

Package: BioVizSeq (via r-universe)

November 21, 2024

Type Package

Title Visualizing the Elements Within Bio-Sequences

Version 0.1.0

Maintainer Shiqi Zhao <zhaosq89@163.com>

Description Visualizing the types and distribution of elements within bio-sequences. At the same time, We have developed a geom layer, geom_rrect(), that can generate rounded rectangles. No external references are used in the development of this package.

License Artistic-2.0

Encoding UTF-8

Language en-US

RoxygenNote 7.2.3

Imports dplyr, ggplot2, httr, RColorBrewer, seqinr, stringr, tidyr, magrittr

NeedsCompilation no

Author Shiqi Zhao [aut, cre, cph]
(<<https://orcid.org/0000-0002-2360-956X>>)

Repository CRAN

Date/Publication 2024-11-20 10:40:14 UTC

Config/pak/sysreqs libicu-dev libssl-dev zlib1g-dev

Contents

cdd_plot	2
cdd_to_loc	3
fastaleng	4
geom_rrect	4
get_motif_location	6
gff_plot	7
gff_to_loc	8

meme_plot	8
motif_plot	10
motif_seq	11
pfam_plot	12
pfam_to_loc	13
plantcare_classify	13
plantcare_plot	14
plantcare_statistic1	15
plantcare_statistic2	15
plantcare_to_loc	16
smart_plot	17
smart_to_loc	18
upload_fa_to_plantcare	18

Index	20
--------------	-----------

cdd_plot	<i>cdd_plot</i>
----------	-----------------

Description

Visualization of domain in CDD file

Usage

```
cdd_plot(
  cdd_file,
  fasta_file,
  the_order = NULL,
  domain_select = NULL,
  shape = "RoundRect",
  r = 0.3,
  legend_size = 15,
  domain_color = NULL
)
```

Arguments

cdd_file	The path of cdd file.
fasta_file	The path of fasta file.
the_order	The path of order file. A List of Gene ID , One ID Per Line.
domain_select	The domain ID which you want to align with.
shape	RoundRect or Rect.
r	The radius of rounded corners.
legend_size	The size of legend.
domain_color	The color set of domain.

Value

p

Author(s)

Shiqi Zhao

Examples

```
hitdata_path <- system.file("extdata", "hitdata.txt", package = "BioVizSeq")
fa_path <- system.file("extdata", "idpep.fa", package = "BioVizSeq")
cdd_plot(hitdata_path, fa_path)
```

```
order_path <- system.file("extdata", "order.csv", package = "BioVizSeq")
cdd_plot(hitdata_path, fa_path, the_order = order_path)
```

cdd_to_loc	<i>cdd_to_loc</i>
------------	-------------------

Description

Extract the location information of domain from cdd file

Usage

```
cdd_to_loc(cdd_file)
```

Arguments

cdd_file CDD file.

Value

data.frame

Author(s)

Shiqi Zhao

Examples

```
hitdata_path <- system.file("extdata", "hitdata.txt", package = "BioVizSeq")
cdd_file <- readLines(hitdata_path)
domain_loc <- cdd_to_loc(cdd_file)
```

fastaleng	<i>fastaleng</i>
-----------	------------------

Description

Statistical sequence length

Usage

```
fastaleng(fasta_file)
```

Arguments

fasta_file The path of protein fasta file.

Value

data.frame

Author(s)

Shiqi Zhao

Examples

```
fasta_path <- system.file("extdata", "idpep.fa", package = "BioVizSeq")
fastaleng(fasta_path)
```

geom_rrect	<i>geom_rrect</i>
------------	-------------------

Description

Rounded rectangle

Usage

```
geom_rrect(  
  mapping = NULL,  
  data = NULL,  
  stat = "identity",  
  position = "identity",  
  r = 0.2,  
  ...,  
  na.rm = FALSE,  
  show.legend = NA,  
  inherit.aes = TRUE  
)
```

Arguments

mapping	Set of aesthetic mappings created by aes . If specified and <code>inherit.aes = TRUE</code> (the default), it is combined with the default mapping at the top level of the plot. You must supply mapping if there is no plot mapping.
data	A <code>data.frame</code> , or other object, will override the plot data. All objects will be fortified to produce a data frame.
stat	Name of stat to modify data.
position	The position adjustment to use for overlapping points on this layer.
r	The radius of rounded corners.
...	additional parameter, e.g. <code>color</code> , <code>linewidth</code> , <code>alpha</code> .
na.rm	If "FALSE" (default), missing values are removed with a warning. If "TRUE", missing values are silently removed, logical.
show.legend	Whether to show legend, logical.
inherit.aes	Whether to inherit aesthetic mappings, logical, defaults to "TRUE".

