying anova to the lm.rrpp fit, as design matrices for the linear model must be created first. Therefore, SS.type is an argument for lm.rrpp and effect.type is an argument for anova.lm.rrpp. If MANOVA statistics are preferred, eigenvalues can be added with manova.update and statistics summarized with summary.manova.lm.rrpp. See manova.update for examples.

The coef.lm.rrpp function can be used to test the specific coefficients of an lm.rrpp fit. The test statistics are the distances (d), which are also standardized (Z-scores). The Z-scores might be easier to compare, as the expected values for random distances can vary among coefficient vectors (Adams and Collyer 2016).

ANOVA vs. MANOVA:

Two SSCP matrices are calculated for each linear model effect, for every random permutation: R (Residuals or Random effects) and H, the difference between SSCPs for "full" and "reduced" models. (Full models contain and reduced models lack the effect tested; SSCPs are hypothesized to be the same under a null hypothesis, if there is no effect. The difference, H, would have a trace of 0 if the null hypothesis were true.) In RRPP, ANOVA and MANOVA correspond to two different ways to calculate statistics from R and H matrices.

ANOVA statistics are those that find the trace of R and H SSCP matrices before calculating subsequent statistics, including sums of squares (SS), mean squares (MS), and F-values. These statistics can be calculated with univariate data and provide univariate-like statistics for multivariate data. These statistics are dispersion measures only (covariances among variables do not contribute) and are the same as "distance-based" stats proposed by Goodall (1991) and Anderson (2001). MANOVA stats require multivariate data and are implicitly affected by variable covariances. For MANOVA, the inverse of R times H (invR.H) is first calculated for each effect, then eigenanalysis is performed on these matrix products. Multivariate statistics are calculated from the positive, real eigenvalues. In general, inferential conclusions will be similar with either approach, but effect sizes might differ.

ANOVA tables are generated by anova.lm.rrpp on lm.rrpp fits and MANOVA tables are generated by summary.manova.lm.rrpp, after running manova.update on lm.rrpp fits.

Currently, mixed model effects are only possible with $ANOVA statistics, not $MANOVA.

More detail is found in the vignette, ANOVA versus MANOVA.

Notes for RRPP 0.5.0 and subsequent versions:

The output from lm.rrpp has changed, compared to previous versions. First, the $LM component of output no longer includes both OLS and GLS statistics, when GLS fits are performed. Only GLS statistics (coefficients, residuals, fitted values) are provided and noted with a "gls." tag. GLS statistics can include those calculated when weights are input (similar to the lm argument). Unlike previous versions, GLS and weighted LS statistics are not labeled differently, as weighted LS is one form of generalized LS estimation. Second, a new object, $Models, is included in output, which contains the linear model fits (lm attributes) for all reduced and full models that are possible to estimate fits.

Notes for RRPP 0.3.1 and subsequent versions:

F-values via RRPP are calculated with residual SS (RSS) found uniquely for any model terms, as per Anderson and ter Braak (2003). This method uses the random pseudo-data generated by each term's null (reduced) model, meaning RSS can vary across terms. Previous versions used an intercept-only model for generating random pseudo-data. This generally has appropriate type I error rates but can have elevated type I error rates if the observed RSS is small relative to total SS. Allowing term by term unique RSS alleviates this concern.

Value

An object of class lm.rrpp is a list containing the following

call	The matched call.
LM	Linear Model objects, including data (Y), coefficients, design matrix (X), sample size (n), number of dependent variables (p), dimension of data space (p.prime), QR decomposition of the design matrix, fitted values, residuals, weights, offset, model terms, data (model) frame, random coefficients (through permutations), random vector distances for coefficients (through permutations), whether OLS or GLS was performed, and the mean for OLS and/or GLS methods. Note that the data returned resemble a model frame rather than a data frame; i.e., it contains the values used in analysis, which might have been transformed according to the formula. The response variables are always labeled Y.1, Y.2, ..., in this frame.
ANOVA	Analysis of variance objects, including the SS type, random SS outcomes, random MS outcomes, random R-squared outcomes, random F outcomes, random Cohen's f-squared outcomes, P-values based on random F outcomes, effect sizes for random outcomes, sample size (n), number of variables (p), and degrees of freedom for model terms (df). These objects are used to construct ANOVA tables.
PermInfo	Permutation procedure information, including the number of permutations (perms), The method of residual randomization (perm.method), and each permutation's sampling frame (perm.schedule), which is a list of reordered sequences of 1:n, for how residuals were randomized.
Models	Reduced and full model fits for every possible model combination, based on terms of the entire model, plus the method of SS estimation.

Author(s)

Michael Collyer

References

Anderson MJ. 2001. A new method for non-parametric multivariate analysis of variance. Austral Ecology 26: 32-46.

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ter Braak, C.J.F. 1992. Permutation versus bootstrap significance tests in multiple regression and ANOVA. pp .79–86 In Bootstrapping and Related Techniques. eds K-H. Jockel, G. Rothe & W. Sendler.Springer-Verlag, Berlin. lm for more on linear model fits.

See Also

procD.lm and procD.pgls within geomorph;

Examples

```
## Not run:

# Examples use geometric morphometric data
# See the package, geomorph, for details about obtaining such data

data("PupfishHeads")
names(PupfishHeads)

# Head Size Analysis (Univariate)-------------------------------------------------------

fit <- lm.rrpp(log(headSize) ~ sex + locality/year, SS.type = "I",
data = PupfishHeads, print.progress = FALSE, iter = 999)
summary(fit)
anova(fit, effect.type = "F") # Maybe not most appropriate
anova(fit, effect.type = "Rsq") # Change effect type, but still not
# most appropriate

# Mixed-model approach (most appropriate, as year sampled is a random
# effect:

anova(fit, effect.type = "F", error = c("Residuals", "locality:year",
"Residuals"))

# Change to Type III SS

fit <- lm.rrpp(log(headSize) ~ sex + locality/year, SS.type = "III",
data = PupfishHeads, print.progress = FALSE, iter = 999,
verbose = TRUE)
summary(fit)
anova(fit, effect.type = "F", error = c("Residuals", "locality:year",
"Residuals"))

# Coefficients Test

coef(fit, test = TRUE)

# Predictions (holding alternative effects constant)

sizeDF <- data.frame(sex = c("Female", "Male"))
rownames(sizeDF) <- c("Female", "Male")
sizePreds <- predict(fit, sizeDF)
summary(sizePreds)
plot(sizePreds)

# Diagnostics plots of residuals

plot(fit)

# Body Shape Analysis (Multivariate) -----------

data(Pupfish)
```

```
names(Pupfish)

# Note:

dim(Pupfish$coords) # highly multivariate!
fit <- lm.rrpp(coords ~ log(CS) + Sex*Pop, SS.type = "I",
data = Pupfish, print.progress = FALSE, iter = 999,
verbose = TRUE)
summary(fit, formula = FALSE)
anova(fit)
coef(fit, test = TRUE)

# Predictions (holding alternative effects constant)

shapeDF <- expand.grid(Sex = levels(Pupfish$Sex),
Pop = levels(Pupfish$Pop))
rownames(shapeDF) <- paste(shapeDF$Sex, shapeDF$Pop, sep = ".")
shapeDF

shapePreds <- predict(fit, shapeDF)
summary(shapePreds)
summary(shapePreds, PC = TRUE)

# Plot prediction

plot(shapePreds, PC = TRUE)
plot(shapePreds, PC = TRUE, ellipse = TRUE)

# Diagnostics plots of residuals

plot(fit)

# PC-plot of fitted values

groups <- interaction(Pupfish$Sex, Pupfish$Pop)
plot(fit, type = "PC", pch = 19, col = as.numeric(groups))

# Regression-like plot

plot(fit, type = "regression", reg.type = "PredLine",
    predictor = log(Pupfish$CS), pch=19,
    col = as.numeric(groups))

# Body Shape Analysis (Distances) ----------

D <- dist(Pupfish$coords) # inter-observation distances
length(D)
Pupfish$D <- D

fitD <- lm.rrpp(D ~ log(CS) + Sex*Pop, SS.type = "I",
data = Pupfish, print.progress = FALSE, iter = 999)

# These should be the same:
```

```
summary(fitD, formula = FALSE)
summary(fit, formula = FALSE)

# GLS Example (Univariate) -----------

data(PlethMorph)
fitOLS <- lm.rrpp(TailLength ~ SVL, data = PlethMorph,
print.progress = FALSE, iter = 999)
fitGLS <- lm.rrpp(TailLength ~ SVL, data = PlethMorph, Cov = PlethMorph$PhyCov,
print.progress = FALSE, iter = 999)

anova(fitOLS)
anova(fitGLS)

sizeDF <- data.frame(SVL = sort(PlethMorph$SVL))

# Prediction plots

# By specimen
plot(predict(fitOLS, sizeDF)) # Correlated error
plot(predict(fitGLS, sizeDF)) # Independent error

# With respect to independent variable (using abscissa)
plot(predict(fitOLS, sizeDF), abscissa = sizeDF) # Correlated error
plot(predict(fitGLS, sizeDF), abscissa = sizeDF) # Independent error


# GLS Example (Multivariate) -----------

Y <- as.matrix(cbind(PlethMorph$TailLength,
PlethMorph$HeadLength,
PlethMorph$Snout.eye,
PlethMorph$BodyWidth,
PlethMorph$Forelimb,
PlethMorph$Hindlimb))
PlethMorph$Y <- Y
fitOLSm <- lm.rrpp(Y ~ SVL, data = PlethMorph,
print.progress = FALSE, iter = 999)
fitGLSm <- lm.rrpp(Y ~ SVL, data = PlethMorph,
Cov = PlethMorph$PhyCov,
print.progress = FALSE, iter = 999)

anova(fitOLSm)
anova(fitGLSm)

# Prediction plots

# By specimen
plot(predict(fitOLSm, sizeDF)) # Correlated error
plot(predict(fitGLSm, sizeDF)) # Independent error

# With respect to independent variable (using abscissa)
plot(predict(fitOLSm, sizeDF), abscissa = sizeDF) # Correlated error
```

```
plot(predict(fitGLSm, sizeDF), abscissa = sizeDF) # Independent error

## End(Not run)
```

`lm.rrpp.ws` *Linear Model Evaluation with RRPP performed within subjects*

Description

Function performs a linear model fit over many random permutations of data, using a randomized residual permutation procedure restricted to subjects.

Usage

```
lm.rrpp.ws(
  f1,
  subjects,
  iter = 999,
  turbo = FALSE,
  seed = NULL,
  int.first = FALSE,
  RRPP = TRUE,
  data,
  Cov = NULL,
  delta = 0.001,
  gamma = c("sample", "equal"),
  print.progress = FALSE,
  verbose = FALSE,
  Parallel = FALSE,
  ...
)
```

Arguments

`f1`	A formula for the linear model (e.g., y~x1+x2).
`subjects`	A variable that can be found in the data frame indicating the research subjects for the analysis. This variable must be in the data frame. Is can be either numeric (if its slot in the data frame is known) or a character, e.g., "sub_id". It is imperative that it is ordered the same as the data but that the data do not have row names the same as subjects. For example, the subjects variable in the data frame might be sub_id: sub1, sub1, sub1, sub2, sub2, sub2, ... and the row names of the data might be obs1, obs2, obs3, obs4, obs5, obs6, ... The data do not need to have row names but the subjects variable has to be provided.
`iter`	Number of iterations for significance testing
`turbo`	A logical value that if TRUE, suppresses coefficient estimation in every random permutation. This will affect subsequent analyses that require random coefficients (see `coef.lm.rrpp`) but might be useful for large data sets for which only ANOVA is needed.

seed	An optional argument for setting the seed for random permutations of the resampling procedure. If left NULL (the default), the exact same P-values will be found for repeated runs of the analysis (with the same number of iterations). If seed = "random", a random seed will be used, and P-values will vary. One can also specify an integer for specific seed values, which might be of interest for advanced users.
int.first	A logical value to indicate if interactions of first main effects should precede subsequent main effects
RRPP	A logical value indicating whether residual randomization should be used for significance testing
data	A data frame for the function environment, see rrpp.data.frame. A data frame is required for this analysis.
Cov	An optional argument for including a covariance matrix to address the non-independence of error in the estimation of coefficients (via GLS). If included, any weights are ignored. This matrix must match in dimensions either the number of subject levels or the number of observations.
delta	A within-subject scaling parameter for covariances, ranging from 0 to 1. If delta = 0, a sight value (0.001) is added to assure variances of the covariance matrix are 0.1 percent larger than covariances.
gamma	A sample-size scaling parameter that is adjusted to be 1 ("equal") scaling or the square-root of the sample size for subject observations ("sample").
print.progress	A logical value to indicate whether a progress bar should be printed to the screen. This is helpful for long-running analyses.
verbose	A logical value to indicate if all possible output from an analysis should be retained. Generally this should be FALSE, unless one wishes to extract, e.g., all possible terms, model matrices, QR decomposition, or random permutation schemes.
Parallel	Either a logical value to indicate whether parallel processing should be used, a numeric value to indicate the number of cores to use, or a predefined socket cluster. This argument defines parallel processing via the parallel library. If TRUE, this argument invokes forking or socket cluster assignment of all processor cores, except one. If FALSE, only one core is used. A numeric value directs the number of cores to use, but one core will always be spared. If a predefined socket cluster (Windows) is provided, the cluster information will be passed to parallel.
...	Arguments typically used in lm, such as weights or offset, passed on to lm.rrpp for estimation of coefficients. If both weights and a covariance matrix are included, weights are ignored (since inverses of weights are the diagonal elements of weight matrix, used in lieu of a covariance matrix.)

Details

The function fits a linear model using ordinary least squares (OLS) or generalized least squares (GLS) estimation of coefficients over any number of random permutations of the data, but the permutations are mostly restricted to occur with subject blocks for any model terms other than subjects. All functionality should resemble that of lm.rrpp. However, an argument for research

subjects is also required. The purpose of this function is to account for the non-independence among observations of research subjects (due to sampling within subjects), while also allowing for the non-independence among subjects to be considered (Adams and Collyer, submitted).

By comparison, the covariance matrix option in `lm.rrpp` must have a one-to-one match to observations, which can be matched by the row names of the data. In this function, the covariance matrix can be the same one used in `lm.rrpp` but the number of observations can be greater. For example, if subjects are species or some other level of taxonomic organization, data can comprise measurements on individuals. Users have the option to expand the covariance matrix for subjects or input one they have generated.

Irrespective of covariance matrix type, the row names of the data matrix must match the subjects. This step assures that the analysis can proceed in `lm.rrpp`. It is also best to make sure to use an `rrpp.data.frame`, so that the subjects can be a name in that data frame. For example, if research subjects are species and data (observations) are collected from individuals within species, then a procedure like the following should produce results:

rownames(Y) <- species

rdf <- rrpp.data.frame(Y = Y, subjects = species, x = x)

fit <- lm.rrpp.ws(Y ~ species * x, subject = species, data = rdf, Cov = myCov, ...)

where ... means other arguments. The covariances in the the Covariance matrix can be sorted by the subjects factor but data will not be sorted. Therefore, names matching the subjects is essential. Additionally, subjects must be a factor in the data frame or a factor in the global environment. It cannot be part of a list. Something like subjects <- mylist$species will not work. Assuring that data and subjects are in the same `rrpp.data.frame` object as data is the best way to avoid errors.

Most attributes for this analysis are explained with `lm.rrpp`. The notable different attributes for this function are that: (1) a covariance matrix for the non-independence of subjects can be either a symmetric matrix that matches in dimensions the number of subjects or the number of observations; (2) a parameter (delta) that can range between near 0 and 1 to calibrate the covariances between observations of different subjects; and (3) a parameter (gamma) that is either 1 (equal) or the square-root of the subject sample size (sample) to calibrate the covariances among observations within subjects. If delta = 0, it is expected that the covariance between individual observations, between subjects, is the same as expected from the covariance matrix, as if observations were the single observations made on subjects. As delta approaches 1, the observations become more independent, as if it is expected that the many observations would not be expected to be as correlated as if from one observation. Increasing delta might be useful, if, for example, many individuals are sampled within species, from different locations, different age groups, etc. Alternatively, the sample size (n_i) for subject i can also influence the trend of inter-subject covariances. If more individual observations are sampled, the correlation between subjects might be favored to be smaller compared to fewer observations. The covariances can be adjusted to allow for greater independence among observations to be assumed for larger samples.

A design matrix, **X**, is constructed with 0s and 1s to indicate subjects association, and it is used to expand the covariance matrix (**C**) by **XCt(X)**, where **t(X)** is the matrix transpose. The parameters in **X** are multiplied by exp(-delta * gamma) to scale the covariances. (If delta = 0 and gamma = 1, they are unscaled.)

These options for scaling covariances could be important for data with hierarchical organization. For example, data sampled from multiple species with expected covariances among species based on phylogenetic distances, might be expected to not covary as strongly if sampling encounters other strata like population, sex, and age. An a priori expectation is that covariances among observations

would be expected to be smaller than between species, if only one observation per species were made.

If one wishes to have better control over between-subject and within-subject covariances, based on either a model or empirical knowledge, a covariance matrix should be generated prior to analysis. One can input a covariance matrix with dimensions the same as **XCt(X)**, if they prefer to define covariances in an alternative way. A function to generate such matrices based on separate inter-subject and intra-subject covariance matrices is forthcoming.

IMPORTANT. It is assumed that either the levels of the covariance matrix (if subject by subject) match the subject levels in the subject argument, or that the order of the covariance matrix (if observation by observation) matches the order of the observations in the data. No attempt is made to reorder a covariance matrix by observations and row-names of data are not used to re-order the covariance matrix. If the covariance matrix is small (same in dimension as the number of subject levels), the function will compile a large covariance matrix that is correct in terms of order, but this is based on the subjects argument, only.

The covariance matrix is important for describing the expected covariances among observations, especially knowing observations between and within subjects are not independent. However, the randomization of residuals in a permutation procedure (RRPP) is also important for testing inter-subject and intra-subject effects. There are two RRPP philosophies used. If the variable for subjects is part of the formula, the subject effect is evaluated with type III sums of squares and cross-products (estimates SSCPs between a model with all terms and a model lacking subject term), and RRPP performed for all residuals of the reduced model. Effects for all other terms are evaluated with type II SSCPs and RRPP restricted to randomization of reduced model residuals, within subject blocks. This assures that subject effects are held constant across permutations, so that intra-subject effects are not confounded by inter-subject effects.

More details will be made and examples provided after publication of articles introducing the novel RRPP approach.

The `lm.rrpp` arguments not available for this function include: full.resid, block, and SS.type. These arguments are fixed because of the within-subject blocking for tests, plus the requirement for type II SS for within-subject effects.