Details

draws rounded rectangle by using the locations of the four corners (`xmin`, `xmax`, `ymin` and `ymax`) like `geom_rect()`.

Value

ggplot object

Aesthetics

`geom_rrect()` understands the following aesthetics (required aesthetics are in bold):

- `xmin`
- `xmax`
- `ymin`
- `ymax`
- **`alpha`**
- **`colour`**
- **`fill`**
- **`group`**
- **`linetype`**

Learn more about setting these aesthetics in `vignette("ggplot2-specs")`.

Author(s)

Shiqi Zhao

Examples

```
library(ggplot2)
df <- data.frame(
  xmin = c(1, 2, 3),
  xmax = c(2, 3, 4),
  ymin = c(1, 2, 3),
  ymax = c(2, 3, 4),
  category = c("A", "B", "C")
)

p <- ggplot(df) +
  geom_rrect(aes(xmin = xmin, xmax = xmax,
                ymin = ymin, ymax = ymax, fill = category),
            r = 0.4, linewidth = 1, colour = "black")

print(p)
```

get_motif_location *get_motif_location*

Description

Extract the location information of motif from mast or meme file

Usage

```
get_motif_location(motif_file)
```

Arguments

motif_file The motif data of mast or meme file.

Value

list

Author(s)

Shiqi Zhao

Examples

```
meme_path <- system.file("extdata", "meme.xml", package = "BioVizSeq")
meme_file <- readLines(meme_path)
motif_loc <- get_motif_location(meme_file)

mast_path <- system.file("extdata", "mast.xml", package = "BioVizSeq")
mast_file <- readLines(mast_path)
```

```
motif_loc <- get_motif_location(mast_file)
```

`gff_plot`

gff_plot

Description

Visualization of element in gff or gtf file

Usage

```
gff_plot(  
  gff_file,  
  the_order = NULL,  
  shape = "Rect",  
  r = 0.3,  
  legend_size = 15,  
  element_color = NULL  
)
```

Arguments

<code>gff_file</code>	The path of gff file.
<code>the_order</code>	The path of order of mRNA. It is also the mRNA you want to showcase. A List of Gene ID , One ID Per Line.
<code>shape</code>	RoundRect or Rect.
<code>r</code>	The radius of rounded corners.
<code>legend_size</code>	The size of legend.
<code>element_color</code>	The color set of element.

Value

p

Author(s)

Shiqi Zhao

Examples

```
gff_path <- system.file("extdata", "test.gff", package = "BioVizSeq")  
gff_plot(gff_path)
```

gff_to_loc	<i>gff_to_loc</i>
------------	-------------------

Description

Extract the location information of element from gff or gtf file

Usage

```
gff_to_loc(gff_data, mRNA_ID = NULL)
```

Arguments

gff_data	gff file.
mRNA_ID	The mRNA you selected. If NULL, it means selecting all mRNAs.

Value

list

Author(s)

Shiqi Zhao

Examples

```
gff_path <- system.file("extdata", "test.gff", package = "BioVizSeq")
gff_data <- read.table(gff_path, header = FALSE, sep = '\t')
gff_loc <- gff_to_loc(gff_data)

ID_path <- system.file("extdata", "ID_select.csv", package = "BioVizSeq")
mRNA_ID <- readLines(ID_path)
gff_loc <- gff_to_loc(gff_data, mRNA_ID=mRNA_ID)
```

meme_plot	<i>meme_plot</i>
-----------	------------------

Description

Visualization of motif in meme file or mast file

Usage

```
meme_plot(  
  meme_file,  
  the_order = NULL,  
  motif_select = NULL,  
  shape = "RoundRect",  
  show_motif_id = FALSE,  
  r = 0.3,  
  legend_size = 15,  
  motif_color = NULL  
)
```

Arguments

meme_file	The path of meme file or mast file.
the_order	The path of order file. A List of Gene ID , One ID Per Line.
motif_select	The motif ID which you want to align with.
shape	RoundRect or Rect.
show_motif_id	Display the name of the motif.
r	The radius of rounded corners.
legend_size	The size of legend.
motif_color	The color set of motif.

Value

p

Author(s)

Shiqi Zhao

Examples

```
meme_path <- system.file("extdata", "meme.xml", package = "BioVizSeq")  
meme_plot(meme_path)  
  
mast_path <- system.file("extdata", "mast.xml", package = "BioVizSeq")  
meme_plot(mast_path)  
  
meme_plot(meme_path, motif_select="1", show_motif_id = TRUE)  
  
order_path <- system.file("extdata", "order.csv", package = "BioVizSeq")  
meme_plot(meme_path, the_order=order_path, motif_select="1")
```

`motif_plot`*motif_plot*

Description

Draws multiple rounded rectangle.