Value

An object of class `lm.rrpp.ws` is a list containing the following

`call`	The matched call.
`LM`	Linear Model objects, including data (Y), coefficients, design matrix (X), sample size (n), number of dependent variables (p), dimension of data space (p.prime), QR decomposition of the design matrix, fitted values, residuals, weights, offset, model terms, data (model) frame, random coefficients (through permutations), random vector distances for coefficients (through permutations), whether OLS or GLS was performed, and the mean for OLS and/or GLS methods. Note that the data returned resemble a model frame rather than a data frame; i.e., it contains the values used in analysis, which might have been transformed according to the formula. The response variables are always labeled Y.1, Y.2, ..., in this frame.
`ANOVA`	Analysis of variance objects, including the SS type, random SS outcomes, random MS outcomes, random R-squared outcomes, random F outcomes, random

	Cohen's f-squared outcomes, P-values based on random F outcomes, effect sizes for random outcomes, sample size (n), number of variables (p), and degrees of freedom for model terms (df). These objects are used to construct ANOVA tables.
PermInfo	Permutation procedure information, including the number of permutations (perms), The method of residual randomization (perm.method), and each permutation's sampling frame (perm.schedule), which is a list of reordered sequences of 1:n, for how residuals were randomized.
Models	Reduced and full model fits for every possible model combination, based on terms of the entire model, plus the method of SS estimation.

Author(s)

Michael Collyer

References

Adams, D.C and M.L Collyer. (submitted) Extended phylogenetic regression models for comparing within-species patterns across the Tree of Life. Methods in Ecology and Evolution

ter Braak, C.J.F. 1992. Permutation versus bootstrap significance tests in multiple regression and ANOVA. pp .79–86 In Bootstrapping and Related Techniques. eds K-H. Jockel, G. Rothe & W. Sendler.Springer-Verlag, Berlin. lm for more on linear model fits.

See Also

lm.rrpp; measurement.error

Examples

```
## Not run:
data(fishy)

suppressWarnings(fit <- lm.rrpp.ws(coords ~ subj + groups * reps,
  subjects = "subj",
  data = fishy))

anova(fit)

## End(Not run)
```

---

lmm.rrpp *Mixed Linear Model Evaluation with RRPP performed within subjects*

Description

Function performs a mixed linear model fit over many random permutations of data, using a randomized residual permutation procedure restricted to subjects. This function is likely to evolve greater flexibility in the future.

Usage

```
lmm.rrpp(
  fixed,
  subjects,
  type = c("intercepts", "slopes"),
  slopeTerm = NULL,
  estimation = c("REML", "ML", "LS"),
  iter = 999,
  data,
  ...
)
```

Arguments

fixed	A formula for the fixed effects of linear model (e.g., y~x1+x2).
subjects	A variable that can be found in the data frame indicating the research subjects for the analysis. This variable must be in the data frame. Is can be either numeric (if its slot in the data frame is known) or a character, e.g., "sub_id". It is imperative that it is ordered the same as the data but that the data do not have row names the same as subjects. For example, the subjects variable in the data frame might be sub_id: sub1, sub1, sub1, sub2, sub2, sub2, ... and the row names of the data might be obs1, obs2, obs3, obs4, obs5, obs6, ... The data do not need to have row names but the subjects variable has to be provided.
type	An indication whether to use random intercepts ("intercepts") or random intercepts and slopes ("slopes") for random effects associated with subjects.
slopeTerm	An optional character value (e.g., "mass") indicating which term in the data frame is to be used in the estimation of random slopes. This term should also be in the fixed formula.
estimation	One of "LS", "ML", or "REML", for guiding how random effects should be estimated.
iter	Number of iterations for significance testing
data	A data frame for the function environment, see rrpp.data.frame. A data frame is required for this analysis.
...	Arguments typically used in lm, lm.rrpp, or lm.rrpp.ws such as weights or offset, seed for permutations, etc., passed on to lm.rrpp.ws for estimation of coefficients.

Details

The function fits a mixed linear model using ordinary least squares (OLS) or generalized least squares (GLS) estimation of coefficients for fixed effects, and maximum likelihood (ML) or restricted ML (REML) for random effects, over any number of random permutations of the data, but the permutations are mostly restricted to occur with subject blocks for any model terms other than subjects. Most functionality should resemble that of lm.rrpp and lm.rrpp.ws, with some caveats. First, estimation of random effects uses the methodology of the lmer function (Bates et al., 2015), for only one random term based on a subjects designation. Second, this means that a "random

slopes" model involves an interaction between subjects and a numeric variable (not a categorical variable). Third, although any number of fixed effects can be used, only one fixed effect can interaction with subjects. Finally, the function is intended to allow analysis of variance (ANOVA), like with `lm.rrpp.ws`, but not treated random effects as fixed. There is not a coefficients test associated with this function, as with other `lm.rrpp` that include only fixed effects.

Although this function allows covariance matrices to be used for generalized least squares (GLS) estimation of fixed effects, it does not currently expand the covariance mnatrix based only on subjects, as does `lm.rrpp.ws`. However, that function can be used to obtain a covariance matrix for this function. The covariance structure (Lambda matrix) among subjects is estimated via ML or REML (Bates et al., 2015) and is not impacted by the expected covariances among fixed effects, however, the expected covariances will influence interactions between fixed and random effects. The Lambda matrix is estimated as a single matrix, even for multivariate data. This matrix is obtained by projecting data onto the first component of a singular value decomposition of the product between the data and a hat matrix of the the fixed effects (basically projection following a principal component analysis of model fitted values). This is a computationally proficient step that is similar to averaging covariance parameters (theta) over all variables. If independent Lambda matrices are desired, then this function can be used on, e.g., each principal component of data, followed by combining the coefficients. For ANOVA, the precise estimation of Lambda is not as important for tests of fixed effects, as it means a linear transformation of coefficients that would be consistent across random permutations of RRPP. It might be more important for model comparison and selection, in which case whether to use LS, ML, REML, and whether to use separate Lambda matrices is a much deeper consideration.

A data frame (preferably an `rrpp.data.frame` object) must be used for this function. Not all "downstream" functions (e.g., logLik, predict) are guaranteed to work as expected, as this function is still very much under development.

The `lm.rrpp` arguments not available for this function include: full.resid, block, and SS.type. These arguments are fixed because of the within-subject blocking for tests, plus the requirement for type II SS for within-subject effects.

Value

An object of class `lmm.rrpp` is a list containing the following

`call`	The matched call.
`LM`	Linear Model objects, including data (Y), coefficients, design matrix (X), sample size (n), number of dependent variables (p), dimension of data space (p.prime), QR decomposition of the design matrix, fitted values, residuals, weights, offset, model terms, data (model) frame, random coefficients (through permutations), random vector distances for coefficients (through permutations), whether OLS or GLS was performed, and the mean for OLS and/or GLS methods. Note that the data returned resemble a model frame rather than a data frame; i.e., it contains the values used in analysis, which might have been transformed according to the formula. The response variables are always labeled Y.1, Y.2, ..., in this frame. Most important objects for mixed models are also found here.
`ANOVA`	Analysis of variance objects, including the SS type, random SS outcomes, random MS outcomes, random R-squared outcomes, random F outcomes, random Cohen's f-squared outcomes, P-values based on random F outcomes, effect sizes for random outcomes, sample size (n), number of variables (p), and degrees of

	freedom for model terms (df). These objects are used to construct ANOVA tables.
PermInfo	Permutation procedure information, including the number of permutations (perms), The method of residual randomization (perm.method), and each permutation's sampling frame (perm.schedule), which is a list of reordered sequences of 1:n, for how residuals were randomized.
devcomp	Deviance components, a list of components used in deviance and likelihood calculations.

Author(s)

Michael Collyer

References

Douglas Bates, Martin Maechler, Ben Bolker, Steve Walker (2015). Fitting Linear Mixed-Effects Models Using lme4. Journal of Statistical Software, 67(1), 1-48. doi:10.18637/jss.v067.i01.

Limitations: This function currently only works for "Random Intercepts" and "Random slopes" models, where the intercepts or slopes of a single subject term (random effect) are free to vary. If a Random Slopes model is used, a single covariate (numeric variable) must be specified for slopes. Generally, this is term that is also among the fixed effects. These limitations are imposed to allow ML or REML estimation. More complex models can be entertained with `lmm.rrpp.ws`, but using only "LS" estimation. If "LS" estimation is used in this function, results should be the same with a comparable formula, using `lmm.rrpp.ws`

If a ML or REML fit is near singular (one or more covariance parameters are 0), the analysis will be arrested rather than continued with some random effects rendered as 0. This should be an indication that the model is over-fit with the type of estimation used.

These limitations might be addressed with future versions of this function, as theoretical advances are made.

See Also

`lm.rrpp`; `lm.rrpp.ws`; `measurement.error`

Examples

```
## Not run:
# Random intercepts models

data(ReptAbund)
fit_ls <- lmm.rrpp(fixed = reptile_abundance ~ Year,
subjects = "Site",
type = "intercepts",
estimation = "LS",
data = ReptAbund)

fit_ml <- lmm.rrpp(fixed = reptile_abundance ~ Year,
subjects = "Site",
type = "intercepts",
```

```
estimation = "ML",
data = ReptAbund)

fit_reml <- lmm.rrpp(fixed = reptile_abundance ~ Year,
subjects = "Site",
type = "intercepts",
estimation = "REML",
data = ReptAbund)

anova(fit_ls, error = c("Site", "Residuals"))
anova(fit_ml, error = c("Site", "Residuals"))
anova(fit_reml, error = c("Site", "Residuals"))

coef(fit_ls, type = "list")
coef(fit_ml, type = "list")
coef(fit_reml, type = "list")

# Random slopes models
# none should not work!

fit_ls <- lmm.rrpp(fixed = reptile_abundance ~ Year,
subjects = "Site",
type = "slopes",
slopeTerm = "Year",
estimation = "LS",
data = ReptAbund)

fit_ml <- lmm.rrpp(fixed = reptile_abundance ~ Year,
subjects = "Site",
type = "slopes",
slopeTerm = "Year",
estimation = "ML",
data = ReptAbund)


fit_reml <- lmm.rrpp(fixed = reptile_abundance ~ Year,
subjects = "Site",
type = "slopes",
slopeTerm = "Year",
estimation = "REML",
data = ReptAbund)

anova(fit_ls, error = c("Year:Site", "Year:Site", "Residuals"))
anova(fit_ml, error = c("Year:Site", "Year:Site", "Residuals"))

coef(fit_ls, type = "list")
coef(fit_ml, type = "list")

## End(Not run)
```

---

logLik.lm.rrpp *Calculate the log-likelihood of a lm.rrpp fit*

Description

logLik.lm.rrpp returns the log-likelihood of an lm.rrpp object or an lmm.rrpp object. Ridge regularization will be performed for ill-conditioned or singular residual covariance matrices, but dimension reduction could be augmented via projection, using the arguments, tol and pc.no. See ordinate for details.

Usage

```
## S3 method for class 'lm.rrpp'
logLik(
  object,
  tol = NULL,
  pc.no = NULL,
  Z = TRUE,
  verbose = FALSE,
  gls.null = FALSE,
  ...
)
```

Arguments

object	Object from lm.rrpp
tol	A value indicating the magnitude below which components should be omitted, following projection. See ordinate for details.
pc.no	Optionally, a number specifying the maximal number of principal components, passed onto ordinate, as argument, rank.
Z	A logical value for whether to calculate Z scores based on RRPP.
verbose	A logical value for whether to return random log-likelihood values, if Z-scores are calculated.
gls.null	A logical value for if a fit has a GLS estimation, should the null model (intercept) also have a GLS estimation, for estimating Z. Should be FALSE if the log-likelihood is measured to compare different GLS estimations for a covariance matrices
...	further arguments passed to or from other methods

Details

Note that options are limited for lmm.rrpp objects. It is currently not possible to use RRPP to find Z scores.

Author(s)

Michael Collyer

looCV *Diagnostic cross-validation tool including ordination based on fitted values*

Description

Function performs a leave-one-out cross-validation estimate (method = "fit") or ordination scores from estimates (method = "PC"). The latter can be helpful for determining if apparent group differences in ordination plots arise merely from data dimensionality. The choice of method essentially chooses one of two functions.

Usage

```
looCV(fit, method = c("fit", "PC"), ...)
```

Arguments

fit	A lm.rrpp fit.
method	Whether to find cross-validated fitted values (fit) or principal component scores (PC) of fitted values.
...	Arguments passed to ordinate

Details

The fit method is standard, using n - 1 observations as training data, and estimating the response for 1 datum, not included in the training data, based on the model design of an original model fit. This method provides alternative fitted values than the original fit.

The PC method uses the strategy of Thioulouse et al. (2021) to perform N ordinations for N observations, in which each of the N observations are left out of the estimation of linear model coefficients, but the vector of data for the left-out observation is projected on the eigenvectors of the fitted values obtained from the leave-one-out cross-validation (jackknife) strategy. The purpose of this diagnostic tool is to determine whether apparent "group differences" in an ordination plot (using the function, ordinate) are because of high-dimensional data (number of variables exceed number of observations) rather than real differences. An apparent group difference is common for high-dimensional data, when variables are far greater in number than observations (Cardini et al., 2019). However, leave-one-out cross-validation can help elucidate whether an observed visual difference is spurious.

This function differs from the strategy of Thioulouse et al. (2021) in two important ways. First, this function uses the linear model design from a lm.rrpp fit, and can contain any number of independent variables, rather than a single factor for groups. Second, after obtaining leave-one-out cross-validated scores, a Procrustes alignment between cross-validated scores and "observed" (real) scores is performed, which minimizes summed squared distances between the alternative ordinations. This latter step assures comparisons are appropriate.

The type = "PC" plot from plot.lm.rrpp has the same scores as obtained from ordinate(Y, A = H), using the ordinate function, where H is a hat matrix (that can be calculated from plot.lm.rrpp output), and Y is a matrix of data. This function updates H for every possible case that one row of Y is left out (meaning the rotation matrix from ordinate is updated N times). If the H matrix

is robust in spite of dropped data and design matrix parameters, the result will be similar to the original ordination. If apparent group differences are spurious, H will tend to change, as will data projections.

The functions summary.looCV and plot.looCV are essential for evaluating results. These support functions compare eigenvalues and projected scores, between observed and cross-validated cases.

This function should be viewed as a diagnostic tool and not as a data transformation tool! The cross-validated scores will not retain Euclidean distances among observations. This could cause problems in analyses that substitute cross-validated scores as data.

Value

An object of class looCV is a list containing the following

d	List of eigenvalues, for observed and cross-validated cases.
scores	List of principal component scores, for observed and cross-validated cases.

Author(s)

Michael Collyer

References

Thioulouse, J., Renaud, S., Dufour, A. B., & Dray, S. (2021). Overcoming the Spurious Groups Problem in Between-Group PCA. Evolutionary Biology, In press.

Cardini, A., O'Higgins, P., & Rohlf, F. J. (2019). Seeing distinct groups where there are none: spurious patterns from between-group PCA. Evolutionary Biology, 46(4), 303-316.

See Also

summary.looCV, plot.looCV, looCV_table

Examples

```
# Example with real group differences

data(Pupfish)
fit <- lm.rrpp(coords ~ Pop * Sex,
data = Pupfish, iter = 0)
CV1f <- looCV(fit)

summary(CV1f)

CV1p <- looCV(fit, method = "PC")

summary(CV1p)

group <- interaction(Pupfish$Pop, Pupfish$Sex)
plot(CV1p, flip = 1, pch = 19, col = group)

# Example with apparent but not real group differences
```

```
n <- NROW(Pupfish$coords)
p <- NCOL(Pupfish$coords)
set.seed(1001)
Yr <- matrix(rnorm(n * p), n, p) # random noise

fit2 <-lm.rrpp(Yr ~ Pop * Sex,
data = Pupfish, iter = 0)
CV2p <- looCV(fit2, method = "PC")
summary(CV2p)
group <- interaction(Pupfish$Pop, Pupfish$Sex)
plot(CV2p, pch = 19, col = group)
```

`looCV_table` *Table for looCV comparisons*

Description

Function simply produces a table of MSE and cross-validated MSE for any number of compared models. The table is sorted from lowest to highest cross-validated MSE. The looCV objects must use method = 'fit' or a table cannot be generated.

Usage

```
looCV_table(...)
```

Arguments

`...`	Any number of `looCV` objects

Author(s)

Michael Collyer

See Also

`looCV`, `model.comparison`

Examples

```
# Example with snakeHS data

data(snakeHS)
fitS <- lm.rrpp(HS ~ SVL, data = snakeHS,
                iter = 0)
fitSR <- lm.rrpp(HS ~ SVL + Region, data = snakeHS,
                 iter = 0)
fitSxR <- lm.rrpp(HS ~ SVL * Region, data = snakeHS,
                  iter = 0)
```

```
CVS <- looCV(fitS)
CVSR <- looCV(fitSR)
CVSxR <- looCV(fitSxR)

summary(CVS)
summary(CVSR)
summary(CVSxR)

looCV_table(CVS, CVSR, CVSxR)
```

`lr_test` *Likelihood ratio test for a linear model, based on RRPP*

Description

Function performs likelihood ratio tests on an lm.rrpp fit, using RRPP or FRPP. Likelihood ratio statistics are calculated for every random permutation, and the effect size is estimated from the distribution of random statistics. The likelihood ratio tests has some resemblance to MANOVA, especially using Wilks' lambda. Sums of squares and cross-products (SSCP) matrices are calculated over the random permutations of a `lm.rrpp` fit. SSCP matrices are computed, as are the inverse of R times H (invR.H), where R is a SSCP for the residuals or random effects and H is the difference between SSCP matrices of full and reduced models (see `manova.update`). From invR.H, Wilks lambda is first estimated, and the likelihood ratio stat is then estimated as -n * log(Wilks).

This function does one of two things. It either performs an update using `manova.update`, using Wilks' lambda as the test statistic, converting Wilks' lambda to likelihood ratio statistics or it uses the results from a previously performed update to calculate new statistics.

Usage

```
lr_test(fit, verbose = FALSE, ...)
```

Arguments

`fit`	Linear model fit from `lm.rrpp` or a fit that has already been updated with `manova.update`.
`verbose`	Logical value for whether to include all random Wilks' lambda and likelihood ratio statistics from random permutations.
`...`	Arguments passed onto `manova.update`.

Author(s)

Michael Collyer

References

Adams, D. C., and M. L. Collyer. 2024. Extended phylogenetic regression models for comparing within-species patterns across the tree of life. Methods in Ecology and Evolution. In review.

Examples

```
# Body Shape Analysis (Multivariate) ----------------

## Not run:
data(Pupfish)

# Although not recommended as a practice, this example will use only
# three principal components of body shape for demonstration.
# A larger number of random permutations should also be used.

Pupfish$shape <- ordinate(Pupfish$coords)$x[, 1:3]


fit <- lm.rrpp(shape ~ log(CS) + Sex, SS.type = "I",
data = Pupfish, print.progress = FALSE, iter = 499)
summary(fit, formula = FALSE)
anova(fit) # ANOVA table

# MANOVA

fit.m <- manova.update(fit, print.progress = FALSE, tol = 0.001)
summary(fit.m, test = "Roy")
summary(fit.m, test = "Wilks")


# Likelihood Ratio Test

LRT <- lr_test(fit.m)
summary(LRT)


## End(Not run)
```

`mahal_dist` *Calculate the pairwise Mahalanobis distances between observations*

Description

This function emulates the `dist` function but allows a covariance matrix (Cov) to be included for standardizing distances. It is assumed that the Covariance matrix makes sense with respect to the data, and that the number of variables match between data and covariance matrix.

Usage

```
mahal_dist(x, Cov, ...)
```

Arguments

x	A numeric matrix of data frame.
Cov	A covariance matrix with the same number of variables as the data.
...	Other arguments passed to dist.

Details

No tests are performed on distances but could be performed with the pairwise function. Distances are only calculated if the covariance matrix is not singular.

Value

An object of class "dist".

Author(s)

Michael Collyer

Examples

```
# Using the Pupfish data (see lm.rrpp help for more detail)

data(Pupfish)
Pupfish$Y <- ordinate(Pupfish$coords)$x[, 1:3]
fit <- lm.rrpp(Y ~ Sex * Pop, SS.type = "I",
data = Pupfish, print.progress = FALSE, iter = 0)
X <- unique(as.matrix(model.matrix(fit)))
means <- X %*% coef(fit)
rownames(means) <- unique(interaction(Pupfish$Sex, Pupfish$Pop))
means
S <- getResCov(fit)
dist(means)
mahal_dist(means, S)
```

---

mammalHR *Mammal Home Range Data*

Description

Home range and body mass data for 49 species of mammals.

Details

The data set is from a study of mammal species home range size (squared kilometers) and its covariation with body mass or whether mammals are carnivores or ungulates (Garland et al., 1993). the data set also includes a covariance matrix based on phylogenetic relatedness among species, to account for the non-independence of observations. These data are suitable data for demonstrating generalized least squares (GLS) estimation, used in the book, Permutational Biometry (Collyer & Adams, 2025).

References

Garland, T., Dickerman, A. W., Janis, C. M., and Jones, J. A. (1993). Phylogenetic analysis of covariance by computer simulation. Systematic Biology, 42:265–292.

Collyer, M. L., & Adams D. C. (2025). Permutational Biometry: Volume 1, Univariate Data. Iowa State University Digital Press. NOT YET PUBLISHED.

`mammalHRphy` *Phylogenetic Data for the Mammal Home Range Data*

Description

Phylogenetic tree data that can be used along with `mammalHR`.

Details

A class `phylo` object containing data for the phylogenetic tree that pertains to the `mammalHR` data set. Although these data are not required for the generalized least squares (GLS) estimation example, used in the book, Permutational Biometry (Collyer & Adams, 2025), they provide additional graphical support for the example.

References

Collyer, M. L., & Adams D. C. (2025). Permutational Biometry: Volume 1, Univariate Data. Iowa State University Digital Press. NOT YET PUBLISHED.

`manova.update` *MANOVA update for lm.rrpp model fits*

Description

Function updates a lm.rrpp fit to add $MANOVA, which like $ANOVA, provides statistics or matrices typically associated with multivariate analysis of variance (MANOVA).

MANOVA statistics or sums of squares and cross-products (SSCP) matrices are calculated over the random permutations of a `lm.rrpp` fit. SSCP matrices are computed, as are the inverse of R times H (invR.H), where R is a SSCP for the residuals or random effects and H is the difference between SSCP matrices of full and reduced models (see below). From invR.H, MANOVA statistics are calculated, including Roy's maximum root (eigenvalue), Pillai trace, Hotelling-Lawley trace, and Wilks lambda (via `summary.manova.lm.rrpp`).

The manova.update to add $MANOVA to `lm.rrpp` fits requires more computation time than the $ANOVA statistics that are computed automatically in `lm.rrpp`. Generally, the same inferential conclusions will be found with either approach, when observations outnumber response variables. For high-dimensional data (more variables than observations) data are projected into a Euclidean space of appropriate dimensions (rank of residual covariance matrix). One can vary the tolerance for eigenvalue decay or specify the number of PCs, if a smaller set of PCs than the maximum is desired.

This is advised if there is strong correlation among variables (the data space could be simplified to fewer dimensions), as spurious results are possible. Because distributions of MANOVA stats can be generated from the random permutations, there is no need to approximate F-values, like with parametric MANOVA. By restricting analysis to the real, positive eigenvalues calculated, all statistics can be calculated (but Wilks lambda, as a product but not a trace, might be less reliable as variable number approaches the number of observations).

ANOVA vs. MANOVA:

Two SSCP matrices are calculated for each linear model effect, for every random permutation: R (Residuals or Random effects) and H, the difference between SSCPs for "full" and "reduced" models. (Full models contain and reduced models lack the effect tested; SSCPs are hypothesized to be the same under a null hypothesis, if there is no effect. The difference, H, would have a trace of 0 if the null hypothesis were true.) In RRPP, ANOVA and MANOVA correspond to two different ways to calculate statistics from R and H matrices.

ANOVA statistics are those that find the trace of R and H SSCP matrices before calculating subsequent statistics, including sums of squares (SS), mean squares (MS), and F-values. These statistics can be calculated with univariate data and provide univariate-like statistics for multivariate data. These statistics are dispersion measures only (covariances among variables do not contribute) and are the same as "distance-based" stats proposed by Goodall (1991) and Anderson (2001). MANOVA stats require multivariate data and are implicitly affected by variable covariances. For MANOVA, the inverse of R times H (invR.H) is first calculated for each effect, then eigen-analysis is performed on these matrix products. Multivariate statistics are calculated from the positive, real eigenvalues. In general, inferential conclusions will be similar with either approach, but effect sizes might differ.

Two important differences between manova.update and `summary.manova` (for `lm` objects) are that manova.update does not attempt to normalize residual SSCP matrices (unneeded for non-parametric statistical solutions) and (2) uses a generalized inverse of the residual SSCP, if needed, when the number of variables could render eigen-analysis problematic. This approach is consistent with covariance regularization methods that attempt to make covariance matrices positive-definite for calculating model likelihoods or multivariate statistics. If the number of observations far exceeds the number of response variables, observed statistics from manova.update and `summary.manova` will be quite similar. If the number of response variables approaches or exceeds the number of observations, manova.update statistics will be much more reliable.

ANOVA tables are generated by `anova.lm.rrpp` on lm.rrpp fits and MANOVA tables are generated by `summary.manova.lm.rrpp`, after running manova.update on lm.rrpp fits.

Currently, mixed model effects are only possible with $ANOVA statistics, not $MANOVA.

More detail is found in the vignette, ANOVA versus MANOVA.

Usage

```
manova.update(
  fit,
  error = NULL,
  tol = 1e-07,
  PC.no = NULL,
  print.progress = FALSE,
  verbose = NULL
)
```

Arguments

fit	Linear model fit from lm.rrpp
error	An optional character string to define R matrices used to calculate invR.H. (Currently only Residuals can be used and this argument defaults to NULL. Future versions will update this argument.)
tol	A tolerance value for culling data dimensions to prevent spurious results. The distribution of eigenvalues for the data will be examined and if the decay becomes less than the tolerance, the data will be truncated to principal components ahead of this point. This will possibly prevent spurious results calculated from eigenvalues near 0. If tol = 0, all possible PC axes are used, which is likely not a problem if observations outnumber variables. If tol = 0 and the number of variables exceeds the number of observations, the value of tol will be made slightly positive to prevent problems with eigen-analysis.
PC.no	A value that, if not NULL, can override the tolerance argument, and forces a desired number of data PCs to use for analysis. If a value larger than the possible number of PCs is chosen, the full compliment of PCs (the full data space) will be used. If a number larger than tol would permit is chosen, the minimum number of PCs between the tol argument and PC.no argument is returned.
print.progress	A logical value to indicate whether a progress bar should be printed to the screen. This is helpful for long-running analyses.
verbose	Either a NULL or logical value for whether to retain all MANOVA result (if TRUE). If NULL, the verbose argument used for the lm.rrpp is retained for MANOVA update. Essentially, verbose indicates whether to retain all SSCP matrices and all invR.H matrices, for every model effect, in every RRPP iteration.

Value

An object of class lm.rrpp is updated to include class manova.lm.rrpp, and the object, $MANOVA, which includes

SSCP	Terms and Model SSCP matrices.
invR.H	The inverse of the residuals SSCP times the H SSCP.
eigs	The eigenvalues of invR.H.
e.rank	Rank of the error (residuals) covariance matrix. Currently NULL only.
PCA	Principal component analysis of data, using either tol or PC.no.
manova.pc.dims	Resulting number of PC vectors in the analysis.
e.rank	Rank of the residual (error) covariance matrix, irrespective of the number of dimensions used for analysis.

Author(s)

Michael Collyer

References

Goodall, C.R. 1991. Procrustes methods in the statistical analysis of shape. Journal of the Royal Statistical Society B 53:285-339.

Anderson MJ. 2001. A new method for non-parametric multivariate analysis of variance. Austral Ecology 26: 32-46.

Examples

```
## Not run:
# Body Shape Analysis (Multivariate) ----------------

data(Pupfish)

# Although not recommended as a practice, this example will use only
# three principal components of body shape for demonstration.
# A larger number of random permutations should also be used.

Pupfish$shape <- ordinate(Pupfish$coords)$x[, 1:3]

Pupfish$logSize <- log(Pupfish$CS)

fit <- lm.rrpp(shape ~ logSize + Sex, SS.type = "I",
data = Pupfish, print.progress = FALSE, iter = 499)
summary(fit, formula = FALSE)
anova(fit) # ANOVA table

# MANOVA

fit.m <- manova.update(fit, print.progress = FALSE, tol = 0.001)
summary(fit.m, test = "Roy")
summary(fit.m, test = "Pillai")

fit.m$MANOVA$eigs$obs # observed eigenvalues
fit.m$MANOVA$SSCP$obs # observed SSCP
fit.m$MANOVA$invR.H$obs # observed invR.H

# Distributions of test statistics

summ.roy <- summary(fit.m, test = "Roy")
dens <- apply(summ.roy$rand.stats, 1, density)
par(mfcol = c(1, length(dens)))
for(i in 1:length(dens)) {
     plot(dens[[i]], xlab = "Roy max root", ylab = "Density",
     type = "l", main = names(dens)[[i]])
     abline(v = summ.roy$rand.stats[1, i], col = "red")
}
par(mfcol = c(1,1))

## End(Not run)
```

`measurement.error` *Evaluation of measurement error for two or more multivariate measurements, for common research subjects*

Description

Function performs analyses concerned with the repeatability (reliability) of multivariate data (measurements) collected from the same research subjects. Although there is no requirement for repeated measurements on all research subjects, the analysis assumes that multiple observations are made.

Usage

```
measurement.error(
  data,
  Y,
  subjects,
  replicates,
  groups = NULL,
  groups.first = FALSE,
  iter = 999,
  seed = NULL,
  multivariate = FALSE,
  use.PCs = TRUE,
  tol = 0.001,
  Parallel = FALSE,
  turbo = TRUE,
  print.progress = FALSE,
  verbose = FALSE
)
```

Arguments

`data`	A required data frame, either of class `data.frame` or class `rrpp.data.frame`. This function cannot be used without a data frame. All arguments for data and variables are names that must exist in the data frame.
`Y`	A name for a matrix (n x p) of data for n observations and p variables that can be found in the data frame. For example, Y = "morphData".
`subjects`	A name for a vector or factor of research subjects, found within the data frame (each subject should occur twice or more). The length of the vector in the data frame must equal the number of observations and will be coerced into a factor. For example, subjects = "ID".
`replicates`	A name for a vector or factor for replicate measurements for research subjects, found within the data frame. The length of the vector in the data frame must equal the number of observations and will be coerced into a factor. For example, replicates = "Rep".

groups	An optional name for a vector in the data frame, coercible to factor, to be included in the linear model (as an interaction with replicates). This would be of interest if one were concerned with systematic ME occurring perhaps differently among certain strata within the data. For example, systematic ME because of an observer bias might only be observed with females or males, in which case the argument might be: groups = "Sex".
groups.first	A logical value for whether to use groups as a term before subjects, so that it can be included in an ANOVA table (if TRUE). Otherwise, a group effect is likely subsumed by a subject effect, since subjects are unique to groups.
iter	Number of iterations for significance testing
seed	An optional argument for setting the seed for random permutations of the resampling procedure. If left NULL (the default), the exact same P-values will be found for repeated runs of the analysis (with the same number of iterations). If seed = "random", a random seed will be used, and P-values will vary. One can also specify an integer for specific seed values, which might be of interest for advanced users.
multivariate	Logical value for whether to include multivariate analyses. Intraclass correlation matrices and relative eigenanalysis are based on products of sums of squares and cross-products (SSCP) matrices, some of which must be inverted and potentially require significant computation time. If FALSE, only statistics based on dispersion of values are calculated.
use.PCs	A logical argument for whether to use the principal components of the data. This might be helpful for relative eigenanalysis, and if p > n, in which case inverting singular covariance matrices would not be possible.
tol	A value indicating the magnitude below which components should be omitted, if use.PCs is TRUE. (Components are omitted if their standard deviations are less than or equal to tol times the standard deviation of the first component.) See ordinate for more details.
Parallel	The same argument as in lm.rrpp to govern parallel processing (either a logical value – TRUE or FALSE – or the number of threaded cores to use). See lm.rrpp for additional details.
turbo	Logical value for whether to suppress coefficient estimation in RRPP iteration, thus turbo-charging RRPP.
print.progress	A logical value to indicate whether a progress bar should be printed to the screen.
verbose	A logical value to indicate if all the output from an lm.rrpp analysis should be retained. If FALSE, only the needed output for summaries and plotting is retained.

Details

This function performs analyses as described in Collyer and Adams (2024) to assess systematic and random components of measurement error (ME). It basically performs ANOVA with RRPP, but with different restricted randomization strategies. The reliability of research subject variation can be considered by restricting randomization within replicates; the consistency of replicate measures can be considered by restricting randomization within subjects. Inter-subject variation remains constant across all random permutations within subjects and inter-replicate variation remains constant across

all random permutations within replicates. Type II sums of squares and cross-products (SSCP) are calculated to assure conditional estimation.

The results include univariate-like statistics based on dispersion of values and eigenanalysis performed on a signal to noise matrix product of SSCP matrices (sensu Bookstein and Mitteroecker, 2014) including the inverse of the random component of ME and the systematic component of ME. The multivariate test is a form of multivariate ANOVA (MANOVA), using RRPP to generate sampling distributions of the major eigenvalue (Roy's maximum root). Likelihood-ratio tests can also be performed using `lr_test`.

Intraclass correlation coefficients (ICC) can also be calculated (using `ICCstats`), both based on dispersion of values and covariance matrices, as descriptive statistics. Details are provided in `ICCstats`.

Value

Objects of class "measurement.error" return the same objects as a `lm.rrpp` fit, plus a list of the following:

`AOV` Analysis of variance to test for systematic error, based on dispersion of values.

`mAOV` Multivariate AOV based on product of the inverse of the random component (SSCP) of ME times the systematic component of ME.

`SSCP` The sums of squares and cross-products matrices for model effects.

`SSCP.ME.product`
The products of the inverse of the random ME SSCP and the SSCP matrices for systematic ME,. These are the same matrix products used for eigenanalysis. This is the observed matrix.

`SSCP.ME.product.std`
A list of the symmetric forms of standardized SSCP.ME.products that yield orthogonal eigenvectors.

Author(s)

Michael Collyer

References

Collyer, M.L. and D.C. Adams. 2024. Interrogating Random and Systematic Measurement Error in Morphometric Data. Evolutionary Biology, 51, 179–20.

Bookstein, F.L., & Mitteroecker, P. (2014). Comparing covariance matrices by relative eigenanalysis, with applications to organismal biology. Evolutionary Biology, 41(2), 336-350.

See Also

`lm.rrpp.ws`, `manova.update`, `lr_test`

Examples

```
## Not run:
# Measurement error analysis on simulated data of fish shapes

data(fishy)

# Example two digitization replicates of the same research subjects
rep1 <- matrix(fishy$coords[1,], 11, 2, byrow = TRUE)
rep2 <- matrix(fishy$coords[61,], 11, 2, byrow = TRUE)
plot(rep1, pch = 16, col = gray(0.5, alpha = 0.5), cex = 2, asp = 1)
points(rep2, pch = 16, col = gray(0.2, alpha = 0.5), cex = 2, asp = 1)

# Analysis unconcerned with groups

ME1 <- measurement.error(
  Y = "coords",
  subjects = "subj",
  replicates = "reps",
  data = fishy)

anova(ME1)
ICCstats(ME1, subjects = "Subjects", with_in = "Systematic ME")
plot(ME1)

# Analysis concerned with groups

ME2 <- measurement.error(
  Y = "coords",
  subjects = "subj",
  replicates = "reps",
  groups = "groups",
  data = fishy)

anova(ME2)
ICCstats(ME2, subjects = "Subjects",
  with_in = "Systematic ME", groups = "groups")
P <- plot(ME2)
focusMEonSubjects(P, subjects = 18:20, shadow = TRUE)

## End(Not run)
```

`model.comparison` *Model Comparisons, in terms of the log-likelihood, covariance trace, or Z-score.*

Description

Function calculates either log-likelihoods or traces of covariance matrices for comparison with respect to parameter penalties, or calculates Z-scores from RRPP, which can be profiled across a gradient (predictor).