Usage

```
motif_plot(  
  motif_loc,  
  gene_length,  
  the_order = NULL,  
  motif_select = NULL,  
  shape = "RoundRect",  
  show_motif_id = FALSE,  
  r = 0.3,  
  legend_size = 15,  
  motif_color = NULL  
)
```

Arguments

<code>motif_loc</code>	A data.frame contains for columns: ID, motif, start, end.
<code>gene_length</code>	A data.frame of the length of biosequences. Two columns: ID, length.
<code>the_order</code>	A List of Gene ID , One ID Per Line.
<code>motif_select</code>	The motif ID which you want to align with.
<code>shape</code>	RoundRect or Rect.
<code>show_motif_id</code>	Display the name of the motif.
<code>r</code>	The radius of rounded corners.
<code>legend_size</code>	The size of legend.
<code>motif_color</code>	The color set of motif.

Details

`motif_plot()` draws multiple rounded rectangle to represent the above elements of biosequences, but not limited to biosequences

Value

P

Author(s)

Shiqi Zhao

Examples

```
df <- data.frame(
  ID = rep(c("geneA", "geneB", "geneC"), each = 3),
  motif = rep(c("1", "2", "3"), times = 3),
  start = c(1, 3, 6, 1, 6, 10, 10, 7, 17),
  end = c(3, 5, 11, 3, 8, 15, 12, 9, 22)
)

length_data <- data.frame(
  ID = c("geneA", "geneB", "geneC"),
  length = c(15, 27, 30)
)

order_data <- c("geneB", "geneA", "geneC")

motif_plot(df, length_data)
motif_plot(df, length_data, the_order = order_data)
```

motif_seq

motif_seq

Description

Get motif sequence from meme file or mast file

Usage

```
motif_seq(meme_file)
```

Arguments

meme_file The path of meme file or mast file.

Value

data.frame

Author(s)

Shiqi Zhao

Examples

```
meme_path <- system.file("extdata", "meme.xml", package = "BioVizSeq")
meme_file <- readLines(meme_path)
motifseq<- motif_seq(meme_file)

mast_path <- system.file("extdata", "mast.xml", package = "BioVizSeq")
```

```
mast_file <- readLines(mast_path)
motifseq<- motif_seq(mast_file)
```

pfam_plot

pfam_plot

Description

Visualization of domain in pfam result file

Usage

```
pfam_plot(
  pfam_file,
  the_order = NULL,
  domain_select = NULL,
  shape = "RoundRect",
  r = 0.3,
  legend_size = 15,
  domain_color = NULL
)
```

Arguments

pfam_file	The path of meme file or mast file.
the_order	The path of order file. A List of Gene ID , One ID Per Line.
domain_select	The domain ID which you want to align with.
shape	RoundRect or Rect.
r	The radius of rounded corners.
legend_size	The size of legend.
domain_color	The color set of domain.

Value

p

Author(s)

Shiqi Zhao

Examples

```
pfam_path <- system.file("extdata", "iprscan.tsv", package = "BioVizSeq")
order_path <- system.file("extdata", "order.csv", package = "BioVizSeq")
pfam_plot(pfam_path)
pfam_plot(pfam_path, the_order=order_path)
```

pfam_to_loc	<i>pfam_to_loc</i>
-------------	--------------------

Description

Extract the location information of domain from pfam result

Usage

```
pfam_to_loc(pfam_data)
```

Arguments

pfam_data The result file (.tsv) of pfam (via InterPro).

Value

list

Author(s)

Shiqi Zhao

Examples

```
pfam_path <- system.file("extdata", "iprscan.tsv", package = "BioVizSeq")
pfam_file <- read.table(pfam_path, sep='\t', header = FALSE)
motif_loc <- pfam_to_loc(pfam_file)
```

plantcare_classify	<i>plantcare_classify</i>
--------------------	---------------------------

Description

Classify the functions of cis element from Plantcare

Usage

```
plantcare_classify(plantcare_file)
```

Arguments

plantcare_file The result file (.tab) of Plantcare.

Value

data.frame

Author(s)

Shiqi Zhao

Examples

```

plantcare_path <- system.file("extdata", "plantCARE_output.tab", package = "BioVizSeq")
plantcare_file <- read.table(plantcare_path, header = FALSE, sep = '\t', quote="")
plantcare_data <- plantcare_classify(plantcare_file)

```

plantcare_plot	<i>plantcare_plot</i>
----------------	-----------------------

Description

Visualization of cis-element in plantcare result file

Usage

```

plantcare_plot(
  plantcare_file,
  promoter_length = 2000,
  the_order = NULL,
  shape = "Rect",
  r = 6,
  legend_size = 15,
  element_color = NULL
)

```

Arguments

plantcare_file	The path of plantcare result file (.tab).
promoter_length	The promoter length.
the_order	The path of order file. A List of Gene ID , One ID Per Line.
shape	RoundRect or Rect.
r	The radius of rounded corners.
legend_size	The size of legend.
element_color	The color set of cis-element.