Usage

```
model.comparison(
  ...,
  type = c("cov.trace", "logLik", "Z"),
  predictor = NULL,
  tol = NULL,
  pc.no = NULL,
  gls.null = FALSE,
  mod.names = TRUE,
  verbose = FALSE
)
```

Arguments

`...`	Any number of lm.rrpp class objects for model fits to be compared.
`type`	An argument to choose between log-likelihood, covariance trace, or Z results. If Z is chosen, Z-scores are calculated, the same log-likelihoods are calculated as with the log-likelihood type, but also in every RRPP permutation, as describe for the initial mode fits, with choice of null model (below).
`predictor`	An optional vector that can be used to profile the results based on type across a range of numerical values described by the predictor. A spline will also be fit, which will reveal estimated values of the predictor that yield maximum and minimum values of model comparison metric.
`tol`	If type = logLik or Z, tol is a tolerance value between 0 and 1, indicating the magnitude below which components should be omitted (if standard deviations of components are less than the eigenvalue of the first component times the tolerance), for calculating the log-likelihood.
`pc.no`	If type = logLik or Z, an optional value to indicate the number of principal components (maximum rank) to use for calculating the log-likelihood.
`gls.null`	A logical value indicating whether GLS estimation should be used with the null (intercept) model, for calculating Z scores via RRPP of log-likelihoods. This should be FALSE if comparing different GLS estimations of covariance matrices. It should be TRUE if comparing different model fits with the same GLS-estimated covariance matrix.
`mod.names`	A logical value for whether to label models by their object names (TRUE) or formulas (FALSE).
`verbose`	A logical value for whether to include distributions of random log-likelihoods, if applicable.

Details

The function calculates either log-likelihoods or traces of (residual) covariance matrices, plus parameter penalties, to assist in comparative model evaluation or selection. Because high-dimensional data often produce singular or ill-conditioned residual covariance matrices, this function does one of two things: 1) uses the trace of a covariance matrix rather than its determinant; or 2) provides a ridge-regularization (Warton, 2008) of the covariance matrix, only if it is determined that it is

ill-conditioned. Regardless of implementation, covariance matrices are projected into a principal component (PC) space of appropriate dimensions.

The parameter penalty is based on that proposed by Bedrick and Tsai (1994), equal to 2(pk + p(p + 1)/2), where p is the appropriate dimension (not number of variables) of the covariance matrix. The parameter, k, is the rank of the model design matrix.

In the case that "logLik" is chosen for the argument, type, AIC scores are calculated. These scores may not perfectly match other packages or software that calculate AIC for multivariate data, if ridge regularization was used (and if other packages require p = the number of data variables). When choosing logLik as the type of comparison, it might be a good idea to adjust the tolerance or number of data principal components. The default (NULL) values will use all data dimensions to calculate log-likelihoods, which might cause problems if the number of variables exceeds the number of observations (producing singular residual covariance matrices). However, one should not reduce data dimensions haphazardly, as this can lead to poor estimates of log-likelihood. Furthermore, using the tolerance argument could result in different numbers of principal components used for each model to calculate log-likelihoods, which might be a concern for comparing models. If both tol and pc.no arguments are used, the solution will use the fewest PCs produced by either argument. Because the trace of a covariance matrix is not sensitive to matrix singularity, no PC adjustment is used for the cov.trace argument.

This function can also calculate Z-scores from RRPP on model log-likelihoods, which can be compared directly or profiled along a gradient (predictor). This might be useful for for comparing generalized least-squares (GLS) models, for example, along a gradient of a parameter used to scale the covariance matrix for GLS estimation. See Collyer et al. 2022 for an example of using RRPP on log-likelihoods with different covariance matrices.

Users can construct their own tables from the results but this function does not attempt to summarize results, as interpreting results requires some arbitrary decisions. The `anova` function explicitly tests multiple models and can be used for nested model comparisons.

Results can also be plotted using the generic `plot` function.

Value

An object of class `model.comparison` is a data frame with either log-likelihoods, covariance traces, or, Z-scores, plus parameter penalties. AIC scores might be included, if applicable. If verbose results are returned, random log-likelihoods are also included.

Author(s)

Michael Collyer

References

Bedrick, E.J., and C.L. Tsai. 1994. Model selection for multivariate regression in small samples. Biometrics, 226-231.

Warton, D.I., 2008. Penalized normal likelihood and ridge regularization of correlation and covariance matrices. Journal of the American Statistical Association. 103: 340-349.

Collyer, M.L., E.K. Baken, & D.C. Adams. A standardized effect size for evaluating and comparing the strength of phylogenetic signal. Methods in Ecology and Evolution. 13: 367–382.

Examples

```
## Not run:
data(Pupfish)
Pupfish$logSize <- log(Pupfish$CS)
fit1 <- lm.rrpp(coords ~ logSize, data = Pupfish, iter = 0,
print.progress = FALSE)
fit2 <- lm.rrpp(coords ~ Pop, data = Pupfish, iter = 0,
print.progress = FALSE)
fit3 <- lm.rrpp(coords ~ Sex, data = Pupfish, iter = 0,
print.progress = FALSE)
fit4 <- lm.rrpp(coords ~ logSize + Sex, data = Pupfish, iter = 0,
print.progress = FALSE)
fit5 <- lm.rrpp(coords ~ logSize + Pop, data = Pupfish, iter = 0,
print.progress = FALSE)
fit6 <- lm.rrpp(coords ~ logSize + Sex * Pop, data = Pupfish, iter = 0,
print.progress = FALSE)

modComp1 <- model.comparison(fit1, fit2, fit3, fit4, fit5,
fit6, type = "cov.trace")
modComp2 <- model.comparison(fit1, fit2, fit3, fit4, fit5,
fit6, type = "logLik", tol = 0.01)

summary(modComp1)
summary(modComp2)

par(mfcol = c(1,2))
plot(modComp1)
plot(modComp2)

# Comparing fits with covariance matrices
# an example for scaling a phylogenetic covariance matrix with
# the scaling parameter, lambda

data("PlethMorph")
Cov <- PlethMorph$PhyCov
lambda <- seq(0, 1, 0.1)

Cov1 <- scaleCov(Cov, scale. = lambda[1])
Cov2 <- scaleCov(Cov, scale. = lambda[2])
Cov3 <- scaleCov(Cov, scale. = lambda[3])
Cov4 <- scaleCov(Cov, scale. = lambda[4])
Cov5 <- scaleCov(Cov, scale. = lambda[5])
Cov6 <- scaleCov(Cov, scale. = lambda[6])
Cov7 <- scaleCov(Cov, scale. = lambda[7])
Cov8 <- scaleCov(Cov, scale. = lambda[8])
Cov9 <- scaleCov(Cov, scale. = lambda[9])
Cov10 <- scaleCov(Cov, scale. = lambda[10])
Cov11 <- scaleCov(Cov, scale. = lambda[11])


fit1 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov1)
fit2 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov2)
```

```
fit3 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov3)
fit4 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov4)
fit5 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov5)
fit6 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov6)
fit7 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov7)
fit8 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov8)
fit9 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov9)
fit10 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov10)
fit11 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov11)

par(mfrow = c(1,1))

MC1 <- model.comparison(fit1, fit2, fit3, fit4, fit5, fit6,
fit7, fit8, fit9, fit10, fit11,
type = "logLik")
MC1
plot(MC1)

MC2 <- model.comparison(fit1, fit2, fit3, fit4, fit5, fit6,
fit7, fit8, fit9, fit10, fit11,
type = "logLik", predictor = lambda)
MC2
plot(MC2)


MC3 <- model.comparison(fit1, fit2, fit3, fit4, fit5, fit6,
fit7, fit8, fit9, fit10, fit11,
type = "Z", predictor = lambda)
MC3
plot(MC3)

## End(Not run)
```

`model.frame.lm.rrpp` *Extract model frame from a lm.rrpp object*

Description

`model.frame.lm.rrpp` returns the model frame constructed for an `lm.rrpp` object.

Usage

```
## S3 method for class 'lm.rrpp'
model.frame(formula, ...)
```

Arguments

`formula`	Object from `lm.rrpp`
`...`	further arguments passed to or from other methods

Author(s)

Michael Collyer

`model.matrix.lm.rrpp` *Extract the model design matrix from an lm.rrpp object*

Description

`model.matrix.lm.rrpp` returns the design matrix constructed for an `lm.rrpp` object.

Usage

```
## S3 method for class 'lm.rrpp'
model.matrix(object, ...)
```

Arguments

`object`	Object from `lm.rrpp`
`...`	further arguments passed to or from other methods

Author(s)

Michael Collyer

`motionpaths` *Simulated motion paths*

Description

Simulated motion paths

Author(s)

Dean Adams

References

Adams, D. C., and M. L. Collyer. 2009. A general framework for the analysis of phenotypic trajectories in evolutionary studies. Evolution 63:1143-1154.

na.omit.rrpp.data.frame *Handle missing values in rrpp.data.frame objects*

Description

Handle missing values in rrpp.data.frame objects

Usage

```
## S3 method for class 'rrpp.data.frame'
na.omit(object, ...)
```

Arguments

object	object (from rrpp.data.frame)
...	further arguments (currently not used)

Author(s)

Michael Collyer

Examples

```
y <- matrix(rnorm(15), 5, 3)
x <- rnorm(5)
rdf <- rrpp.data.frame(x = x, y = y, d = dist(y))
rdf$x[1] <- NA # create missing data
rdf

ndf <- na.omit(rdf)
ndf
```

ordinate *Ordination tool for data aligned to another matrix*

Description

Function performs a singular value decomposition of ordinary least squares (OLS) or generalized least squares (GLS) residuals, aligned to an alternative matrix, plus projection of data onto vectors obtained.

Usage

```
ordinate(
  Y,
  A = NULL,
  Cov = NULL,
  transform. = TRUE,
  scale. = FALSE,
  tol = NULL,
  rank. = NULL,
  newdata = NULL
)
```

Arguments

Y	An n x p data matrix.
A	An optional n x n symmetric matrix or an n x k data matrix, where k is the number of variables that could be associated with the p variables of Y. If NULL, an n x n identity matrix will be used.
Cov	An optional n x n covariance matrix to describe the non-independence among observations in Y, and provide a GLS-centering of data. Note that Cov and A can be the same, if one wishes to align GLS residuals to the same matrix used to obtain them. Note also that no explicit GLS-centering is performed on A. If this is desired, A should be GLS-centered beforehand.
transform.	An optional argument if a covariance matrix is provided to transform GLS-centered residuals, if TRUE. If FALSE, only GLS-centering is performed. Only if transform = TRUE (the default) can one expect the variances of ordinate scores in a principal component analysis to match eigenvalues.
scale.	a logical value indicating whether the variables should be scaled to have unit variance before the analysis takes place. The default is FALSE.
tol	A value indicating the magnitude below which components should be omitted. (Components are omitted if their standard deviations are less than or equal to tol times the standard deviation of the first component.) With the default null setting, no components are omitted (unless rank. is provided). Other settings for tol could be tol = sqrt(.Machine$double.eps), which would omit essentially constant components, or tol = 0, to retain all components, even if redundant. This argument is exactly the same as in prcomp
rank.	Optionally, a number specifying the maximal rank, i.e., maximal number of aligned components to be used. This argument can be set as alternative or in addition to tol, useful notably when the desired rank is considerably smaller than the dimensions of the matrix. This argument is exactly the same as in prcomp
newdata	An optional data frame of values for the same variables of Y to be projected onto aligned components. This is only possible with OLS (transform. = FALSE).

Details

The function performs a singular value decomposition, **t(A)Z = UDt(V)**, where **Z** is a matrix of residuals (obtained from **Y**; see below) and **A** is an alignment matrix with the same number of rows

as **Z**. (**t** indicates matrix transposition.) **U** and **V** are the matrices of left and right singular vectors, and **D** is a diagonal matrix of singular values. **V** are the vectors that describe maximized covariation between **Y** and **A**. If **A** = **I**, an n x n identity matrix, **V** are the eigen vectors (principal components) of **Y**.

Z represents a centered and potentially standardized form of **Y**. This function can center data via OLS or GLS means (the latter if a covariance matrix to describe the non-independence among observations is provided). If standardizing variables is preferred, then **Z** both centers and scales the vectors of **Y** by their standard deviations.

Data are projected onto aligned vectors, **ZV**. If a GLS computation is made, the option to transform centered values (residuals) before projection is available. This is required for orthogonal projection, but from a transformed data space. Not transforming residuals maintains the Euclidean distances among observations and the OLS multivariate variance, but the projection is oblique (scores can be correlated).

The versatility of using an alignment approach is that alternative data space rotations are possible. Principal components are thus the vectors that maximize variance with respect to the data, themselves, but "components" of (co)variation can be described for any inter-matrix relationship, including phylogenetic signal, ecological signal, ontogenetic signal, size allometry, etc. More details are provided in Collyer and Adams (2021).

Much of this function is consistent with the `prcomp` function, except that centering data is not an option (it is required).

SUMMARY STATISTICS: For principal component plots, the traditional statistics to summarize the analysis include eigenvalues (variance by component), proportion of variance by component, and cumulative proportion of variance. When data are aligned to an alternative matrix, the statistics are less straightforward. A summary of of such an analysis (performed with `summary.ordinate`) will produce these additional statistics:

Singular Value Rather than eigenvalues, the singular values from singular value decomposition of the cross-product of the scaled alignment matrix and the data.

Proportion of Covariance Each component's singular value divided by the sum of singular values. The cumulative proportion is also returned. Note that these values do not explain the amount of covariance between the alignment matrix and data, but explain the distribution of the covariance. Large proportions can be misleading.

RV by Component The partial RV statistic by component. Cumulative values are also returned. The sum of partial RVs is Escoffier's RV statistic, which measures the amount of covariation between the alignment matrix and data. Caution should be used in interpreting these values, which can vary with the number of observations and number of variables. However, the RV is more reliable than proportion of singular value for interpretation of the strength of linear association for aligned components. (It is most analogous to proportion of variance for principal components.)

Value

An object of class `ordinate` is a list containing the following

`x`	Aligned component scores for all observations
`xn`	Optional projection of new data onto components.
`d`	The portion of the squared singular values attributed to the aligned components.

sdev	Standard deviations of d; i.e., the scale of the components.
rot	The matrix of variable loadings, i.e. the singular vectors, **V**.
center	The OLS or GLS means vector used for centering.
transform	Whether GLS transformation was used in projection of residuals (only possible in conjunction with GLS-centering).
scale	The scaling used, or FALSE.
alignment	Whether data were aligned to principal axes or the name of another matrix.
GLS	A logical value to indicate if GLS-centering and projection was used.

Author(s)

Michael Collyer

References

Collyer, M.L. and D.C. Adams. 2021. Phylogenetically-aligned Component Analysis. Methods in Ecology and evolution. In press.

Revell, L. J. 2009. Size-correction and principal components for interspecific comparative studies. Evolution, 63:3258-3268.

See Also

plot.ordinate, prcomp, plot.default, gm.prcomp within geomorph

Examples

```
# Examples use residuals from a regression of salamander
# morphological traits against body size (snout to vent length, SVL).
# Observations are species means and a phylogenetic covariance matrix
# describes the relatedness among observations.

data("PlethMorph")
Y <- as.data.frame(PlethMorph[c("TailLength", "HeadLength",
"Snout.eye", "BodyWidth",
"Forelimb", "Hindlimb")])
Y <- as.matrix(Y)
R <- lm.rrpp(Y ~ SVL, data = PlethMorph,
iter = 0, print.progress = FALSE)$LM$residuals

# PCA (on correlation matrix)

PCA.ols <- ordinate(R, scale. = TRUE)
PCA.ols$rot
prcomp(R, scale. = TRUE)$rotation # should be the same

# phyPCA (sensu Revell, 2009)
# with projection of untransformed residuals (Collyer & Adams 2020)

PCA.gls <- ordinate(R, scale. = TRUE,
```

```
transform. = FALSE,
Cov = PlethMorph$PhyCov)

# phyPCA with transformed residuals (orthogonal projection,
# Collyer & Adams 2020)

PCA.t.gls <- ordinate(R, scale. = TRUE,
transform. = TRUE,
Cov = PlethMorph$PhyCov)

 # Align to phylogenetic signal (in each case)

 PaCA.ols <- ordinate(R, A = PlethMorph$PhyCov, scale. = TRUE)

 PaCA.gls <- ordinate(R, A = PlethMorph$PhyCov,
 scale. = TRUE,
 transform. = FALSE,
 Cov = PlethMorph$PhyCov)

 PaCA.t.gls <- ordinate(R, A = PlethMorph$PhyCov,
 scale. = TRUE,
 transform. = TRUE,
 Cov = PlethMorph$PhyCov)

 # Summaries

 summary(PCA.ols)
 summary(PCA.gls)
 summary(PCA.t.gls)
 summary(PaCA.ols)
 summary(PaCA.gls)
 summary(PaCA.t.gls)

 # Plots

 par(mfrow = c(2,3))
 plot(PCA.ols, main = "PCA OLS")
 plot(PCA.gls, main = "PCA GLS")
 plot(PCA.t.gls, main = "PCA t-GLS")
 plot(PaCA.ols, main = "PaCA OLS")
 plot(PaCA.gls, main = "PaCA GLS")
 plot(PaCA.t.gls, main = "PaCA t-GLS")
 par(mfrow = c(1,1))

 # Changing some plot aesthetics (the arguments in plot.ordinate and
 # plot.default are important for changing plot parameters)

 P1 <- plot(PaCA.gls, main = "PaCA GLS", include.axes = TRUE)

 P2 <- plot(PaCA.gls, main = "PaCA GLS", include.axes = TRUE,
 col = 4, pch = 21, bg = PlethMorph$group)
 add.tree(P2, PlethMorph$tree, edge.col = 4)
```

```
P3 <- plot(PaCA.gls, main = "PaCA GLS", include.axes = TRUE,
col = 4, pch = 21, bg = PlethMorph$group,
flip = 1)
add.tree(P3, PlethMorph$tree, edge.col = 4)
```

`pairwise` *Pairwise comparisons of lm.rrpp fits*

Description

Function generates distributions of pairwise statistics for a lm.rrpp fit and returns important statistics for hypothesis tests.

Usage

```
pairwise(
  fit,
  fit.null = NULL,
  groups,
  covariate = NULL,
  verbose = FALSE,
  print.progress = FALSE
)
```

Arguments

`fit`	A linear model fit using `lm.rrpp`.
`fit.null`	An alternative linear model fit to use as a null model for RRPP, if the null model of the fit is not desired. Note, for FRPP this argument should remain NULL and FRPP must be established in the lm.rrpp fit (RRPP = FALSE). If the null model is uncertain, using `reveal.model.designs` will help elucidate the inherent null model used.
`groups`	A factor or vector that is coercible into a factor, describing the levels of the groups for which to find LS means or slopes. Normally this factor would be part of the model fit, but it is not necessary for that to be the case in order to obtain results.
`covariate`	A numeric vector for which to calculate slopes for comparison If NULL, LS means will be calculated instead of slopes. Normally this variable would be part of the model fit, but it is not necessary for that to be the case in order to obtain results.
`verbose`	A logical value for whether to include extra results, specifically the Mahalanobis distances among means, which requires the calculation of residual covariance matrices for each permutation. This should be generally FALSE, unless Mahalanobis distances are desired, in which case it must be TRUE. Verbose computations can require much additional time.

`print.progress` If a null model fit is provided, a logical value to indicate whether analytical results progress should be printed on screen. Unless large data sets are analyzed, this argument is probably not helpful.

Details

Based on an lm.rrpp fit, this function will find fitted values over all permutations and based on a grouping factor, calculate either least squares (LS) means or slopes, and pairwise statistics among them. Pairwise statistics have multiple flavors, related to vector attributes:

Distance between vectors, "dist" Vectors for LS means or slopes originate at the origin and point to some location, having both a magnitude and direction. A distance between two vectors is the inner-product of of the vector difference, i.e., the distance between their endpoints. For LS means, this distance is the difference between means. For multivariate slope vectors, this is the difference in location between estimated change for the dependent variables, per one-unit change of the covariate considered. For univariate slopes, this is the absolute difference between slopes.

Standardized distance between vectors, "stdist" Same as the distance between vectors, but distances are divided by the model standard error (square-root of the trace of the residual covariance matrix, adjusted by sample size). Pairwise tests with this statistic should be consistent with ANOVA results.

Mahalanobis distance between vectors, "mdist" Similar to the standardized distance between vectors but the inverse of the residual covariance matrix is used in calculation of the distance, rather than dividing the Euclidean distance between means and dividing by the standard error. If the residual covariance matrix is singular, Mahalanobis distances will not be calculated. Pairwise tests with this statistic should be consistent with MANOVA results.

Vector correlation, "VC" If LS mean or slope vectors are scaled to unit size, the vector correlation is the inner-product of the scaled vectors. The arccosine (acos) of this value is the angle between vectors, which can be expressed in radians or degrees. Vector correlation indicates the similarity of vector orientation, independent of vector length.

Difference between vector lengths, "DL" If the length of a vector is an important attribute – e.g., the amount of multivariate change per one-unit change in a covariate – then the absolute value of the difference in vector lengths is a practical statistic to compare vector lengths, rather than the estimates the vectors make. Let d1 and d2 be the distances (length) of vectors. Then |d1 - d2| is a statistic that compares their lengths. For slope vectors, this is a comparison of rates. For comparison, if vectors are rates, "dist" finds the difference between estimates per unit change of, e.g., time, size, etc., which could be large, even for small rates of change, if vectors point in dissimilar directions. "DL" is a comparison of rates, irrespective of direction.

Variance, "var" Vectors of residuals from a linear model indicate can express the distances of observed values from fitted values. Mean squared distances of values (variance), by group, can be used to measure the amount of dispersion around estimated values for groups. Absolute differences between variances are used as test statistics to compare mean dispersion of values among groups. Variance degrees of freedom equal n, the group size, rather than n-1, as the purpose is to compare mean dispersion in the sample. (Additionally, tests with one subject in a group are possible, or at least not a hindrance to the analysis.)

The `summary.pairwise` function is used to select a test statistic for the statistics described above, as "dist", "VC", "DL", and "var", respectively. If vector correlation is tested, the `angle.type` argument can be used to choose between radians and degrees.

The null model is defined via lm.rrpp, but one can also use an alternative null model as an optional argument. In this case, residual randomization in the permutation procedure (RRPP) will be performed using the alternative null model to generate fitted values. If full randomization of values (FRPP) is preferred, it must be established in the lm.rrpp fit and an alternative model should not be chosen. If one is unsure about the inherent null model used if an alternative is not specified as an argument, the function reveal.model.designs can be used.

Observed statistics, effect sizes, P-values, and one-tailed confidence limits based on the confidence requested will be summarized with the summary.pairwise function. Confidence limits are inherently one-tailed as the statistics are similar to absolute values. For example, a distance is analogous to an absolute difference. Therefore, the one-tailed confidence limits are more akin to two-tailed hypothesis tests. (A comparable example is to use the absolute value of a t-statistic, in which case the distribution has a lower bound of 0.)

Notes for RRPP 0.6.2 and subsequent versions:

In previous versions of pairwise, summary.pairwise had three test types: "dist", "VC", and "var". When one chose "dist", for LS mean vectors, the statistic was the inner-product of the vector difference. For slope vectors, "dist" returned the absolute value of the difference between vector lengths, which is "DL" in 0.6.2 and subsequent versions. This update uses the same calculation, irrespective of vector types. Generally, "DL" is the same as a contrast in rates for slope vectors, but might not have much meaning for LS means. Likewise, "dist" is the distance between vector endpoints, which might make more sense for LS means than slope vectors. Nevertheless, the user has more control over these decisions with version 0.6.2 and subsequent versions.

Notes for RRPP 2.0.4 and subsequent versions:

The test types, standardized distance between vectors, "stdist", and Mahalanobis distances between vectors were added. The former simply divides the distance between vectors by model standard error (square-root of the trace of the residual covariance matrix, adjusted by sample size). This is a multivariate generalization of a t-statistic for the difference between means. It is not the same as Hotelling squared-T-statistic, which requires incorporating the inverse of the residual covariance matrix in the calculation of the distance, a calculation that also requires a non-singular covariance matrix. However, the Mahalanobis distances are similar (and proportional) to the Hotelling squared-T-statistic. Pairwise tests using Mahalanobis distances represent a non-parametric analog to the parametric Hotelling squared-T test. Both tests should be better for GLS model fits compared to Euclidean distances between means, as the total sums of squares are more likely to vary across random permutations. In general, if ANOVA is performed a pairwise test with "stdist" is a good association; if MANOVA is performed, a pairwise test with "mdist" is a good association.

Notes for RRPP 2.2.0 and subsequent versions:

It is possible to adjust the P-values with p.adjust when using summary.pairwise. However, one should use extreme caution because probabilities calculated with permutations are left-censored by the number of permutations. For example, the smallest P-value using 1,000 permutations is 0.001, even if the true P-value is much smaller. Using P-value adjustments is fraught with inferential errors unless one uses a very large number of permutations.

Alternatively, one can adjust the confidence level in summary.pairwise, which is much safer. For example, one might use a 99.5 (confidence = 0.995), thus finding confidence limits for statistics for which only 0.5 percent of values are larger. This is a safe option, regardless of the number of permutations.

Value

An object of class `pairwise` is a list containing the following

`LS.means`	LS means for groups, across permutations.
`slopes`	Slopes for groups, across permutations.
`means.dist`	Pairwise distances between means, across permutations.
`std.means.dist`	Pairwise distances between means, across permutations, standardized.
`mah.means.dist`	Pairwise Mahalanobis distances between means, across permutations.
`means.vec.cor`	Pairwise vector correlations between means, across permutations.
`means.lengths`	LS means vector lengths, by group, across permutations.
`means.diff.length`	Pairwise absolute differences between mean vector lengths, across permutations.
`slopes.dist`	Pairwise distances between slopes (end-points), across permutations.
`slopes.vec.cor`	Pairwise vector correlations between slope vectors, across permutations.
`slopes.lengths`	Slope vector lengths, by group, across permutations.
`slopes.diff.length`	Pairwise absolute differences between slope vector lengths, across permutations.
`n`	Sample size
`p`	Data dimensions; i.e., variable number
`PermInfo`	Information for random permutations, passed on from lm.rrpp fit and possibly modified if an alternative null model was used.

Author(s)

Michael Collyer

References

Collyer, M.L., D.J. Sekora, and D.C. Adams. 2015. A method for analysis of phenotypic change for phenotypes described by high-dimensional data. Heredity. 115:357-365.

Adams, D.C and M.L. Collyer. 2018. Multivariate phylogenetic ANOVA: group-clade aggregation, biological challenges, and a refined permutation procedure. Evolution. In press.

See Also

`lm.rrpp`

Examples

```
## Not run:
# Examples use geometric morphometric data on pupfishes
# See the package, geomorph, for details about obtaining such data

# Body Shape Analysis (Multivariate) --------------
```

```
data("Pupfish")

# Note:

dim(Pupfish$coords) # highly multivariate!

Pupfish$logSize <- log(Pupfish$CS)

# Note: one should use all dimensions of the data but with this
# example, there are many.  Thus, only three principal components
# will be used for demonstration purposes.

Pupfish$Y <- ordinate(Pupfish$coords)$x[, 1:3]

## Pairwise comparisons among LS means

# Note: one should increase RRPP iterations but a
# smaller number is used here for demonstration
# efficiency.  Generally, iter = 999 will take less
# than 1s for these examples with a modern computer.

fit1 <- lm.rrpp(Y ~ logSize + Sex * Pop, SS.type = "I",
data = Pupfish, print.progress = FALSE, iter = 199)
summary(fit1, formula = FALSE)
anova(fit1)

pup.group <- interaction(Pupfish$Sex, Pupfish$Pop)
pup.group
PW1 <- pairwise(fit1, groups = pup.group)
PW1

# distances among means
summary(PW1, confidence = 0.95, test.type = "dist")
summary(PW1, confidence = 0.95, test.type = "dist", stat.table = FALSE)

# using P-value adjustment
summary(PW1, confidence = 0.95, test.type = "dist",
p.adjust.method = "bonferroni")
summary(PW1, confidence = 0.95,
test.type = "dist", stat.table = FALSE,
p.adjust.method = "bonferroni")

# using 99% confidence
summary(PW1, confidence = 0.99, test.type = "dist")
summary(PW1, confidence = 0.99, test.type = "dist", stat.table = FALSE)

# standardized distances among means
summary(PW1, confidence = 0.95, test.type = "stdist")

# Mahalanobis (generalized) distances among means
summary(PW1, confidence = 0.95, test.type = "mdist")
```

```
# absolute difference between mean vector lengths
summary(PW1, confidence = 0.95, test.type = "DL")

# correlation between mean vectors (angles in degrees)
summary(PW1, confidence = 0.95, test.type = "VC",
   angle.type = "deg")

# Can also compare the dispersion around means
summary(PW1, confidence = 0.95, test.type = "var")

## Pairwise comparisons of slopes

fit2 <- lm.rrpp(Y ~ logSize * Sex * Pop, SS.type = "I",
data = Pupfish, print.progress = FALSE, iter = 199)
summary(fit2, formula = FALSE)
anova(fit1, fit2)

# Using a null fit that excludes all factor-covariate
# interactions, not just the last one

PW2 <- pairwise(fit2, fit.null = fit1, groups = pup.group,
covariate = Pupfish$logSize, print.progress = FALSE)
PW2

# distances between slope vectors (end-points)
summary(PW2, confidence = 0.95, test.type = "dist")
summary(PW2, confidence = 0.95, test.type = "dist", stat.table = FALSE)

# absolute difference between slope vector lengths
summary(PW2, confidence = 0.95, test.type = "DL")

# correlation between slope vectors (and angles)
summary(PW2, confidence = 0.95, test.type = "VC",
   angle.type = "deg")

# Can also compare the dispersion around group slopes
summary(PW2, confidence = 0.95, test.type = "var")

## End(Not run)
```

`pairwise.model.Z` *Pairwise comparisons of model effects*

Description

Function generates pairwise statistics for comparing model fits and returns important statistics for hypothesis tests.

Usage

```
pairwise.model.Z(
  ...,
  nsamp = NULL,
  two.tailed = TRUE,
  predictor = NULL,
  tol = NULL,
  pc.no = NULL,
  gls.null = FALSE,
  p.adjust.method = "none"
)
```

Arguments

...	Either an object of class model.comparison, or several objects of class lm.rrpp. If the former, arguments of type = 'Z' and verbose = TRUE are required. If the latter, a model comparison analysis will first be performed with these arguments.
nsamp	An optional vector containing the sample sizes for each model fit
two.tailed	A logical value to indicate whether a two-tailed test (typical and default) should be performed.
predictor	An optional vector to be passed to model.comparison, if used.
tol	An optional value to be passed to model.comparison, if used.
pc.no	An optional value to be passed to model.comparison, if used.
gls.null	An optional logical value to be passed to model.comparison, if used.
p.adjust.method	Optional logical value to adjust P-values, based on p.adjust. Defaults to "none".

Details

The function statistically compares the effect sizes of two or more models fit and evaluated using RRPP. Input for the function is a list of fitted models of the class 'model.comparison', whose options included type = 'Z' and verbose = TRUE when the models were compared with that function.

A two-sample test is performed on each pair of models, comparing the strength of model fits to one another (Collyer and Adams 2025). This might be used to compare the strength of fit of the data to differing statistical models (as in model selection) or for comparing the fit across differing datasets for the same model to determine whether the strength of fit in one dataset is greater than that found in another (see Collyer and Adams 2025). In the latter case, one is advised to include a vector containing the sample sizes of each dataset, so that two-sample tests may account for differences in sample size.

Value

A list containing the following

sample.z	A vector of model effect sizes.
pairwise.z	A matrix of pairwise test statistics comparing model effect sizes.

pairwise.P	A matrix of pairwise significance levels.
tails	Number of tails used for P-value calculation.

Author(s)

Dean Adams and Michael Collyer

References

Collyer, M.L., and D.C. Adams. 2025. Permutational Biometry. Volume 1: Univariate Data. Iowa State University Digital Press. (Forthcoming).

Examples

```
## Not run:
 data(Pupfish)
 Pupfish$logSize <- log(Pupfish$CS)
 fit1 <- lm.rrpp(coords ~ logSize, data = Pupfish,
 print.progress = FALSE)
 fit2 <- lm.rrpp(coords ~ Pop, data = Pupfish,
 print.progress = FALSE)
 fit3 <- lm.rrpp(coords ~ Sex, data = Pupfish,
 print.progress = FALSE)
 fit6 <- lm.rrpp(coords ~ logSize + Sex * Pop, data = Pupfish,
 print.progress = FALSE)
 Mod.C <- model.comparison(fit1, fit2, fit3, fit6,
 pc.no = 4, type = "Z", verbose = TRUE)
 res <- pairwise.model.Z(Mod.C)
 summary(res, stats.table = TRUE)
 summary(res, stats.table = FALSE)

## End(Not run)
```

---

phytoplankton *Phytoplankton Biomass and Eutrophication*

Description

A data set containing variables for phytoplanton biomass, phosphorous levels, trophic status, and locations (coordinates) of 70 temperate lakes.

Details

The data in this data set are samples from a larger study that compared phytoplankton biomass from temperate lakes in Alberta, Canada, with different levels of eutrophication (Loewen et al., 2020). These data are suitable for considering the spatial non-independence of observations in generalized least squares (GLS) estimation, for the example in the book, Permutational Biometry (Collyer & Adams, 2025).

References

Loewen, C. J. G., Wyatt, F. R., Mortimer, C. A., Vinebrooke, R. D., and Zurawell, R. W. (2020). Multiscale drivers of phytoplankton communities in north-temperate lakes. Ecological Applications, 30(5):e02102.

Collyer, M. L., & Adams D. C. (2025). Permutational Biometry: Volume 1, Univariate Data. Iowa State University Digital Press. NOT YET PUBLISHED.

`PlethCinHoff` *Plethodon comparative SVL data*

Description

Snout-to-vent-length (SVL) data for two species of salamanders

Details

The data set is a subset of data from two species of salamanders (Plethodon cinereus and P. hoffmani) in eastern North America (based on Adams and Rohlf, 2000; Adams, 2000), collected from the Smithsonian Museum (Washington DC, USA). Data include species, locality (whether in sympatry or allopatry), and SVL in mm. These data are suitable data for demonstrating single factor and factorial analysis of variance (ANOVA). (ANCOVA), used in the book, Permutational Biometry (Collyer & Adams, 2025).

Author(s)

Dean Adams

References

Adams, D. C. (2000). Divergence of trophic morphology and resource use among populations of Plethodon cinereus and P. hoffmani in Pennsylvania. In Bruce, R. C., Jaeger, R. G., and Houck, L. D., editors, The Biology of Plethodontid Salamanders, pages 383–394. Springer US.

Adams, D. C. and Rohlf, F. J. (2000). Ecological character displacement in Plethodon: Biomechanical differences found from a geometric morphometric study. Proceedings of the National Academy of Sciences, 97:4106–4111.

Collyer, M. L., & Adams D. C. (2025). Permutational Biometry: Volume 1, Univariate Data. Iowa State University Digital Press. NOT YET PUBLISHED.

PlethMorph *Plethodon comparative morphological data*

Description

Data for 37 species of plethodontid salamanders. Variables include snout to vent length (SVL) as species size, tail length, head length, snout to eye length, body width, forelimb length, and hind limb length, all measured in mm. A grouping variable is also included for functional guild size. A variable for species names is also included. The data set also includes a phylogenetic covariance matrix based on a Brownian model of evolution, to assist in generalized least squares (GLS) estimation.

Details

The covariance matrix was estimated with the vcv.phylo function of the R package, ape, based on the tree described in Adams and Collyer (2018).

Author(s)

Michael Collyer and Dean Adams

References

Adams, D.C and Collyer, M.L. 2018. Multivariate phylogenetic anova: group-clade aggregation, biological challenges, and a refined permutation procedure. Evolution, 72: 1204-1215.

plot.interSubVar *Plot Function for RRPP*

Description

This function produces a heat map for inter-subject variability, based on results from a `measurement.error` object. The function, `interSubVar`, must first be used on the `measurement.error` object to obtain variability statistics. This function use the `image` function to produce plots. It does little to manipulate such plots, but any argument for `image` can be manipulated here, as well as the graphical parameters that can be adjusted within `image`.

Usage

```
## S3 method for class 'interSubVar'
plot(x, ...)
```

Arguments

`x`	Object from `interSubVar`
`...`	Arguments passed onto `image` and `plot`.

Author(s)

Michael Collyer

`plot.kcomp` *Plot Function for RRPP*

Description

Plot Function for RRPP

Usage

```
## S3 method for class 'kcomp'
plot(x, axis1 = 1, axis2 = 2, flip = NULL, include.axes = TRUE, ...)
```

Arguments

`x`	An object of class `kcomp`
`axis1`	A value indicating which component should be displayed as the X-axis (default = K1)
`axis2`	A value indicating which component should be displayed as the Y-axis (default = K2)
`flip`	An argument that if not NULL can be used to flip components in the plot. The values need to match axis1 or axis2. For example, if axis1 = 3 and axis2 = 4, flip = 1 will not change either axis; flip = 3 will flip only the horizontal axis; flip = c(3, 4) will flip both axes.
`include.axes`	A logical argument for whether axes should be shown at x = 0 and y = 0. This is different than the axes argument in the generic `plot.default` function, which controls the edges of the plot (providing a box effect or not). Using include.axes = TRUE does not allow aesthetic control of the axes. If desired, it is better to use include.axes = FALSE and augment the plot object with `abline` (choosing h = 0 and v = 0 in separate applications).
`...`	other arguments passed to plot (helpful to employ different colors or symbols for different groups). See

Value

An object of class "plot.kcomp" is a list with components that can be used in other plot functions, such as the type of plot, points, a group factor, and other information depending on the plot parameters used.