Value

P

Author(s)

Shiqi Zhao

Examples

```
plantcare_path <- system.file("extdata", "plantCARE_output.tab", package = "BioVizSeq")  
plantcare_plot(plantcare_path, promoter_length = 2000)
```

plantcare_statistic1 *plantcare_statistic1*

Description

Count the number of cis element from Plantcare for heatmap

Usage

```
plantcare_statistic1(plantcare_data)
```

Arguments

plantcare_data The result of plantcare_classify().

Value

data.frame

Author(s)

Shiqi Zhao

Examples

```
plantcare_path <- system.file("extdata", "plantCARE_output.tab", package = "BioVizSeq")  
plantcare_file <- read.table(plantcare_path, header = FALSE, sep = '\t', quote="")  
plantcare_data <- plantcare_classify(plantcare_file)  
statistic_data1 <- plantcare_statistic1(plantcare_data)
```

plantcare_statistic2 *plantcare_statistic2*

Description

Count the number of cis element from Plantcare for Bar chart

Usage

```
plantcare_statistic2(plantcare_data)
```

Arguments

plantcare_data The result of plantcare_classify().

Value

data.frame

Author(s)

Shiqi Zhao

Examples

```
plantcare_path <- system.file("extdata", "plantCARE_output.tab", package = "BioVizSeq")
plantcare_file <- read.table(plantcare_path, header = FALSE, sep = '\t', quote="")
plantcare_data <- plantcare_classify(plantcare_file)
statistic_data2 <- plantcare_statistic2(plantcare_data)
```

plantcare_to_loc *plantcare_to_loc*

Description

Extract the location information of cis-element from Plantcare

Usage

```
plantcare_to_loc(plantcare_data)
```

Arguments

plantcare_data The result of plantcare_classify().

Value

data.frame

Author(s)

Shiqi Zhao

Examples

```
plantcare_path <- system.file("extdata", "plantCARE_output.tab", package = "BioVizSeq")
plantcare_file <- read.table(plantcare_path, header = FALSE, sep = '\t', quote="")
plantcare_data <- plantcare_classify(plantcare_file)
plantcare_loc <- plantcare_to_loc(plantcare_data)
```

smart_plot	<i>smart_plot</i>
------------	-------------------

Description

Visualization of domain in SMART result file

Usage

```
smart_plot(  
    fasta_file,  
    the_order = NULL,  
    domain_select = NULL,  
    shape = "RoundRect",  
    r = 0.3,  
    legend_size = 15,  
    domain_color = NULL  
)
```

Arguments

fasta_file	The path of protein fasta file.
the_order	The path of order file. A List of Gene ID , One ID Per Line.
domain_select	The domain ID which you want to align with.
shape	RoundRect or Rect.
r	The radius of rounded corners.
legend_size	The size of legend.
domain_color	The color set of domain.

Value

p

Author(s)

Shiqi Zhao

smart_to_loc	<i>smart_to_loc</i>
--------------	---------------------

Description

Extract the location information of domain from SMART result

Usage

```
smart_to_loc(input_file, do_pfam = TRUE)
```

Arguments

input_file	The path of protein fasta file.
do_pfam	Include the pfam domain or not.

Value

list

Author(s)

Shiqi Zhao

upload_fa_to_plantcare	<i>upload_fa_to_plantcare</i>
------------------------	-------------------------------

Description

Upload the promoter file to Plantcare database

Usage

```
upload_fa_to_plantcare(fasta_file, email)
```

Arguments

fasta_file	The path of promoter file.
email	e-mail address.

Details

Due to the file size limitation of plantcare on fasta, upload_fa_to_plantcare() first splits fasta file. Then uploads the splited fasta files to the plantcare database, and automatically returns the results to the email provided by the user.

```
upload_fa_to_plantcare("the path/test.fasta", "your e-mail address")
```

Value

plantcare_result

Author(s)

Shiqi Zhao

Index

aes, 5
alpha, 5

cdd_plot, 2
cdd_to_loc, 3
colour, 5

fastaleng, 4
fill, 5

geom_rrect, 4
get_motif_location, 6
gff_plot, 7
gff_to_loc, 8
group, 5

linetype, 5

meme_plot, 8
motif_plot, 10
motif_seq, 11

pfam_plot, 12
pfam_to_loc, 13
plantcare_classify, 13
plantcare_plot, 14
plantcare_statistic1, 15
plantcare_statistic2, 15
plantcare_to_loc, 16

smart_plot, 17
smart_to_loc, 18

upload_fa_to_plantcare, 18

xmax, 5
xmin, 5

ymax, 5
ymin, 5