Author(s)

Michael Collyer

plot.lm.rrpp *Plot Function for RRPP*

Description

Plot Function for RRPP

Usage

```
## S3 method for class 'lm.rrpp'
plot(
  x,
  type = c("diagnostics", "regression", "PC"),
  resid.type = c("p", "n"),
  fitted.type = c("o", "t"),
  predictor = NULL,
  reg.type = c("PredLine", "RegScore"),
  ...
)
```

Arguments

x	plot object (from lm.rrpp)
type	Indicates which type of plot, choosing among diagnostics, regression, or principal component plots. Diagnostic plots are similar to lm diagnostic plots, but for multivariate data. Regression plots plot multivariate dispersion in some fashion against predictor values. PC plots project data onto the eigenvectors of the covariance matrix for fitted values.
resid.type	If type = "diagnostics", an optional argument for whether Pearson ("p") or normalized ("n") residuals should be used. These residuals are the same for ordinary least-squares (OLS) estimation but differ for generalized least-squares (GLS) estimation. For the latter, normalizing residuals requires multiplying them by the transformation matrix obtained for GLS estimation.
fitted.type	As with resid.type, whether fitted values use observed ("o") or transformed ("t") values.
predictor	An optional vector if "regression" plot type is chosen, and is a variable likely used in lm.rrpp. This vector is a vector of covariate values equal to the number of observations.
reg.type	If "regression" is chosen for plot type, this argument indicates whether prediction line (PredLine) or regression score (RegScore) plotting is performed. For explanation of prediction line, see Adams and Nistri (2010). For explanation of regression score, see Drake and Klingenberg (2008).
...	other arguments passed to plot (helpful to employ different colors or symbols for different groups). See plot.default and par

Author(s)

Michael Collyer

References

Drake, A. G., and C. P. Klingenberg. 2008. The pace of morphological change: Historical transformation of skull shape in St Bernard dogs. Proc. R. Soc. B. 275:71-76.

Adams, D. C., and A. Nistri. 2010. Ontogenetic convergence and evolution of foot morphology in European cave salamanders (Family: Plethodontidae). BMC Evol. Biol. 10:1-10.

Examples

```
## Not run:
# Univariate example
data(PlethMorph)
fitGLS <- lm.rrpp(TailLength ~ SVL, data = PlethMorph, Cov = PlethMorph$PhyCov,
 print.progress = FALSE, iter = 0)

 par(mfrow = c(2, 2))
 plot(fitGLS)
 plot(fitGLS, resid.type = "n") # use normalized (transformed) residuals
 plot(fitGLS, resid.type = "n", fitted.type = "t") # use also transformed fitted values

 # Multivariate example

Y <- as.matrix(cbind(PlethMorph$TailLength,
PlethMorph$HeadLength,
PlethMorph$Snout.eye,
PlethMorph$BodyWidth,
PlethMorph$Forelimb,
PlethMorph$Hindlimb))
PlethMorph$Y <- Y
fitGLSm <- lm.rrpp(Y ~ SVL, data = PlethMorph,
Cov = PlethMorph$PhyCov,
print.progress = FALSE, iter = 0)

par(mfrow = c(2, 2))
 plot(fitGLSm)
 plot(fitGLSm, resid.type = "n") # use normalized (transformed) residuals
 plot(fitGLSm, resid.type = "n", fitted.type = "t") # use also transformed fitted values
 par(mfrow = c(1, 1))

## End(Not run)
```

---

plot.looCV *Plot Function for RRPP*

Description

Plot Function for RRPP

Usage

```
## S3 method for class 'looCV'
plot(x, axis1 = 1, axis2 = 2, flip = NULL, ...)
```

Arguments

x	An object of class looCV
axis1	A value indicating which component should be displayed as the X-axis (default = C1)
axis2	A value indicating which component should be displayed as the Y-axis (default = C2)
flip	An argument that if not NULL can be used to flip components in the plot. The values need to match axis1 or axis2. For example, if axis1 = 3 and axis2 = 4, flip = 1 will not change either axis; flip = 3 will flip only the horizontal axis; flip = c(3, 4) will flip both axes. Axis will only be flipped in first plot.
...	other arguments passed to plot (helpful to employ different colors or symbols for different groups). See

Author(s)

Michael Collyer

---

plot.measurement.error

Plot Function for RRPP

Description

This function produces multivariate signal-to-noise ratio plots for measurement.error objects. See the function, plot.interSubVar for plotting the inter-subject variability from a measurement.error object, after applying the function, interSubVar.

Usage

```
## S3 method for class 'measurement.error'
plot(
  x,
  separate.by.groups = TRUE,
  add.connectors = TRUE,
  add.labels = FALSE,
  use.std.vectors = FALSE,
  titles = NULL,
  add.legend = TRUE,
  ...
)
```

Arguments

x	Object from measurement.error
separate.by.groups	A logical value for whether to make separate plots for each group, if different groups are available. If FALSE, groups are still represented by different symbols in the plot, unless overridden by plot arguments.
add.connectors	A logical value for whether to add connectors, like vectors, between replicate observations of the same subjects.
add.labels	A logical value for whether to label subjects. Labels are either subject name (if available) or number of occurrence in the data set.
use.std.vectors	A logical value for whether to use vectors obtained from a standardized matrix, which are orthogonal. This is not strictly necessary.
titles	An optional vector or list for augmenting the titles of plots produced. The length of the vector or list should match the number of plots produced by other arguments.
add.legend	A logical value for whether to add a legend to plots. If separate.by.groups is TRUE, adding a legend to plots will be slightly redundant. If certain parameters are augmented by user (point characters, colors), add.legend will be made to be FALSE to prevent misinterpretation of intended plotting scheme.
...	Other arguments passed onto plot

Author(s)

Michael Collyer

---

plot.model.comparison *Plot Function for RRPP*

Description

Plot Function for RRPP

Usage

```
## S3 method for class 'model.comparison'
plot(x, ...)
```

Arguments

x	plot object (from model.comparison)
...	other arguments passed to plot (helpful to employ different colors or symbols for different groups). See plot.default and par

Author(s)

Michael Collyer

plot.ordinate *Plot Function for RRPP*

Description

Plot Function for RRPP

Usage

```
## S3 method for class 'ordinate'
plot(x, axis1 = 1, axis2 = 2, flip = NULL, include.axes = TRUE, ...)
```

Arguments

x	An object of class ordinate
axis1	A value indicating which component should be displayed as the X-axis (default = C1)
axis2	A value indicating which component should be displayed as the Y-axis (default = C2)
flip	An argument that if not NULL can be used to flip components in the plot. The values need to match axis1 or axis2. For example, if axis1 = 3 and axis2 = 4, flip = 1 will not change either axis; flip = 3 will flip only the horizontal axis; flip = c(3, 4) will flip both axes.
include.axes	A logical argument for whether axes should be shown at x = 0 and y = 0. This is different than the axes argument in the generic plot.default function, which controls the edges of the plot (providing a box effect or not). Using include.axes = TRUE does not allow aesthetic control of the axes. If desired, it is better to use include.axes = FALSE and augment the plot object with abline (choosing h = 0 and v = 0 in separate applications).
...	other arguments passed to plot (helpful to employ different colors or symbols for different groups). See

Value

An object of class "plot.ordinate" is a list with components that can be used in other plot functions, such as the type of plot, points, a group factor, and other information depending on the plot parameters used.

Author(s)

Michael Collyer

`plot.predict.lm.rrpp` *Plot Function for RRPP*

Description

Plot Function for RRPP

Usage

```
## S3 method for class 'predict.lm.rrpp'
plot(x, PC = FALSE, ellipse = FALSE, abscissa = NULL, label = TRUE, ...)
```

Arguments

`x`	plot object (from `predict.lm.rrpp`)
`PC`	A logical argument for whether the data space should be rotated to its principal components
`ellipse`	A logical argument to change error bars to ellipses in multivariate plots. It has no function for univariate plots or is abscissa is not NULL.
`abscissa`	An optional vector (numeric of factor) equal in length to predictions to use for plotting as the abscissa (x-axis), in which case predictions are the ordinate (y-axis). This might be helpful if predictions are made for a continuous independent variable. The abscissa would be the same variable used to make predictions (and can be the data.frame used for newdata in `predict.lm.rrpp`).
`label`	A logical argument for whether points should be labeled (in multivariate plots).
`...`	other arguments passed to plot, arrows, points, or text (helpful to employ different colors or symbols for different groups). See `plot.default`, `arrows`, `points`, `par`, and `text`

Author(s)

Michael Collyer

Examples

```
# See lm.rrpp help file for examples.
```

`plot.trajectory.analysis` *Plot Function for RRPP*

Description

Function generates a principal component plot for trajectories

Usage

```
## S3 method for class 'trajectory.analysis'
plot(x, ...)
```

Arguments

`x`	plot object (from `trajectory.analysis`)
`...`	other arguments passed to plot (helpful to employ different colors or symbols for different groups). See `plot.default` and `par`

Details

The function calculates and plots principal components of fitted values from `lm.rrpp` that are passed onto `trajectory.analysis`, and projects data onto them. This function is a set.up, and `add.trajectories` is needed to add trajectories to the plot. By having two stages of control, the plotting functions are more flexible. This function also returns plotting information that can be valuable for making individualized plots, if `add.trajectories` is not preferred.

Value

If an object is assigned, it will return:

`pca`	Principal component analysis performed using `prcomp`.
`pc.points`	Principal component scores for all data.
`trajectory.analysis`	Trajectory analysis passed on.
`trajectories`	pca Observed trajectories projected onto principal components.

Author(s)

Michael Collyer

References

Adams, D. C., and M. M. Cerney. 2007. Quantifying biomechanical motion using Procrustes motion analysis. J. Biomech. 40:437-444.

Adams, D. C., and M. L. Collyer. 2007. The analysis of character divergence along environmental gradients and other covariates. Evolution 61:510-515.

Adams, D. C., and M. L. Collyer. 2009. A general framework for the analysis of phenotypic trajectories in evolutionary studies. Evolution 63:1143-1154.

Collyer, M. L., and D. C. Adams. 2007. Analysis of two-state multivariate phenotypic change in ecological studies. Ecology 88:683-692.

Collyer, M. L., and D. C. Adams. 2013. Phenotypic trajectory analysis: comparison of shape change patterns in evolution and ecology. Hystrix 24: 75-83.

Collyer, M.L., D.J. Sekora, and D.C. Adams. 2015. A method for analysis of phenotypic change for phenotypes described by high-dimensional data. Heredity. 115:357-365.

See Also

`plot.default` and `par`

Examples

```
# See trajectory.analysis help file for examples
```

`podarcis` *Podarcis Wall Lizards from the Mediterranean Basin*

Description

A data set of body size for males and females form different lizard species in different geographic locations.

Details

These data are a subset of the data collected originally by Kaliontzopoulou et al.(2015). Variables include snout-to-vent-length (SVL), sex, species, and habitat preference. These data are suitable for comparing effect sizes in the book, Permutational Biometry (Collyer & Adams, 2025).

References

Kaliontzopoulou, A., Carretero, M. A., and Adams, D. C. (2015). Ecomorphological variation in male and female wall lizards and the macroevolution of sexual dimorphism in relation to habitat use. Journal of Evolutionary Biology, 28:80–94.

Collyer, M. L., & Adams D. C. (2025). Permutational Biometry: Volume 1, Univariate Data. Iowa State University Digital Press. NOT YET PUBLISHED.

powerTrans *Power Transformation*

Description

A function that transforms a vector of statistics or data to be approximately normally distributed.

Usage

```
powerTrans(
  y,
  eps = 0.001,
  target = NULL,
  interp = FALSE,
  forceYJ = FALSE,
  useStDev = FALSE
)
```

Arguments

y	A vector of values to use.
eps	Tolerance for lambda = 0.
target	The value to target in the distribution. (If null, the first value in the vector is used.) This will be moved to the first value in results.
interp	Logical value for whether to spline interpolate lambda (ideal for vectors of small length). This will default if the number of values is less than 100, as in boxcox.
forceYJ	A logical value whether to force a Yeo-Johnson transformation for non-negative values. Negative values trigger the transformation, but positive values can be transformed similarly.
useStDev	A logical value whether to initially use standard deviates of y. This might be useful if the range of y is restricted, say between 0 and 1.

Details

This function uses a Box-Cox power transformation for a vector of values, (presumably statistics from RRPP) with the following conditions:

(1) If any values are 0 or negative, either a Yeo-Johnson transformation or a linear transformation of y - min(y) + 1e-7 is performed to optimize lambda with only positive values. There might be cases where one works better than the other. This function will determine which produces more normally distributed data, indicating which transformation was used.

(2) The lambda parameter is optimized over a range of -2 to +2, as is typical for most applications.

(3) The first (or targeted) value (assumed to be the observed value in a distribution of random values) is removed from the vector to perform optimization of lambda, but is returned to the vector for transformation. This assures that large, aberrant values, such as might be found via RRPP for large effects, do not have leverage for the optimization.

A Yeo-Johnson transformation can be forced, which for positive values merely shifts the distribution by adding 1. For most RRPP statistics (SS. MS, Rsq, F, d), this is not needed for effect sizes, as values are positive. However, for certain distributions, this might be a good idea for comparisons, because values might be strongly positive or bimodal in nature.

Using standard deviates (useStDev) is a good idea for statistics with aberrant ranges (like 0 to 1, or 1,000 to 2,000). If used, a Yeo-Johnson or linear transformation is implicit, since standardizing introduces negative values.

Value

A list containing the following:

`opt.lambda`	The optimized lambda parameter.
`transformed`	The power-transformed values.
`lambda`	If spline interpolation is used, the values of lambda used.
`loglik`	If spline interpolation is used, the log-likelihoods calculated for lambda.
`additional_trans`	How additional transformation was performed: one of a linear transformation, a Yeo-Johnson transformation, or none. The result could also indicate whether standard deviates of y were used.

Author(s)

Michael Collyer

References

Box, G. E. P. and Cox, D. R. (1964) An analysis of transformations (with discussion). Journal of the Royal Statistical Society B, 26, 211–252.

Yeo, I. and Johnson, R. (2000) A new family of power transformations to improve normality or symmetry. Biometrika, 87, 954-959.

---

predict.lm.rrpp *predict for lm.rrpp model fits*

Description

Computes predicted values from an `lm.rrpp` model fit, using bootstrapped residuals to generate confidence intervals. (Residuals are the residuals of the lm.rppp fit, not its null model. The bootstrap procedure resamples residual vectors with replacement.) The bootstrap permutations use the same number of iterations and seed as used in the `lm.rrpp` model fit. A `predict.lm.rrpp` object can be plotted using various options. See `plot.predict.lm.rrpp`.

Note that if data offsets are used (if the offset argument is used when fitting a `lm.rrpp` model), they are ignored for estimating coefficients over iterations. Offsets are subtracted from data in `lm` and added to predicted values in `predict.lm`, effectively adjusting the intercept and then un-adjusting it for predictions. This causes problems if the newdata have a different number of observations than the original model fit.

Usage

```
## S3 method for class 'lm.rrpp'
predict(object, newdata = NULL, block = NULL, confidence = 0.95, ...)
```

Arguments

object	Object from lm.rrpp.
newdata	Data frame of either class data.frame or rrpp.data.frame. If null, the data frame from the lm.rrpp fit will be used, effectively calculating all fitted values and their confidence intervals. If a numeric variable is missing from newdata, an attempt to average the values will be made in prediction; i.e., least squares means for factor levels can be found. All factors used in the lm.rrpp fit should be represented in the newdata data frame, with appropriate factor levels.
block	An optional factor for blocks within which to restrict resampling permutations.
confidence	The desired confidence interval level for prediction.
...	Other arguments (currently none)

Author(s)

Michael Collyer

Examples

```
## Not run:
# See examples for lm.rrpp to see how predict.lm.rrpp works in conjunction
# with other functions

data(Pupfish)

# CS is centroid (fish) size
fit <- lm.rrpp(coords ~ log(CS)  + Sex*Pop,
SS.type = "I", data = Pupfish, iter = 999)

# Predictions (holding alternative effects constant)

shapeDF <- expand.grid(Sex = levels(Pupfish$Sex), Pop = levels(Pupfish$Pop))
rownames(shapeDF) <- paste(shapeDF$Sex, shapeDF$Pop, sep = ".")
shapeDF

shapePreds <- predict(fit, shapeDF)
summary(shapePreds)
summary(shapePreds, PC = TRUE)

shapePreds99 <- predict(fit, shapeDF, confidence = 0.99)
summary(shapePreds99, PC = TRUE)

# Plot prediction

plot(shapePreds, PC = TRUE)
plot(shapePreds, PC = TRUE, ellipse = TRUE)
```

```
plot(shapePreds99, PC = TRUE)
plot(shapePreds99, PC = TRUE, ellipse = TRUE)

## End(Not run)
```

`prep.lda` *Linear discriminant function for lm.rrpp model fits*

Description

Function creates arguments for `lda` or `qda` from a `lm.rrpp` fit.

Usage

```
prep.lda(
  fit,
  tol = 1e-07,
  PC.no = NULL,
  newdata = NULL,
  inherent.groups = FALSE,
  ...
)
```

Arguments

`fit`	A linear model fit using `lm.rrpp`.
`tol`	A tolerance used to decide if the matrix of data is singular. This value is passed onto both `lda` and `prcomp`, internally.
`PC.no`	An optional argument to define the specific number of principal components (PC) used in analysis. The minimum of this value or the number of PCs resulting from the tol argument will be used.
`newdata`	An optional matrix (or object coercible to a matrix) for classification. If NULL, all observed data are used.
`inherent.groups`	A logical argument in case one wishes to have the inherent groups in the model fit revealed. If TRUE, no other analysis will be done than to reveal the groups. This argument should always be FALSE to perform a classification analysis.
`...`	Arguments passed to `lda`. See `lda` for details

Details

This function uses a `lm.rrpp` fit to produce the data and the groups to use in `lda` or `qda`.There are two general purposes of this function that are challenging when using `lda`, directly. First, this function finds the inherent groups in the `lm.rrpp` fit, based on factor levels. Second, this function find pseudodata - rather than the observed data - that involve either or both a principal component

projection with appropriate (or user-prescribed) dimensions and a transformation. The principal component projection incorporates GLS mean-centering, where appropriate. Transformation involves holding non-grouping model terms constant. This is accomplished by using the fitted values from the lm.rrpp fit and the residuals of a lm.rrpp fit with grouping factors, alone. When, the lm.rrpp fit contains only grouping factors, this function produces raw data projected on principal components.

Regardless of variables input, data are projected onto PCs. The purpose of this function is to predict group association, and working in PC space facilitates this objective.

This is a new function and not all limits and scenarios have been tested before its release. Please report any issues or limitations or strange results to the maintainer.

Notes for RRPP 0.5.0 and subsequent versions:

Prior to version 0.5.0, the function, classify, was available. This function has been deprecated. It mimicked lda with added features that are largely retained with prep.lda. However, prep.lda facilitates the much more diverse options available with lda.

Value

A list of arguments that can be passed to lda. As a minimum, these arguments include $x, $grouping, and $tol. If newdata is not NULL, $newdata, using the same transformation and PCs as for the data, will also be included.

Author(s)

Michael Collyer

See Also

lda, predict.lda, qda, predict.qda

Examples

```
# Using the Pupfish data (see lm.rrpp help for more detail)

data(Pupfish)
Pupfish$logSize <- log(Pupfish$CS)
fit <- lm.rrpp(coords ~ logSize + Sex * Pop, SS.type = "I",
data = Pupfish, print.progress = FALSE, iter = 0)

prep.lda(fit, inherent.groups = TRUE) # see groups available
lda.args <- prep.lda(fit, CV = TRUE, PC.no = 6)
lda.args$x
lda.args$grouping

# not run:
# library(MASS)
# LDA <- do.call(lda, lda.args)
# LDA$posterior
# table(lda.args$grouping, LDA$class)
```

print.anova.lm.rrpp *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'anova.lm.rrpp'
print(x, ...)
```

Arguments

x	print/summary object (from lm.rrpp)
...	other arguments passed to print/summary

Author(s)

Michael Collyer

print.betaTest *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'betaTest'
print(x, ...)
```

Arguments

x	Object from betaTest
...	Other arguments passed onto betaTest

Author(s)

Michael Collyer

print.coef.lm.rrpp *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'coef.lm.rrpp'
print(x, ...)
```

Arguments

x	Object from coef.lm.rrpp
...	Other arguments passed onto coef.lm.rrpp

Author(s)

Michael Collyer

print.ICCstats *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'ICCstats'
print(x, ...)
```

Arguments

x	print/summary object (from ICCstats)
...	other arguments passed to print/summary

Author(s)

Michael Collyer

print.kcomp *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'kcomp'
print(x, ...)
```

Arguments

x	Object from kcomp
...	Other arguments passed onto print.kcomp

Author(s)

Michael Collyer

print.lm.rrpp *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'lm.rrpp'
print(x, ...)
```

Arguments

x	print/summary object (from lm.rrpp)
...	other arguments passed to print/summary

Author(s)

Michael Collyer

print.looCV *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'looCV'
print(x, ...)
```

Arguments

x	Object from looCV
...	Other arguments passed onto print.looCV

Author(s)

Michael Collyer

print.lr_test *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'lr_test'
print(x, ...)
```

Arguments

x	Object from lr_test
...	Other arguments passed onto print.lr_test

Author(s)

Michael Collyer

print.measurement.error *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'measurement.error'
print(x, ...)
```

Arguments

x	Object from measurement.error
...	Other arguments passed onto measurement.error

Author(s)

Michael Collyer

Examples

```
## Not run:

# Measurement error analysis on simulated data of fish shapes

data(fishy)

# Analysis unconcerned with groups

ME1 <- measurement.error(
  Y = "coords",
  subjects = "subj",
  replicates = "reps",
  data = fishy)

anova(ME1)
ICCstats(ME1, subjects = "Subjects", with_in = "Systematic ME")
plot(ME1)

# Analysis concerned with groups

ME2 <- measurement.error(
  Y = "coords",
  subjects = "subj",
  replicates = "reps",
  groups = "groups",
```

```
  data = fishy)

anova(ME2)
ICCstats(ME2, subjects = "Subjects",
  with_in = "Systematic ME", groups = "groups")
P <- plot(ME2)
focusMEonSubjects(P, subjects = 18:20, shadow = TRUE)

## End(Not run)
```

`print.model.comparison` *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'model.comparison'
print(x, ...)
```

Arguments

`x`	Object from `model.comparison`
`...`	Other arguments passed onto model.comparison

Author(s)

Michael Collyer

`print.ordinate` *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'ordinate'
print(x, ...)
```

Arguments

x	Object from ordinate
...	Other arguments passed onto print.ordinate

Author(s)

Michael Collyer

---

print.pairwise *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'pairwise'
print(x, ...)
```

Arguments

x	Object from pairwise
...	Other arguments passed onto pairwise

Author(s)

Michael Collyer

---

print.pairwise.model.Z *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'pairwise.model.Z'
print(x, stats.table = TRUE, ...)
```

Arguments

x	print/summary object (from pairwise.model.Z)
stats.table	A logical value for whether to condense results into a single table of multiple statistics.
...	other arguments passed to print/summary

Author(s)

Dean Adams and Michael Collyer

---

print.predict.lm.rrpp *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'predict.lm.rrpp'
print(x, PC = FALSE, ...)
```

Arguments

x	Object from predict.lm.rrpp
PC	Logical argument for whether to use predicted values rotated to their PCs
...	Other arguments passed onto predict.lm.rrpp

Author(s)

Michael Collyer

---

print.summary.betaTest *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'summary.betaTest'
print(x, ...)
```

Arguments

x	Object from betaTest
...	Other arguments passed onto betaTest

Author(s)

Michael Collyer

---

print.summary.lm.rrpp *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'summary.lm.rrpp'
print(x, ...)
```

Arguments

x	print/summary object (from summary.lm.rrpp)
...	other arguments passed to print/summary

Author(s)

Michael Collyer

---

print.summary.manova.lm.rrpp *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'summary.manova.lm.rrpp'
print(x, ...)
```

Arguments

x	Object from summary.manova.lm.rrpp
...	Other arguments passed onto summary.manova.lm.rrpp

Author(s)

Michael Collyer

`print.summary.ordinate`

Print/Summary Function for RRPP

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'summary.ordinate'
print(x, ...)
```

Arguments

`x`	Object from `summary.ordinate`
`...`	Other arguments passed onto print.ordinate

Author(s)

Michael Collyer

`print.summary.pairwise`

Print/Summary Function for RRPP

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'summary.pairwise'
print(x, ...)
```

Arguments

`x`	Object from `summary.pairwise`
`...`	Other arguments passed onto summary.pairwise

Author(s)

Michael Collyer

`print.summary.trajectory.analysis`

Print/Summary Function for RRPP

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'summary.trajectory.analysis'
print(x, ...)
```

Arguments

`x`	Object from `summary.trajectory.analysis`
`...`	Other arguments passed onto summary.trajectory.analysis

Author(s)

Michael Collyer

`print.trajectory.analysis`

Print/Summary Function for RRPP

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'trajectory.analysis'
print(x, ...)
```

Arguments

`x`	Object from `trajectory.analysis`
`...`	Other arguments passed onto

Author(s)

Michael Collyer

Pupfish *Landmarks on pupfish*

Description

Landmark data from Cyprinodon pecosensis body shapes, with indication of Sex and Population from which fish were sampled (Marsh or Sinkhole).

Details

These data were previously aligned with GPA. Centroid size (CS) is also provided. See the **geomorph** package for details.

Author(s)

Michael Collyer

References

Collyer, M.L., D.J. Sekora, and D.C. Adams. 2015. A method for analysis of phenotypic change for phenotypes described by high-dimensional data. Heredity. 113: doi:10.1038/hdy.2014.75.

PupfishHeads *Landmarks on pupfish heads*

Description

Landmark data from Cyprinodon pecosensis head shapes, with variables for sex, month and year sampled, locality, head size, and coordinates of landmarks for head shape, per specimen. These data are a subset of a larger data set.

Details

The variable, "coords", are data that were previously aligned with GPA. The variable, "headSize", is the Centroid size of each vector of coordinates. See the **geomorph** package for details.

Author(s)

Michael Collyer

References

Gilbert, M.C. 2016. Impacts of habitat fragmentation on the cranial morphology of a threatened desert fish (Cyprinodon pecosensis). Masters Thesis, Western Kentucky University.

PupfishMR — *Experimental metabolic rate data from pupfish*

Description

Simulated metabolic rate data from experimental pupfish

Details

These data are simulated data meant to mimic a subset of the data from Collyer & Stockwell (2004). Variables include experimental treatment (Control versus artificially infected), mass in grams, standard length in mm, and metabolic rate (oxygen consumption, mg O2 per L per hour). These data do not have a temporal component like the original data, and do not include aberrant or missing observations. These data are suitable data for demonstrating linear regression and analysis of covariance (ANCOVA), used in the book, Permutational Biometry (Collyer & Adams, 2025).

Author(s)

Michael Collyer

References

Collyer, M. L., & Stockwell, C. A. (2004). Experimental evidence for costs of parasitism for a threatened species, White Sands pupfish (Cyprinodon tularosa). Journal of Animal Ecology, 73(5), 821-830.

Collyer, M. L., & Adams D. C. (2025). Permutational Biometry: Volume 1, Univariate Data. Iowa State University Digital Press. NOT YET PUBLISHED.

pval — *Obtain P-value from a vector of statistics*

Description

A function to find the probability of values greater or lesser than target, from a vector of statistics presumably obtained in random permutations.

Usage

```
pval(s, target = NULL, greater = TRUE)
```

Arguments

s	The sampling distribution vector to use.
target	The value to target in the distribution. (If null, the first value in the vector is used.). If the target exists outside the range of s, a probability of 0 or 1 is certain.
greater	Logical value for whether the probability should be "greater than or equal to". Change to greater = FALSE for "less than or equal to".

Author(s)

Michael Collyer

`QRforX` *QR decomposition of linear model design matrices*

Description

This function performs a QR decomposition (factorization) on a linear model design matrix (X) and returns useful results for subsequent analysis. This is intended as an internal function but can be used externally. Because base::qr and Matrix::qr have different options for QR algorithms, this function assures that results are consistent for other RRPP function use, whether X is a dense or sparse matrix.

Usage

```
QRforX(X, returnQ = TRUE, ...)
```

Arguments

`X`	A linear model design matrix, but can be any object coercible to matrix.
`returnQ`	A logical value whether to return the Q matrix. Generating a Q matrix can be computationally intense for large matrices. If it is not explicitly needed, this argument can be FALSE.
`...`	Further arguments passed to base::qr.

Value

An object of class `QR` is a list containing the following:

`Q`	The Q matrix, if requested.
`R`	The R matrix.
`X`	The X matrix, which could be changed from dense to sparse, or vice versa, and redundant columns removed.
`dimnames`	The dimnames associated with Q, R, or X
`rank`	The rank of the X matrix.
`S4`	Logical value for whether Q, R, and X are S4 class objects.

Author(s)

Michael Collyer

Examples

```
## Simple Example
data(Pupfish)
fit <- lm.rrpp(coords ~ Pop, data = Pupfish, print.progress = FALSE)
QR <- QRforX(model.matrix(fit))
QR$Q
QR$R
QR$rank
QR$S4

## Not run, but one could get base::qr and Matrix::qr results as

# base::qr(as.matrix(QR$X))
# Matrix::qr(QR$X)

## Complex example

data("PupfishHeads")
fit <- suppressWarnings(lm.rrpp(headSize ~ sex +
locality/year, data = PupfishHeads))
X <- model.matrix(fit)
dim(X) # Already reduced
colnames(X)
X <- model.matrix(terms(fit), fit$LM$data)
dim(X) # Retains redundant parameters
colnames(X)
QR <- QRforX(X)
dim(QR$X)
dim(QR$Q)
dim(QR$R)
QR$rank
```

`ranef.lmm.rrpp` *ranef for lmm.rrpp model fits*

Description

Returns random effects from a lmm.rrpp object.

Usage

```
## S3 method for class 'lmm.rrpp'
ranef(object, type = c("matrix", "list"), ...)
```

Arguments

`object`	Object from `lmm.rrpp`
`type`	Whether to combine random effects into one matrix or print a list by variable.
`...`	Other arguments passed.

Author(s)

Michael Collyer

`ReptAbund` *Reptile Abundance Data in Western Australia*

Description

Reptile abundance data from several sites in Western Australia, originally collected to examine recovery after a large fire.

Details

The data set is from a study examining reptile abundance from several sites in Western Australia (Davis and Doherty, 2015). The original study examined the recovery of reptile communities in ten remnant localities following a large fire in 2009. Half of localities were in the burn zone and the remaining half were unaffected by the fire. Data were sampled over five years. These data are suitable data for demonstrating mixed linear models, used in the book, Permutational Biometry (Collyer & Adams, 2025).

References

Davis, R. A. and Doherty, T. S. (2015). Rapid recovery of an urban remnant reptile community following summer wildfire. PLOS ONE, 10(5):e0127925.

Collyer, M. L., & Adams D. C. (2025). Permutational Biometry: Volume 1, Univariate Data. Iowa State University Digital Press. NOT YET PUBLISHED.

`residuals.lm.rrpp` *Extract residuals*

Description

Extract residuals

Usage

```
## S3 method for class 'lm.rrpp'
residuals(object, ...)
```

Arguments

`object`	plot object (from `lm.rrpp`)
`...`	Arguments passed to other functions

Author(s)

Michael Collyer

Examples

```
# See examples for lm.rrpp
```

---

reveal.model.designs *Reveal model designs used in lm.rrpp fit*

Description

Function returns every full and reduced model for model terms used in lm.rrpp fits. This function is useful for revealing the null and full model that would be used in the pairwise function, if a specific null model is not declared as an argument (fit.null in the pairwise function). It also helps to demonstrate how sums of squares and cross-products (SSCP) are calculated in lm.rrpp permutations (iterations), from the difference between fitted values for null and full designs.

Usage

```
reveal.model.designs(fit)
```

Arguments

fit	A linear model fit from lm.rrpp.

Author(s)

Michael Collyer

Examples

```
data(Pupfish)
fit1 <- lm.rrpp(coords~ Pop*Sex, data = Pupfish,
SS.type = "I", print.progress = FALSE, iter = 0)
fit2 <- lm.rrpp(coords~ Pop*Sex, data = Pupfish,
SS.type = "II", print.progress = FALSE, iter = 0)
fit3 <- lm.rrpp(coords~ Pop*Sex, data = Pupfish,
SS.type = "III", print.progress = FALSE, iter = 0)

reveal.model.designs(fit1)
reveal.model.designs(fit2)
reveal.model.designs(fit3)
```

`rrpp.data.frame` *Create a data frame for lm.rrpp analysis*

Description

Create a data frame for lm.rrpp analysis, when covariance or distance matrices are used

Usage

```
rrpp.data.frame(...)
```

Arguments

`...`	Components (objects) to combine in the data frame.

Details

This function is not much different than `data.frame` but is more flexible to allow distance matrices and covariance matrices to be included. Essentially, this function creates a list, much like an object of class `data.frame` is also a list. However, `rrpp.data.frame` is less concerned with coercing the list into a matrix and more concerned with matching the number of observations (n). It is wise to use this function with any `lm.rrpp` analysis so that `lm.rrpp` does not have to search the global environment for needed data.

It is assumed that multiple data sets for the same subjects are in the same order.

See `lm.rrpp` for examples.

Author(s)

Michael Collyer

Examples

```
# Why use a rrpp.data.frame?
y <- matrix(rnorm(30), 10, 3)
x <- rnorm(10)
df <- data.frame(x = x, y = y)
df
rdf <- rrpp.data.frame(x = x, y = y)
rdf # looks more like a list

is.list(df)
is.list(rdf)

d <- dist(y) # distance matrix as data

# One can try this but it will result in an error
# df <- data.frame(df, d = d)
rdf <- rrpp.data.frame(rdf, d = d) # works
```

```
fit <- lm.rrpp(d ~ x, data = rdf, iter = 99)
summary(fit)
```

---

scaleCov *Scaling of a Covariance Matrix*

Description

Function performs linear and exponential scaling of a covariance, either including or excluding diagonals or off-diagonal elements.

Usage

```
scaleCov(
  Cov,
  scale. = 1,
  exponent = 1,
  scale.diagonal = FALSE,
  scale.only.diagonal = FALSE
)
```

Arguments

Cov	Square matrix to be scaled.
scale.	The linear scaling parameter. Values are multiplied by this numeric value.
exponent	The exponentiation scaling parameter. Values are raised to this power.
scale.diagonal	Logical to indicate if diagonal should be included.
scale.only.diagonal	Logical to indicate if only the diagonal should be scaled.

Details

The function scales covariances as scale * cov ^exponent, where cov is any covariance or variance in the covariance matrix. Arguments allow inclusion or exclusion or either the diagonal or off-diagonal elements to be scaled. It is assumed that a covariance matrix is scaled, but any square matrix will work.

Value

A square matrix.

Author(s)

Michael Collyer

Examples

```
## Not run:
data(Pupfish)
Pupfish$logSize <- log(Pupfish$CS)
fit1 <- lm.rrpp(coords ~ logSize, data = Pupfish, iter = 0,
print.progress = FALSE)
fit2 <- lm.rrpp(coords ~ Pop, data = Pupfish, iter = 0,
print.progress = FALSE)
fit3 <- lm.rrpp(coords ~ Sex, data = Pupfish, iter = 0,
print.progress = FALSE)
fit4 <- lm.rrpp(coords ~ logSize + Sex, data = Pupfish, iter = 0,
print.progress = FALSE)
fit5 <- lm.rrpp(coords ~ logSize + Pop, data = Pupfish, iter = 0,
print.progress = FALSE)
fit6 <- lm.rrpp(coords ~ logSize + Sex * Pop, data = Pupfish, iter = 0,
print.progress = FALSE)

modComp1 <- model.comparison(fit1, fit2, fit3, fit4, fit5,
fit6, type = "cov.trace")
modComp2 <- model.comparison(fit1, fit2, fit3, fit4, fit5,
fit6, type = "logLik", tol = 0.01)

summary(modComp1)
summary(modComp2)

par(mfcol = c(1,2))
plot(modComp1)
plot(modComp2)

# Comparing fits with covariance matrices
# an example for scaling a phylogenetic covariance matrix with
# the scaling parameter, lambda

data("PlethMorph")
Cov <- PlethMorph$PhyCov
lambda <- seq(0, 1, 0.1)

Cov1 <- scaleCov(Cov, scale. = lambda[1])
Cov2 <- scaleCov(Cov, scale. = lambda[2])
Cov3 <- scaleCov(Cov, scale. = lambda[3])
Cov4 <- scaleCov(Cov, scale. = lambda[4])
Cov5 <- scaleCov(Cov, scale. = lambda[5])
Cov6 <- scaleCov(Cov, scale. = lambda[6])
Cov7 <- scaleCov(Cov, scale. = lambda[7])
Cov8 <- scaleCov(Cov, scale. = lambda[8])
Cov9 <- scaleCov(Cov, scale. = lambda[9])
Cov10 <- scaleCov(Cov, scale. = lambda[10])
Cov11 <- scaleCov(Cov, scale. = lambda[11])


fit1 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov1)
fit2 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov2)
```

```
fit3 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov3)
fit4 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov4)
fit5 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov5)
fit6 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov6)
fit7 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov7)
fit8 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov8)
fit9 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov9)
fit10 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov10)
fit11 <- lm.rrpp(SVL ~ 1, data = PlethMorph, Cov = Cov11)

par(mfrow = c(1,1))

MC1 <- model.comparison(fit1, fit2, fit3, fit4, fit5, fit6,
fit7, fit8, fit9, fit10, fit11,
type = "logLik")
MC1
plot(MC1)

MC2 <- model.comparison(fit1, fit2, fit3, fit4, fit5, fit6,
fit7, fit8, fit9, fit10, fit11,
type = "logLik", predictor = lambda)
MC2
plot(MC2)


MC3 <- model.comparison(fit1, fit2, fit3, fit4, fit5, fit6,
fit7, fit8, fit9, fit10, fit11,
type = "Z", predictor = lambda)
MC3
plot(MC3)

## End(Not run)
```

---

snakeHS *Snake Head Size and Regional Variation*

Description

A data set containing variables for snake head size, snout-to-vent-length, sex, and geographic information.

Details

These data were collected from prairie rattlesnakes (Crotalus viridis viridis), from several geographic locations in the upper Midwest of the United States (Smith and Collyer, 2008). Variables include a spatial measure of head size (HS), snout-to-vent-length (SVL, cm), sex, and the site and region where 107 snakes were captured. These data are suitable for demonstrating model selection in the book, Permutational Biometry (Collyer & Adams, 2025).

References

Smith, M. T. and Collyer, M. L. (2008). The biology of rattlesnakes, chapter Regional variation and sexual dimorphism in head form of the prairie rattlesnake (Crotalus viridid viridis): Comparisons using new analytical techniques and collection methods, pages 79–90. Loma Linda University Press.

Collyer, M. L., & Adams D. C. (2025). Permutational Biometry: Volume 1, Univariate Data. Iowa State University Digital Press. NOT YET PUBLISHED.

`summary.anova.lm.rrpp` *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'anova.lm.rrpp'
summary(object, ...)
```

Arguments

`object`	Object from `predict.lm.rrpp`
`...`	Other arguments passed onto predict.lm.rrpp

Author(s)

Michael Collyer

`summary.betaTest` *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'betaTest'
summary(object, confidence = 0.95, ...)
```

Arguments

object	Object from betaTest
confidence	The desired confidence limit to print with a table of summary statistics. Because distances are directionless, confidence limits are one-tailed.
...	Other arguments passed onto betaTest

Author(s)

Michael Collyer

---

summary.coef.lm.rrpp *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'coef.lm.rrpp'
summary(object, ...)
```

Arguments

object	Object from coef.lm.rrpp
...	Other arguments passed onto coef.lm.rrpp

Author(s)

Michael Collyer

---

summary.ICCstats *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'ICCstats'
summary(object, ...)
```

Arguments

object	print/summary object (from ICCstats)
...	other arguments passed to print/summary

Author(s)

Michael Collyer

---

summary.kcomp *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'kcomp'
summary(object, ...)
```

Arguments

object	Object from kcomp
...	Other arguments passed onto print.kcomp

Author(s)

Michael Collyer

---

summary.lm.rrpp *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'lm.rrpp'
summary(object, formula = TRUE, ...)
```

Arguments

object	print/summary object (from lm.rrpp)
formula	Logical argument for whether to include formula in summary table
...	other arguments passed to print/summary

Author(s)

Michael Collyer

`summary.looCV` *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'looCV'
summary(object, ...)
```

Arguments

`object`	Object from `looCV`
`...`	Other arguments passed onto print.looCV

Author(s)

Michael Collyer

`summary.lr_test` *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'lr_test'
summary(object, ...)
```

Arguments

`object`	Object from `lr_test`
`...`	Other arguments passed onto print.lr_test

Author(s)

Michael Collyer

summary.manova.lm.rrpp

Print/Summary Function for RRPP

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'manova.lm.rrpp'
summary(object, test = c("Roy", "Pillai", "Hotelling-Lawley", "Wilks"), ...)
```

Arguments

object	Object from lm.rrpp, updated with manova.update
test	Type of multivariate test statistic to use.
...	Other arguments passed onto manova.lm.rrpp

Author(s)

Michael Collyer

summary.measurement.error

Print/Summary Function for RRPP

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'measurement.error'
summary(object, ...)
```

Arguments

object	Object from measurement.error
...	Other arguments passed onto measurement.error

Author(s)

Michael Collyer

summary.model.comparison *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'model.comparison'
summary(object, ...)
```

Arguments

object	Object from model.comparison
...	Other arguments passed onto model.comparison

Author(s)

Michael Collyer

summary.ordinate *Print/Summary Function for RRPP*

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'ordinate'
summary(object, ...)
```

Arguments

object	Object from ordinate
...	Other arguments passed onto print.ordinate

Author(s)

Michael Collyer

summary.pairwise *Print/Summary Function for RRPP*

Description

See pairwise for further description.

Usage

```
## S3 method for class 'pairwise'
summary(
  object,
  stat.table = TRUE,
  test.type = c("dist", "stdist", "mdist", "VC", "DL", "var"),
  angle.type = c("rad", "deg"),
  confidence = 0.95,
  show.vectors = FALSE,
  p.adjust.method = "none",
  ...
)
```

Arguments

object	Object from pairwise
stat.table	Logical argument for whether results should be returned in one table (if TRUE) or separate pairwise tables (if FALSE)
test.type	the type of statistic to test. See below should be used in the test.
angle.type	If test.type = "VC", whether angle results are expressed in radians or degrees.
confidence	Confidence level to use for upper confidence limit; default = 0.95 (alpha = 0.05)
show.vectors	Logical value to indicate whether vectors should be printed.
p.adjust.method	Optional logical value to adjust P-values, based on p.adjust. Defaults to "none".
...	Other arguments passed onto pairwise

Details

The following summarize the test that can be performed:

Distance between vectors, "dist" Vectors for LS means or slopes originate at the origin and point to some location, having both a magnitude and direction. A distance between two vectors is the inner-product of of the vector difference, i.e., the distance between their endpoints. For LS means, this distance is the difference between means. For multivariate slope vectors, this is the difference in location between estimated change for the dependent variables, per one-unit change of the covariate considered. For univariate slopes, this is the absolute difference between slopes.

Standardized distance between vectors, "stdist" Same as the distance between vectors, but distances are divided by the model standard error (square-root of the trace of the residual covariance matrix). Pairwise tests with this statistic should be consistent with ANOVA results.

Mahalanobis distance between vectors, "mdist" Similar to the standardized distance between vectors but the inverse of the residual covariance matrix is used in calculation of the distance, rather than dividing the Euclidean distance between means and dividing by the standard error. If the residual covariance matrix is singular, Mahalanobis distances will not be calculated. Pairwise tests with this statistic should be consistent with MANOVA results.

Vector correlation, "VC" If LS mean or slope vectors are scaled to unit size, the vector correlation is the inner-product of the scaled vectors. The arccosine (acos) of this value is the angle between vectors, which can be expressed in radians or degrees. Vector correlation indicates the similarity of vector orientation, independent of vector length.

Difference between vector lengths, "DL" If the length of a vector is an important attribute – e.g., the amount of multivariate change per one-unit change in a covariate – then the absolute value of the difference in vector lengths is a practical statistic to compare vector lengths, rather than the estimates the vectors make. Let d1 and d2 be the distances (length) of vectors. Then |d1 - d2| is a statistic that compares their lengths. For slope vectors, this is a comparison of rates. For comparison, if vectors are rates, "dist" finds the difference between estimates per unit change of, e.g., time, size, etc., which could be large, even for small rates of change, if vectors point in dissimilar directions. "DL" is a comparison of rates, irrespective of direction.

Variance, "var" Vectors of residuals from a linear model indicate can express the distances of observed values from fitted values. Mean squared distances of values (variance), by group, can be used to measure the amount of dispersion around estimated values for groups. Absolute differences between variances are used as test statistics to compare mean dispersion of values among groups. Variance degrees of freedom equal n, the group size, rather than n-1, as the purpose is to compare mean dispersion in the sample. (Additionally, tests with one subject in a group are possible, or at least not a hindrance to the analysis.)

The argument, `test.type` is used to select one of the tests above. See `pairwise` for examples.

Notes for RRPP 0.6.2 and subsequent versions:

In previous versions of pairwise, `summary.pairwise` had three test types: "dist", "VC", and "var". When one chose "dist", for LS mean vectors, the statistic was the inner-product of the vector difference. For slope vectors, "dist" returned the absolute value of the difference between vector lengths, which is "DL" in 0.6.2 and subsequent versions. This update uses the same calculation, irrespective of vector types. Generally, "DL" is the same as a contrast in rates for slope vectors, but might not have much meaning for LS means. Likewise, "dist" is the distance between vector endpoints, which might make more sense for LS means than slope vectors. Nevertheless, the user has more control over these decisions with version 0.6.2 and subsequent versions.

Notes for RRPP 2.0.4 and subsequent versions:

The test types, standardized distance between vectors, "stdist", and Mahalanobis distances between vectors were added. The former simply divides the distance between vectors by model standard error (square-root of the trace of the residual covariance matrix). This is a multivariate generalization of a t-statistic for the difference between means. It is not the same as Hotelling squared-T-statistic, which requires incorporating the inverse of the residual covariance matrix in the calculation of the distance, a calculation that also requires a non-singular covariance matrix.

However, the Mahalanobis distances are similar (and proportional) to the Hotelling squared-T-statistic. Pairwise tests using Mahalanobis distances represent a non-parametric analog to the parametric Hotelling squared-T test. Both tests should be better for GLS model fits compared to Euclidean distances between means, as the total sums of squares are more likely to vary across random permutations. In general, if ANOVA is performed a pairwise test with "stdist" is a good association; if MANOVA is performed, a pairwise test with "mdist" is a good association.

Author(s)

Michael Collyer

`summary.pairwise.model.Z`

Print/Summary Function for RRPP

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'pairwise.model.Z'
summary(object, ...)
```

Arguments

`object`	print/summary object (from `pairwise.model.Z`)
`...`	other arguments passed to print/summary

Author(s)

Dean Adams and Michael Collyer

`summary.predict.lm.rrpp`

Print/Summary Function for RRPP

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'predict.lm.rrpp'
summary(object, ...)
```

Arguments

object	Object from predict.lm.rrpp
...	Other arguments passed onto predict.lm.rrpp

Author(s)

Michael Collyer

---

summary.trajectory.analysis

Print/Summary Function for RRPP

Description

Print/Summary Function for RRPP

Usage

```
## S3 method for class 'trajectory.analysis'
summary(
  object,
  stat.table = TRUE,
  attribute = c("MD", "TC", "SD"),
  angle.type = c("rad", "deg"),
  confidence = 0.95,
  p.adjust.method = "none",
  show.trajectories = FALSE,
  ...
)
```

Arguments

object	Object from trajectory.analysis
stat.table	Logical argument for whether results should be returned in one table (if TRUE) or separate pairwise tables (if FALSE)
attribute	Whether magnitude differences (MD, absolute difference in trajectory path lengths), trajectory correlations (TC), or trajectory shape differences (SD) are summarized.
angle.type	If attribute = "TC", whether angle results are expressed in radians or degrees.
confidence	Confidence level to use for upper confidence limit; default = 0.95 (alpha = 0.05)
p.adjust.method	Optional logical value to adjust P-values, based on p.adjust. Defaults to "none".
show.trajectories	Logical value to indicate whether trajectories should be printed.
...	Other arguments passed onto trajectory.analysis

Author(s)

Michael Collyer

`terms.lm.rrpp` *Extract the terms from an lm.rrpp object*

Description

`terms.lm.rrpp` returns the terms constructed for an `lm.rrpp` object.

Usage

```
## S3 method for class 'lm.rrpp'
terms(x, ...)
```

Arguments

`x`	Object from `lm.rrpp`
`...`	further arguments passed to or from other methods

Author(s)

Michael Collyer

`trajectory.analysis` *Quantify and compare shape change trajectories*

Description

Function estimates attributes of multivariate trajectories

Usage

```
trajectory.analysis(
  fit,
  fit.null = NULL,
  groups,
  traj.pts,
  pca = TRUE,
  print.progress = FALSE
)
```

Arguments

fit	A linear model fit using lm.rrpp.
fit.null	An alternative linear model fit to use as a null model for RRPP, if the null model of the fit is not desired. Note, if RRPP = FALSE (FRPP rather than RRPP), then the null model has only an intercept. If the null model is uncertain, using reveal.model.designs will help elucidate the inherent null model used.
groups	A factor or vector coercible to factor that defines trajectories.
traj.pts	Either a single value or a vector coercible to factor to define trajectory points. If only a single value, it is assumed that the data are already in the form, y1p1, y2p1, y3p1, ..., y2p2, y2p2, y3p2, ..., yjp1, yjp2, yjp3, ..., yjpk, for j variables comprising k trajectory points; i.e., traj.pts = k. If a factor, then a group * traj.pt factorial model is assumed, where traj.pts defines the levels for points within groups.
pca	A logical value to optionally project group:point means onto principal components (perform PCA on a covariance matrix of the means) This option only applies to factorial designs (traj.pts is a factor).
print.progress	A logical value to indicate whether a progress bar should be printed to the screen. This is helpful for long-running analyses.

Details

The function quantifies multivariate trajectories from a set of observations, and assesses variation in attributes of the trajectories via RRPP. A trajectory is defined by a sequence of points in the data space. These trajectories can be quantified for various attributes (their size, orientation, and shape), and comparisons of these attribute enable the statistical comparison of shape change trajectories (Collyer and Adams 2007; Adams and Collyer 2007; Adams and Collyer 2009; Turner et al. 2010; Collyer and Adams 2013).

This function is a modified version of pairwise, retaining the least squares (LS) means as trajectory points. Analysis starts with a lm.rrpp fit (but a procD.lm fit from geomorph can also be used). LS means are calculated using a grouping variable. Data can be trajectories, as a start(sensu Adams and Cerney 2007), or trajectories can be calculated from data using a factorial model (in which case trajectory points are defined by factor levels).

This function produces statistics that can be summarized with the summary.trajectory.analysis function. The summaries are consistent with those in the summary.pairwise function, pertaining to trajectory attributes including, magnitude difference (MD), the difference in path lengths of trajectories; trajectory correlations (TC), better thought of as angular differences between trajectory principal axes; and if trajectories have three or more points, shape difference (SD), the square root of summed squared point differences, after scaling, centering, and rotating trajectories. The SD is the "Procrustes" distance between trajectories (Adams and Collyer 2009), much the same way as the shape difference between anatomical landmark configurations in geometric morphometrics. If attribute = "TC" is chosen for the summary, then the angle type ("rad" or "deg", can be chosen for either radians and degrees, respectively, to return angles between principal axes.)

Plotting can be performed with plot.trajectory.analysis and add.trajectories. The former plots all principal component scores for the data, and allows point-by-point control of plot parameters. The later adds trajectories points and lines, with constrained control. By saving the

plot.trajectory.analysis object, plotting information can be retained and advanced plotting can be performed. See examples below.

Value

An object of class "trajectory.analysis" returns a list of the following:

LS.means	LS.means from pairwise function.
trajectories	Trajectories from every permutation.
PD	Path distances of trajectories from every permutation.
MD	Magnitude differences between trajectories from every permutation.
TC	Trajectory correlations from every permutation.
SD	Trajectory shape differences from every permutation.

Author(s)

Dean Adams and Michael Collyer

References

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Collyer, M. L., and D. C. Adams. 2007. Analysis of two-state multivariate phenotypic change in ecological studies. Ecology 88:683-692.

Collyer, M. L., and D. C. Adams. 2013. Phenotypic trajectory analysis: comparison of shape change patterns in evolution and ecology. Hystrix 24: 75-83.

Collyer, M.L., D.J. Sekora, and D.C. Adams. 2015. A method for analysis of phenotypic change for phenotypes described by high-dimensional data. Heredity. 115:357-365.

Examples

```
## Not run:
### Analysis of sexual dimorphism vectors (factorial approach)
data(Pupfish)
fit <- lm.rrpp(coords ~ Pop * Sex, data = Pupfish, iter = 199)
reveal.model.designs(fit)
TA <- trajectory.analysis(fit, groups = Pupfish$Pop,
traj.pts = Pupfish$Sex, print.progress = FALSE)

# Magnitude difference (absolute difference between path distances)
summary(TA, attribute = "MD")

# Correlations (angles) between trajectories
summary(TA, attribute = "TC", angle.type = "deg")
```

```
# No shape differences between vectors
summary(TA, attribute = "SD")

# Retain results
TA.summary <- summary(TA, attribute = "MD")
TA.summary$summary.table

# Plot results
TP <- plot(TA, pch = as.numeric(Pupfish$Pop) + 20, bg = as.numeric(Pupfish$Sex),
cex = 0.7, col = "gray")
add.trajectories(TP, traj.pch = c(21, 22), start.bg = 1, end.bg = 2)
legend("topright", levels(Pupfish$Pop), pch =  c(21, 22), pt.bg = 1)

### Analysis when data are already trajectories (motion paths)

# data are planar Cartesian coordinates (x, y) across 5 points (10 variables)
data(motionpaths)
fit <- lm.rrpp(trajectories ~ groups, data = motionpaths, iter = 199)
TA <- trajectory.analysis(fit, groups = motionpaths$groups, traj.pts = 5)

# Magnitude difference (absolute difference between path distances)
summary(TA, attribute = "MD")

# Correlations (angles) between trajectories
summary(TA, attribute = "TC", angle.type = "deg")

# Shape differences between trajectories
summary(TA, attribute = "SD")

TP <- plot(TA, pch = 21, bg = as.numeric(motionpaths$groups),
cex = 0.7, col = "gray")
add.trajectories(TP, traj.pch = 21, traj.bg = 1:4)

## End(Not run)
```

`vec.cor.matrix` *Support function for RRPP*

Description

Calculate vector correlations for a matrix (by rows). Used for pairwise comparisons.

Usage

```
vec.cor.matrix(M)
```

Arguments

`M`	Matrix for vector correlations.

Author(s)

Michael Collyer

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